

# **A Seedling Identification Guide**

**For Common Plants on Mt. Rainier and the North Cascades**

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# About this Project



## Project Inspiration

This project arose from my personal interests in plant propagation, plant identification, and scientific writing during my time as an undergraduate student in plant biology at the University of Washington. After a summer fieldwork job that involved seedling identification, I gained an appreciation for this task and its challenges, including the lack of resources available. Although there are numerous identification guides for mature plants, there are remarkably few for seedlings. Yet, researchers and restoration managers need seedling identification resources to help them track seed germination and seedling mortality rates. I sought to fill this knowledge gap by creating a seedling identification guide for thirty plant species commonly found on Mt. Rainier and the North Cascades in Washington. This guide was first used to support field research conducted by the Hille Ris Lambers Lab, a plant community ecology lab at the University of Washington. However, I hope that this guide will serve as a multipurpose resource for scientists, amateur botanists, seed enthusiasts, and anyone who wishes to learn more about the beginning of a plant's life.

# About this Project



## A Case for Seedling Identification

Despite the importance of early life stages to the survival, growth, and development of plants, little is known about the morphology of seedlings. Most plant identification guides focus on mature plants. Seedlings are harder to identify because they are very small, lack prominent features such as flowers and fruits, and have similar leaf shapes. Of the few seedling guides available, there is a heavy focus on crop and weed seedling identification. These resources are targeted toward farmers and invasive species management programs. Seedling guides for native plants are rare, but would be useful for plant ecologists and habitat restoration projects. One notable example is *A Guide to Seedling Identification for 25 Conifers of the Pacific Northwest* by forest ecologist Jerry Franklin.<sup>1</sup> Nevertheless, seedling identification resources for the Pacific Northwest could be greatly improved and expanded, especially for native species that are not trees.

The species in this guide represent a variety of growth forms of plants native to Mt. Rainier and the North Cascades, including trees, shrubs, flowers, and sedges. Most of these species were selected by the Hille Ris Lambers Lab for field research on the effects of climate change on seed germination and seedling survival. Identifying plants is a fundamental part of ecological fieldwork and can be achieved using strategies that range from flipping through plant photographs to following a dichotomous key to performing DNA analysis. This guide provides a morphological approach to seedling identification, which can be supplemented with environmental clues such as climate and elevation.

# Helpful Hints for Seedling Identification

## Basic Terminology<sup>2</sup>

*Cotyledons* are the first leaves of a seedling. Unlike true leaves, cotyledons emerge from the seed. Most flowers have one or two cotyledons, while conifers usually have multiple cotyledons.

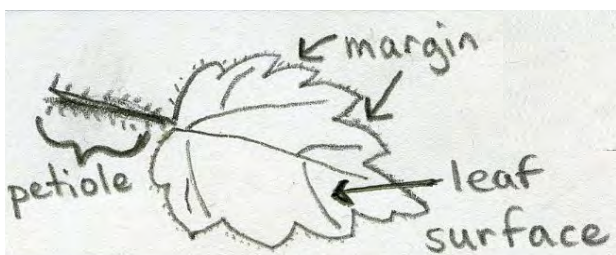
*True leaves* are leaves that emerge after the cotyledons. True leaves often look very different than cotyledons.

*Stomata* are pores that allow plants to breathe in carbon dioxide for photosynthesis and breathe out oxygen and water vapor. Most stomata are found on leaves. Conifer stomata are typically white and arranged on the needle in stripes. Different conifers have different numbers of stomatal stripes, which is useful for identification.

The *petiole* is the stem of a leaf, which attaches the leaf blade to the main stem. The petioles of cotyledons often lengthen as the cotyledons age, and may not be visible when the cotyledons first emerge. The petioles of true leaves are usually longer than the petioles of cotyledons. Some plants do not have petioles.

The *margin* of a leaf is the edge of the leaf, and may be smooth, spiny, jagged, etc.

The *surface* of a leaf is the area or blade of the leaf. The upper surface of the leaf faces the sky, and the lower surface faces the soil. Color and texture may differ between the upper and lower leaf surface.



# Helpful Hints for Seedling Identification

## Tools

Use a magnifying glass! It will help you identify details such as plant hairs and stomatal stripes that may be hard to see without magnification.

If you want to photograph seedlings, use a camera with a macro lens and tripod. A macro lens captures minute details that a regular lens will not. A tripod provides stability and minimizes image blur. If your seedlings are growing in a greenhouse, you may need to move your plants to better lighting to avoid yellow-tinted photographs.

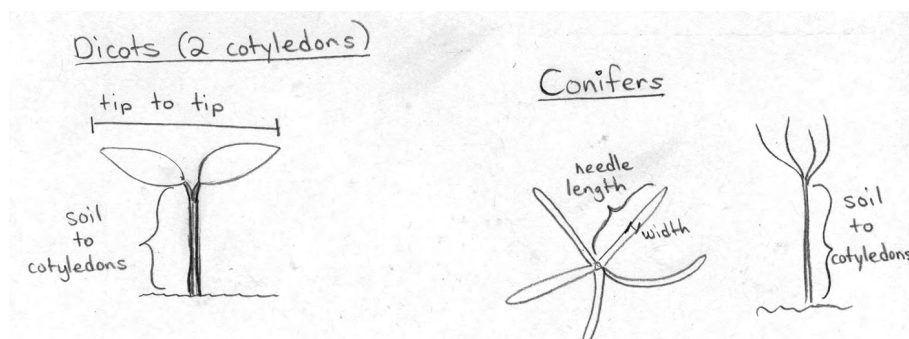
The penny in each photograph provides a visual size reference. Compare seedling dimensions to the diameter of the penny, 0.75 inches (19.05 mm).

## Tips

When identifying seedlings, first examine leaf shape and cotyledon number, and look for the presence of hairs. Other traits such as color can be informative but often vary between seedlings of the same species, especially when seedlings receive different amounts of light and water due to their location. Seed shells stuck to cotyledons may be useful if the seed can be identified.

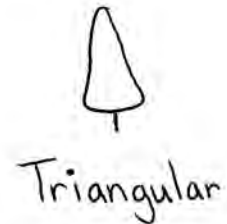
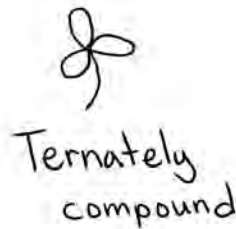
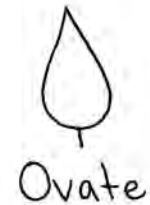
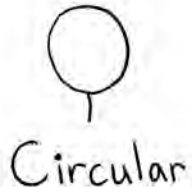
If you are struggling to identify a seedling, try identifying the adult plants in the area. The seedling could have emerged from a seed produced by a nearby plant.

Measurements in this guide serve as a rough size guideline for new seedlings. For plants with two cotyledons, the cotyledon measurement from “tip to tip” is the distance between the cotyledon tips. This leaf span is analogous to the human arm span. For conifers, the length and width of an individual needle is included instead of a “tip to tip” measurement. Secondly, the stem measurement from “soil to cotyledons” describes the distance from soil level to the bottom of the cotyledon leaf blades (not the bottom of the cotyledon petioles). Note that these numbers will increase as the seedling grows.



# Leaf Shape Terminology<sup>2</sup>

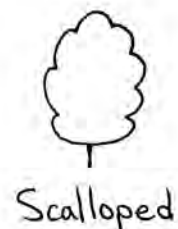
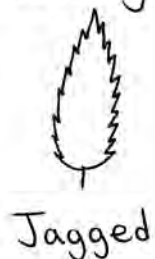
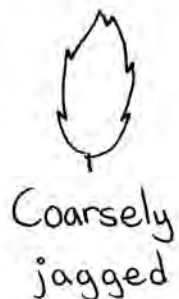
## Leaf Shapes



## Leaf Tips



## Leaf Margins





# Dichotomous Key

Similar to a choose-your-own adventure book, a dichotomous key allows you to reach a conclusion by guiding you through a series of binary decisions. In this case, the conclusion is the correct identification of a seedling and decisions are made based on cotyledon traits. This key only includes the 30 species in this guide, so the seedling that you are trying to identify may not be listed. To use this key, choose between a and b for a given number and follow the directions to go to the next number. If no number is mentioned, proceed to the following line. Continue until you have reached a species or genus. For example, you might suspect that you have a *Xerophyllum tenax* seedling, but want to check. You start by reading 1a and 1b, and choose 1a because the cotyledon is leafy. You follow the directions in 1a and go to 2. After reading 2a and 2b, you select 2a because the seedling has one cotyledon. 2a has no directions, so you go to the next line, 3. Since both 3a and 3b list plant names, this is your last choice. You decide that your hunch was correct because your seedling matches the description in 3a.

**1a.** Cotyledons leafy, not needlelike. Go to 2.

**1b.** Cotyledons needlelike. Go to 16, page 11.

**2a.** One cotyledon.

**3a.** *Xerophyllum tenax*: Thick, flat cotyledon with blunt or rounded tip. Seed often attached to cotyledon tip. No sheath at base.

**3b.** *Carex*: Thin, flat cotyledon that tapers to a pointed tip. Round sheath at base.

**2b.** Two cotyledons.

**4a.** *Rubus*: Hairs on cotyledon margin, but not on cotyledon surface. Cotyledons elliptic or oval. Cotyledons measure 5-10 mm from tip to tip soon after emergence.

**4b.** No hairs on cotyledon margin OR hairs on cotyledon surface and margin.

**5a.** Cotyledons measure less than 5 mm from tip to tip. Very tiny! Go to 6.

**5b.** Cotyledons measure 5 mm or more from tip to tip. Go to 9, page 10.

**6a.** *Tellima grandiflora* & *Tolmiea menziesii*: Cotyledons rounded but slightly triangular, and widest at the base of the leaf. Cotyledon tips are slightly notched. A vein line down the center of the cotyledon is often visible. The first true leaves look like miniature adult true leaves. The first true leaves are round with a scalloped margin and tiny white hairs.

**6b.** Cotyledon leaf tip round and not notched.

**7a.** *Anaphalis margaritacea*: Cotyledons circular or slightly elliptic, and 1-1.5 times as long as wide. Cotyledon petioles lengthen as the seedling ages, drawing the cotyledons apart. Cotyledons are hairless, but true leaves have soft, white hairs.

**7b.** Cotyledons elliptic or oval, not circular. Cotyledons at least 1.5 times as long as wide. 9

# Dichotomous Key

- 8a. Alnus:** Cotyledons elliptic or oval, and widest at the base or middle. Cotyledons usually 1.5-2 times as long as wide. Petioles lengthen as seedling ages. The stem is often reddish. True leaves have serrate, jagged, or lobed margins.
- 8b. Vaccinium:** Cotyledons elliptic or oval, and twice as long as wide. Cotyledon width is relatively constant from leaf base to tip. There is no or little space between the cotyledons since petiole length is insignificant. The stem is often reddish. The first true leaves are the size of cotyledons, but have slightly serrate or jagged margins.
- 9a. Lupine:** Cotyledons thick, fleshy, and bean shaped. The stem is often reddish or purplish, while the cotyledons are dark green. Cotyledons are large, about 15-25 mm from tip to tip.
- 9b.** Cotyledons thin or not bean shaped.
- 10a. Sambucus:** Cotyledons triangular. Cotyledon tip blunt or slightly pointed. Cotyledons measure 10-20 mm from tip to tip soon after emergence, but enlarge greatly as the seedling develops.
- 10b.** Cotyledons not triangular.
- 11a. Acer:** Cotyledons oblong, at least 3 times as long as wide, and 35-45 mm from tip to tip soon after emergence. Cotyledons tough, but commonly creased and crinkled with fold lines.  
Waxy stem.
- 11b.** Cotyledons soft, under 30 mm from tip to tip, or not oblong.
- 12a. Anemone occidentalis:** Cotyledons and first true leaf have separate stems. The base of the cotyledon does not taper to a petiole-like structure. Cotyledons elliptic or oval. True leaves dissected.
- 12b.** Cotyledons and first true leaves found on same stem.
- 13a. Berberis:** The base of the cotyledon does not taper to a petiole-like structure.  
Cotyledons elliptic or oval. Stem 10 mm or longer. Spiny true leaf margin.
- 13b.** The base of the cotyledon tapers to a petiole-like structure that is relatively long and thick, being at least 1/3 of the width and 1/4 of the length of cotyledon leaves.  
Seedlings germinate and grow quickly.
- 14a. Eriogon peregrinus:** Cotyledons as thin as paper. Surface looks and feels smooth.  
Cotyledons elliptic or oval, and widest at the middle. Tips are blunt or rounded.  
Surface of true leaves is hairless, but margins and petioles may have small hairs.
- 14b.** Cotyledons slightly thicker than paper. Stem base is often reddish or pinkish.
- 15a. Arnica latifolia:** Cotyledons spatulate or oblong. Cotyledon apex rounded, never notched. True leaves covered in thin white hairs.
- 15b. Eriophyllum lanatum:** Cotyledons various shapes, often spatulate or elliptic.  
Cotyledon apex notched or rounded. Cotyledon surface has visual texture, but feels smooth. True leaves covered in thick, long, white hairs, hence the common name woolly sunflower.

# Dichotomous Key

## 16a. Needle tip pointed.

**17a.** Double white stomatal stripes on upper cotyledon surface, but no stomatal stripes on lower cotyledon surface. Stomatal stripes may be faint on newly emerged cotyledons, but should be visible with a magnifying glass. 4-8 cotyledons.

**18a.** *Pseudotsuga menziesii*: Cotyledons 5-8, most commonly 6-7. Cotyledons wider than *Picea*. 0.7 to 1.2 mm wide. Seedlings taller than *Picea*. 15 to 35 mm tall. Cotyledon length usually greater than *Picea*. 10-25 mm long. Cotyledons cupped up. Stem often deep red.

**18b.** *Picea*: Cotyledons narrow, less than 0.7 mm wide. Seedlings generally shorter than *Pseudotsuga menziesii*. Less than 20 mm tall soon after emergence. Cotyledon length usually shorter than *Pseudotsuga menziesii*. Less than 13 mm long.

**19a.** *Picea sitchensis*: Cotyledons 4-6, most commonly 5-6. Seedling height and cotyledon width and length are similar to *Picea engelmanni*.

**19b.** *Picea engelmanni*: Cotyledons 4-8, most commonly 5-7. Seedling height and cotyledon width and length are similar to *Picea sitchensis*.

**17b.** Double stomatal stripe on upper cotyledon surface not visible with a magnifying glass OR cotyledon count fewer than 4 or greater than 8.

**20.** *Pinus*: Cotyledons narrow, no more than 0.5 mm wide. Cotyledons flat on newly emerged seedlings, but become more triangular with age. Cotyledons usually cupped upwards.

**20a.** *Pinus contorta*: Cotyledons 4-6, most commonly 5. Cotyledons typically shorter than *Pinus ponderosa*, up to 30 mm long.

**20b.** *Pinus ponderosa*: Cotyledons 8-12, most commonly 10. Cotyledons arranged like a messy hairdo, pointed upwards in multiple directions. Cotyledons typically longer than *Pinus contorta*, up to 60 mm long.

## 16b. Needle tip rounded or blunt.

**21.** *Abies*: Cotyledons 4-7, flat, and wide. Cotyledons usually 1-2 mm in width. Double white stomatal stripes present on upper cotyledon surface, but no stomatal stripes on lower cotyledon surface. Stomatal stripes may be faint on newly emerged cotyledons, but should be visible with a magnifying glass. Stem often reddish.

**21a.** *Abies grandis*: Usually taller than *Abies lasiocarpa*. 20-50 mm tall. Needles often longer than *Abies lasiocarpa*. 15-35 mm long.

**21b.** *Abies lasiocarpa*: Usually shorter than *Abies grandis*. 10-25 mm tall. Needles often shorter than *Abies grandis*. 10-25 mm long.

Conifer cotyledon measurements included in this key were obtained from *A Guide to Seedling Identification for 25 Conifers of the Pacific Northwest* by Jerry Franklin.<sup>1</sup>

# Conifers

Side by side comparison is useful when identifying multiple species of conifer seedlings. For conifer seedlings of the same age, pine (*Pinus*) seedlings are usually bigger than true fir (*Abies*) seedlings, which are bigger than spruce (*Picea*) seedlings. The needle and stem lengths of conifer seedlings provided in this guide were referenced from *A Guide to Seedling Identification for 25 Conifers of the Pacific Northwest* by Jerry Franklin.<sup>1</sup>

Height and Needle Length	Shortest	Medium	Longest
Species	Spruces ( <i>Picea</i> )	True firs ( <i>Abies</i> )	Pines ( <i>Pinus</i> )

Note that white stomatal stripes on conifer needles may be difficult to see soon after cotyledons emerge but more visible as cotyledons age.



# Grand fir (*Abies grandis*)

## Cotyledons

**Number:** 5-7 (most commonly 5-6)

**Color:** Dark green

**Upper needle surface:** Two wide white stomatal stripes

**Lower needle surface:** Plain green, no stomatal stripes

**Shape:** Round or blunt tip, flat and wide needle

**Length:** 15-35 mm long, 1.0-2.2 mm wide<sup>1</sup>

**Angle:** Needles horizontal or slightly curved

## Young stem

**Color:** Usually red, but may be reddish green

**Texture:** Waxy, no hairs

**Length:** 20-50 mm from soil to cotyledons<sup>1</sup>



**Left:** Seedling with seed shell covering cotyledons. **Right:** Seedling with cotyledons only. Note double white stripes on each needle, which are stomata and develop with age.



**Left:** Note red stem. **Middle and right:** Seedlings with cotyledons and true needles. True needles develop double white stomatal stripes and a darker green color with age.

# Subalpine fir (*Abies lasiocarpa*)

## Cotyledons

**Number:** 4-7 (most commonly 5-6)

**Color:** Dark green

**Upper needle surface:** Two wide white stomatal stripes

**Lower needle surface:** Plain green, no stomatal stripes

**Shape:** Round or blunt tip, flat and wide needle

**Length:** 10-25 mm long, 1.0-2.2 mm wide<sup>1</sup>

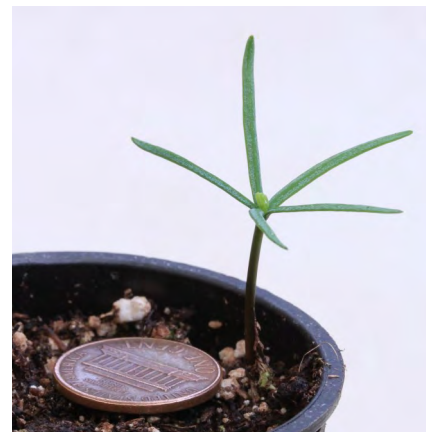
**Angle:** Needles pointed upward, often forming a cup

## Young stem

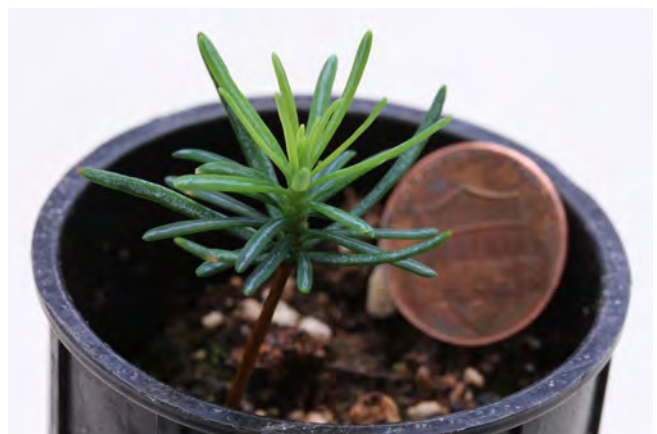
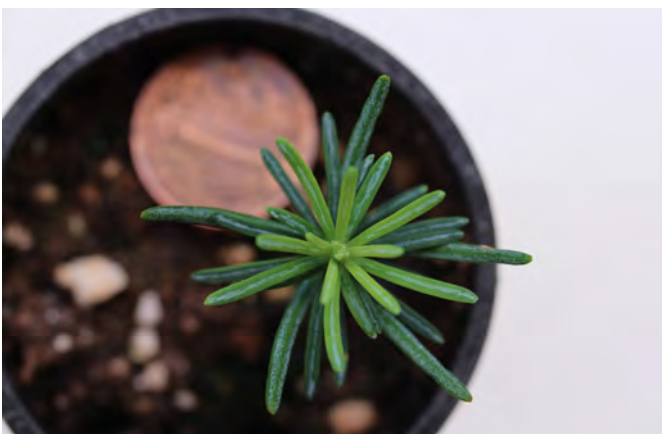
**Color:** Usually red, but may be reddish green or pink

**Texture:** Waxy, no hairs

**Length:** 10-25 mm from soil to cotyledons<sup>1</sup>



Seedlings with cotyledons only. **Left:** Seedling with seed shell attached to cotyledon tips. **Left and right:** Note variation in stem color.



Seedlings with cotyledons and true needles. True needles develop double white stomatal stripes and a darker green color with age. Note that true needles are denser than *Abies grandis* seedlings.

# Engelmann spruce (*Picea engelmanni*)

## Cotyledons

**Number:** 4-8 (most commonly 5-7)

**Color:** Medium to dark green

**Upper needle surface:** Two white stomatal stripes, sometimes difficult to see

**Lower needle surface:** Plain green, no stomatal stripes

**Shape:** Pointed tip, flat needle that increases in thickness with age

**Length:** 6-13 mm long, 0.3-0.6 mm wide<sup>1</sup>

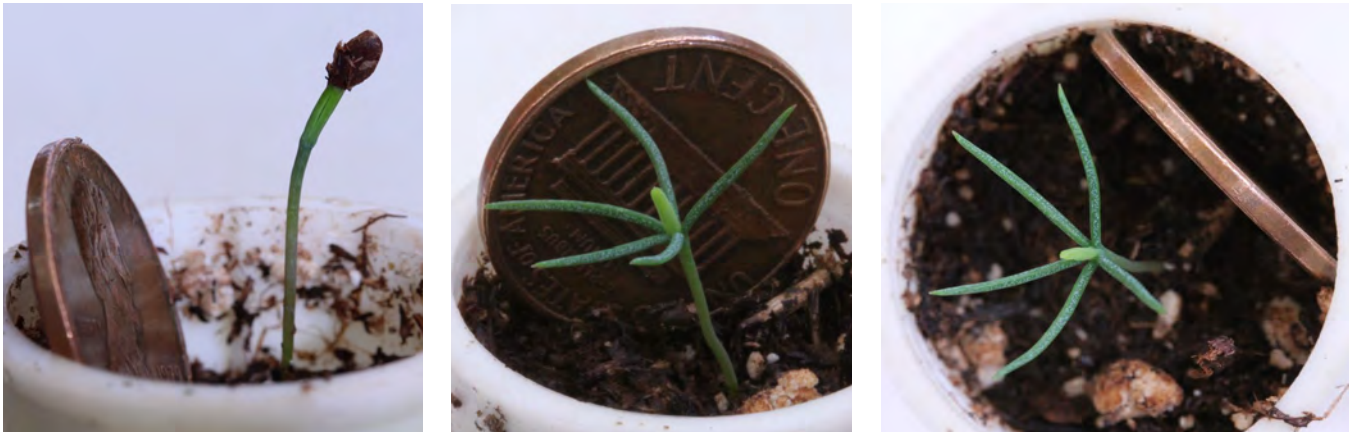
**Angle:** Needles usually cupped upward, but may point in multiple directions

## Young stem

**Color:** Greenish white, green, greenish red, or red

**Texture:** Waxy, no hairs

**Length:** 8-18 mm from soil to cotyledons<sup>1</sup>



**Left:** Seedling with seed shell attached to cotyledon tips. **Middle and right:** Seedlings with cotyledons only. Note *Picea* needles are much shorter than *Abies* and *Pinus* needles in this guide.



**Left:** Seedling with cotyledons only. Note pointed *Picea* needle tips, as opposed to rounded *Abies* needle tips. **Middle and right:** Seedlings with cotyledons and true needles.

# Sitka spruce (*Picea sitchensis*)

## Cotyledons

**Number:** 4-6 (most commonly 5-6)

**Color:** Dark green

**Upper needle surface:** Two white stomatal stripes, sometimes difficult to see

**Lower needle surface:** Plain green, no stomatal stripes

**Shape:** Pointed tip, flat needle that increases in thickness with age

**Length:** 6-11 mm long, 0.3-0.6 mm wide<sup>1</sup>

**Angle:** Needles pointed up or in multiple directions

## Young stem

**Color:** Green or greenish red

**Texture:** Waxy, no hairs

**Length:** 7-15 mm from soil to cotyledons<sup>1</sup>



**Left:** Seedling with seed shell attached to cotyledon tips. **Middle and right:** Seedling with cotyledons only. Note pointed *Picea* needle tips, as opposed to rounded *Abies* needle tips.



Seedlings with cotyledons and true needles. True needles may have bluish tint. See how *Picea* needles are narrower than *Abies* needles.



# Lodgepole pine (*Pinus contorta*)

## Cotyledons

**Number:** 4-6 (most commonly 5)

**Color:** Medium green

**Upper needle surface:** Plain green

**Lower needle surface:** Plain green

**Shape:** Pointed tip, flat needle that becomes triangular with age

**Length:** 16-30 mm long, 0.4-0.5 mm wide<sup>1</sup>

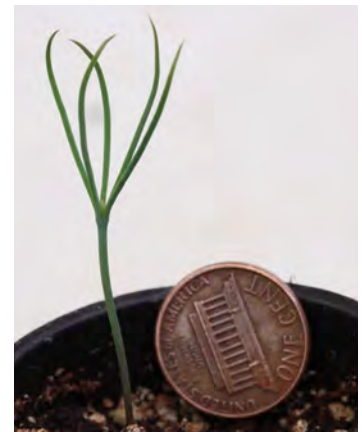
**Angle:** Needles strongly pointed or cupped up, forming a V or U when viewed from the side

## Young stem

**Color:** Green, reddish green, or reddish purple

**Texture:** Waxy, no hairs

**Length:** 10-25 mm from soil to cotyledons<sup>1</sup>



**Left:** Seedling with seed shell attached to cotyledon tips. **Middle and right:** Seedlings with cotyledons only. Note thinner and longer needles than *Abies*.



**Left:** Seedling with cotyledons surrounding central emerging true needles. **Middle and right:** Seedlings with cotyledons and true needles. Needles shorter than *Pinus ponderosa*, shown in reference to penny and edge of cone-tainer.

# Ponderosa pine (*Pinus ponderosa*)

## Cotyledons

**Number:** Many, 8-12 (most commonly 10)

**Color:** Dark green

**Upper needle surface:** Plain green

**Lower needle surface:** Plain green

**Shape:** Pointed tip, flat needle that increases in thickness and becomes triangular with age

**Length:** Very long needles, 25-60 mm long, 0.2-0.5 mm wide<sup>1</sup>

**Angle:** Needles arranged like a messy hairdo and strongly cupped upward

## Young stem

**Color:** Green or reddish green

**Texture:** Waxy, no hairs

**Length:** 15-40 mm from soil to cotyledons<sup>1</sup>



**Left:** Seedling with seed shell attached to cotyledon tips. **Middle and right:** Seedlings with cotyledons. Note that the number of cotyledons is greater than *Pinus contorta*.



**Left:** Seedling with cotyledons surrounding central emerging true leaves. **Middle and right:** Seedlings with cotyledons and true needles. Needles longer than *Pinus contorta*, shown in reference to the penny and edge of cone-tainer.

# Douglas fir (*Pseudotsuga menziesii*)

## Cotyledons

**Number:** 5-8 (most commonly 6-7)

**Color:** Medium green

**Upper needle surface:** Two faint white stomatal stripes

**Lower needle surface:** Plain green

**Shape:** Pointed tip, flat needle that increases in thickness with age

**Length:** 10-25 mm long, 0.7-1.2 mm wide<sup>1</sup>

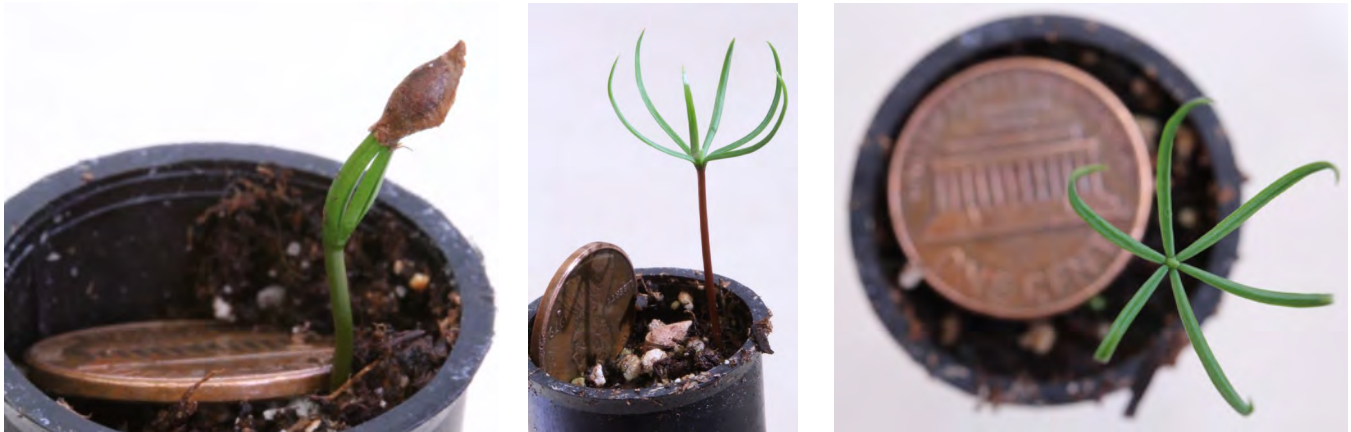
**Angle:** Needles cupped up into a bowl shape, tips often curve inward when viewed from above

## Young stem

**Color:** Usually deep red, but may also be greenish red or whitish green

**Texture:** Waxy, no hairs

**Length:** 15-35 mm from soil to cotyledons<sup>1</sup>



**Left:** Seedling with seed shell attached to cotyledon tips. **Middle:** Note red stem. **Middle and right:** Cotyledons cup upward much more than *Abies*. **Right:** Unlike *Abies*, cotyledon tips curve inward.



Seedlings with cotyledons and true needles. **Middle:** Easy to confuse with *Abies* seedlings, which also have red stems. **Left and right:** Note pointed needle tips, as opposed to rounded *Abies* needle tips.

## Other Trees & Shrubs

Whether or not a seedling will develop into a tree may not be obvious. Some trees, such as alders, invest more energy into leaf growth than stem growth when they are very young. Other trees, such as maples, grow seedlings with rapidly elongating stems. A seedling may not look like a miniature version of the adult plant that it will become.



These seedlings showed little resemblance to adult maple trees when they only bore cotyledons, visible in this photograph as the pair of long, narrow green leaves at the base of each stem. However, the identity of the seedlings became more apparent after the emergence of their true leaves, which are similar to adult maple leaves in shape.

# Vine maple (*Acer circinatum*)

## Two cotyledons

**Shape:** Oblong, width slightly greater at leaf apex, round or blunt tip, smooth margin

**Color:** Medium green

**Texture:** Hairless, tough leaves, may have fold lines due to struggle to emerge from seed

**Length:** 35-45 mm from tip to tip

## Young stem

**Color:** Varies, may be light red, reddish purplish green, greenish red, or light green

**Texture:** Waxy

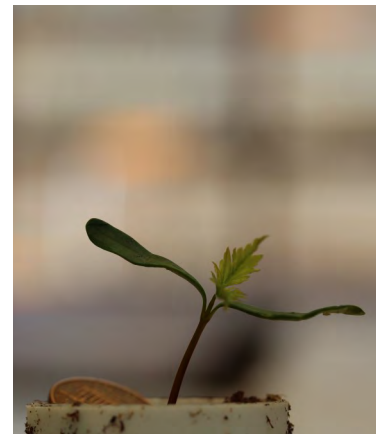
**Length:** 15-25 mm from soil to cotyledons

## First true leaves

**Shape:** Resemble mature maple leaves, narrow when young

**Color:** Bright yellowish green, light green, greenish purple, or purplish red

**Texture:** Hairless



**Left:** Seedling emerging from seed. **Middle and right:** Seedlings with cotyledons and young true leaves.



**Left:** Seedling with cotyledons and first pair of true leaves. **Middle:** Close up of first true leaf. **Right:** Later true leaves have deeper lobes. **Middle and right:** Note how true leaves widen with age. Also observe variation in leaf color.

# Rocky Mountain maple (*Acer glabrum*)

## Two cotyledons

**Shape:** Oblong, width slightly greatest at leaf midpoint, round or blunt tip, smooth margin

**Color:** Medium green

**Texture:** Hairless, tough leaves, may have fold lines due to struggle to emerge from seed

**Length:** 35-45 mm from tip to tip

## Young stem

**Color:** Light green, greenish red, or red

**Texture:** Waxy

**Length:** 15-20 mm from soil to cotyledons, usually shorter than *Acer circinatum*

## First true leaves

**Shape:** Resemble elongated mature maple leaves, narrow when young

**Color:** Light green, may have tinges of red and yellow

**Texture:** Hairless



Seedlings with cotyledons only. Observe crinkled fold lines on cotyledons.



**Left:** Seedling with cotyledons and first true leaves. **Middle and right:** Older seedlings with true leaves. **Right:** Note first pair of true leaves is longer than later true leaves. See how leaf lobes are shallower than *Acer circinatum*.

# Red alder (*Alnus rubra*)

## Two cotyledons

**Shape:** Elliptic or oval, smooth margin, seed shell often remains attached to young cotyledons

**Color:** Medium green

**Texture:** Hairless

**Length:** Very small, less than 5 mm from tip to tip

## Young stem

**Color:** Whitish green, reddish green, or reddish purple

**Texture:** Hairless

**Length:** Less than 5 mm from soil to cotyledons

## First true leaves

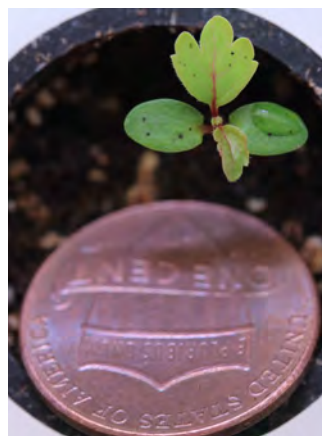
**Shape:** Circular to ovate, lobing at leaf apex, also lobing along sides of margin in older seedlings

**Color:** Medium green or yellowish green, middle vein sometimes reddish

**Texture:** Small, short, sparse white hairs on leaf stem and margin



**Left and middle:** Seedlings with cotyledons only. **Right:** Seedling with cotyledons and first true leaves. Note red stem.



Seedlings with cotyledons and true leaves. Note that tip of leaf margin lobes are more rounded than *Alnus viridis*.

**Right:** See how base of leaf margin is smooth even on older true leaves, unlike *Alnus viridis*.

# Green alder (*Alnus viridis*)

## Two cotyledons

**Shape:** Elliptic or oval, smooth margin, seed shell often remains attached to young cotyledons

**Color:** Medium-dark green

**Texture:** Hairless

**Length:** Very small, less than 5 mm from tip to tip

## Young stem

**Color:** White, whitish green, or red

**Texture:** Hairless

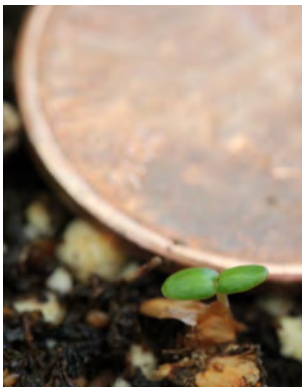
**Length:** Less than 5 mm from soil to cotyledons

## First true leaves

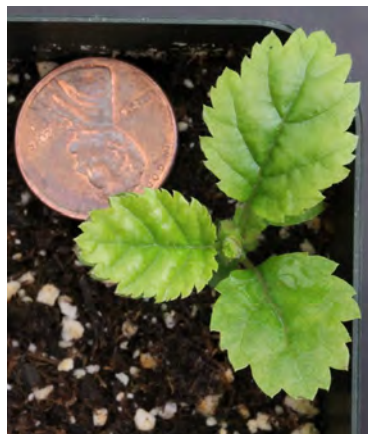
**Shape:** Circular to ovate, jagged margin at leaf apex and later all along margin

**Color:** Medium green or yellowish green

**Texture:** Small, short, sparse white hairs on leaf stem and margin



**Left:** Seedling with cotyledons and seed shell remnant attached to stem. **Middle and right:** Seedlings with cotyledons and first true leaves. Observe variation in jagged margin in first true leaves.



**Left:** Seedling with first true leaves. **Middle and right:** Older seedlings. Note jagged leaf margin forms sharper points than *Alnus rubra* margin. See how leaf margin is jagged almost to the petiole junction on older true leaves, unlike *Alnus rubra*.



# Tall Oregon grape (*Berberis aquifolium*)

## Two cotyledons

**Shape:** Elliptic or oval, smooth margin

**Color:** Medium green, may be yellowish or reddish

**Texture:** Hairless

**Length:** 10-20 mm from tip to tip

## Young stem

**Color:** Whitish green, green, greenish red, or red

**Texture:** Hairless

**Length:** 10-15 mm from soil to cotyledons

## First true leaves

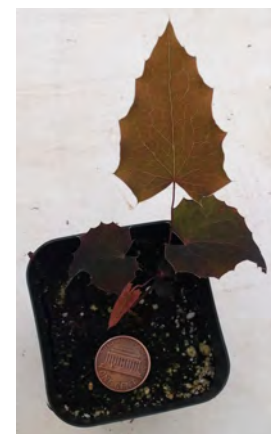
**Shape:** Cordate, spiny margin with spines at leaf apex pointing in same direction as leaf tip and spines below apex pointing approximately perpendicular to leaf margin

**Color:** Greenish red, reddish purple, or purplish green, lighter veins, white lower leaf surface

**Texture:** Leaves become leathery with age



**Left and middle:** Seedlings with cotyledons only. **Middle:** Note seed shell still attached to seedling. **Right:** First true leaf.



**Left and middle:** Seedlings with cotyledons and first true leaves. See how the V that makes the heart shape at the base of leaf is deeper than *Berberis nervosa*. **Right:** Older seedling with true leaves.

# Dwarf Oregon grape (*Berberis nervosa*)

## Two cotyledons

**Shape:** Elliptic or oval, smooth margin

**Color:** Green, reddish green, or reddish purplish green

**Texture:** Hairless

**Length:** 10-20 mm from tip to tip

## Young stem

**Color:** Reddish purple, reddish green, or green

**Texture:** Hairless

**Length:** 10-15 mm from soil to cotyledons

## First true leaves

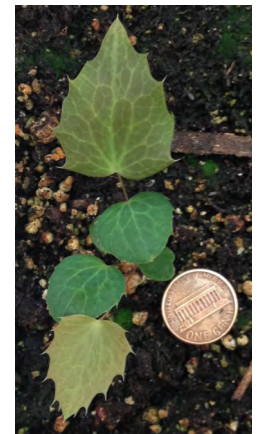
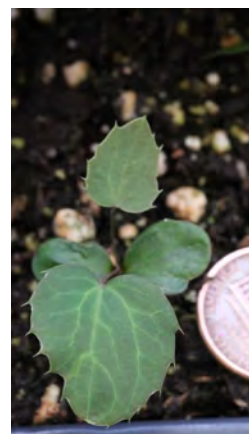
**Shape:** Cordate, spiny margin with spines pointing in same direction as leaf tip

**Color:** Green, greenish red, purplish green, or red, lighter veins, white leaf lower surface

**Texture:** Leaves become leathery with age



Seedlings with cotyledons and emerging pair of true leaves.



Seedlings with cotyledons and true leaves. Note spines on leaf margin point in the same general direction as the leaf tip, unlike *Berberis aquifolium*, which has spines pointing away from the leaf tip in the bottom half of the leaf.

# Thimbleberry (*Rubus parviflorus*)

## Two cotyledons

**Shape:** Elliptic or oval, smooth margin

**Color:** Medium green upper leaf surface, slightly lighter green lower leaf surface

**Texture:** Tiny hairs stick out horizontally from leaf margin, more visible with magnifying glass

**Length:** 5-10 mm from tip to tip

## Young stem

**Color:** Green or greenish white, sometimes with a pink tinge

**Texture:** Hairs stick straight out along stem

**Length:** Less than 5 mm from soil to cotyledons

## First true leaves

**Shape:** Triangular or cordate, coarsely jagged margin at leaf apex and sides, smooth margin at leaf base

**Color:** Medium green leaf surface, light green or purplish veins

**Texture:** White hairs on upper and lower leaf surface, leaf margins, and petiole



**Left and middle:** Seedlings with cotyledons only. **Left:** Note hairs on leaf margin. **Middle:** Note hairs on stem. **Right:** Seedling with cotyledons and first true leaves.



**Left:** One of the first true leaves. **Middle and right:** Older plants with true leaves. **Right:** Mature true leaf.

# Trailing blackberry (*Rubus ursinus*)

## Two cotyledons

**Shape:** Elliptic or oval, smooth margin

**Color:** Medium green

**Texture:** White hairs on leaf margin, more visible with magnifying glass

**Length:** 5-10 mm from tip to tip

## Young stem

**Color:** Light green or white

**Texture:** White hairs, tiny dots of red fluid secreted at hair tips in older seedlings

**Length:** Less than 5 mm from soil to cotyledons

## First true leaves

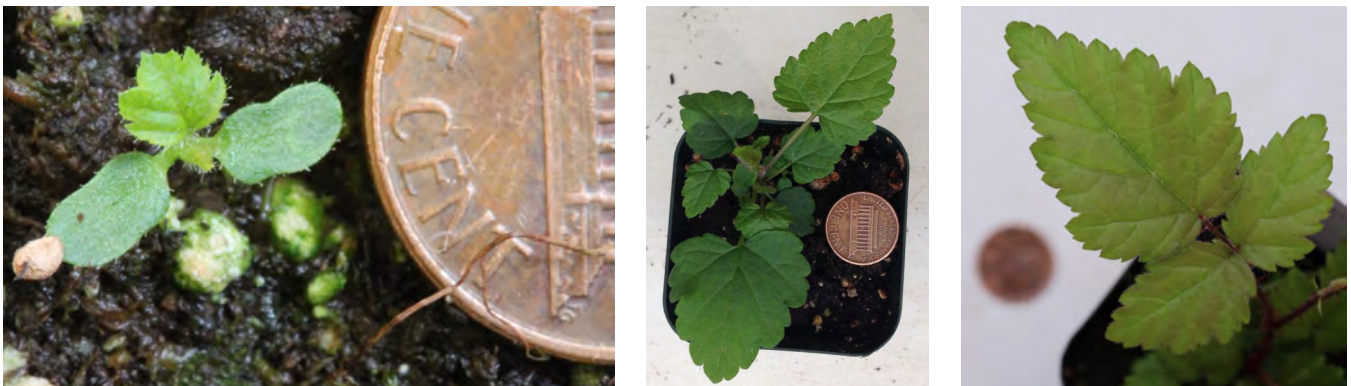
**Shape:** Circular or cordate, coarsely jagged margin at leaf apex and sides, smooth leaf base margin

**Color:** Medium green

**Texture:** Dense white hairs on petiole, dark pink thorns on petiole in older seedlings, also hairs on upper and lower leaf surface and margins



**Left and middle:** Seedlings with cotyledons only. **Right:** Seedling with cotyledons and first true leaf. Note hairs along margin of cotyledons and true leaves.



**Left:** Seedling with cotyledons and first true leaf. **Middle and right:** Older plants with true leaves. **Right:** Mature true leaf.

# Blue elderberry (*Sambucus cerulea*)

## Two cotyledons

**Shape:** Triangular, blunt or slightly pointed tip, smooth margin

**Color:** Light to medium green

**Texture:** Hairless

**Length:** 10-20 mm from tip to tip

## Young stem

**Color:** Light or whitish green

**Texture:** Hairless

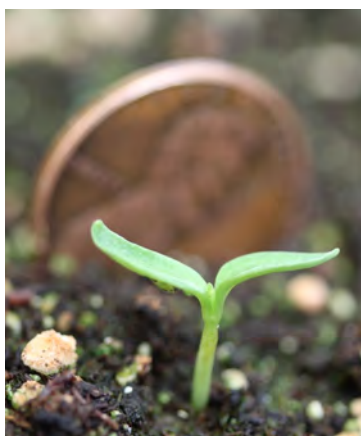
**Length:** 5-10 mm from soil to cotyledons

## First true leaves

**Shape:** Ovate, coarsely jagged margin, pointed or slightly rounded tip

**Color:** Light to medium green

**Texture:** Small white hairs of medium density on petiole, sparse hairs on lower leaf surface



Seedlings with cotyledons only. **Left:** Seed still attached to cotyledons. **Right:** Cotyledons elongate as seedling develops.



**Left:** Seedling with cotyledons and first pair of true leaves. See how leaf base of first pair of true leaves is straight, lacking the inward notch found in *Sambucus racemosa*. **Middle:** Older seedling. **Right:** True leaf of older seedling.

# Red elderberry (*Sambucus racemosa*)

## Two cotyledons

**Shape:** Triangular, blunt or slightly pointed tip, smooth margin

**Color:** Light to medium green

**Texture:** Hairless

**Length:** 10-20 mm from tip to tip

## Young stem

**Color:** Light or whitish green

**Texture:** Hairless

**Length:** 5-10 mm from soil to cotyledons

## First true leaves

**Shape:** Cordate, coarsely jagged margin, pointed or slightly rounded tip

**Color:** Light to medium green

**Texture:** White hairs on upper leaf surface, margins, and petiole



**Left and middle:** Seedlings with cotyledons only. **Right:** Seedling with cotyledons and first pair of true leaves. See how first true leaves are more rounded than *Sambucus cerulea* and leaf base notches inward.



Older seedlings. Note how second pair of true leaves differs in shape from first pair of true leaves.

# Cascade blueberry (*Vaccinium deliciosum*)

## Two cotyledons

**Shape:** Elliptic or oval, twice as long as wide, smooth margin, no or little space between cotyledons

**Color:** Green or reddish green

**Texture:** Hairless

**Length:** Extremely small; less than 5 mm from tip to tip

## Young stem

**Color:** Light green, reddish green, red, or reddish purple

**Texture:** Hairless

**Length:** Less than 5 mm from soil to cotyledons

## First true leaves

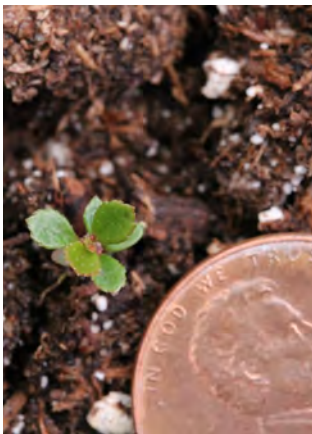
**Shape:** Obovate or oval, rounded leaf apex with blunt or slightly pointed tip, petiole absent or very short, margin slightly jagged at leaf apex, also slightly jagged on leaf sides in older seedlings

**Color:** Medium green, may be yellowish or reddish, sometimes with a red leaf margin

**Texture:** White hairs on upper leaf surface, margins, and petiole



Seedlings with cotyledons only. Note variation in color and leaf shape in individual seedlings of the same species.



Seedlings with cotyledons and true leaves. Note variation in shape of true leaves on the same plant.

# Red huckleberry (*Vaccinium parviflorum*)

## Two cotyledons

**Shape:** Elliptic or oval, twice as long as wide, smooth margin, no or little space between cotyledons

**Color:** Green

**Texture:** Hairless

**Length:** Extremely small; less than 5 mm from tip to tip

## Young stem

**Color:** Light green

**Texture:** Hairless

**Length:** Less than 5 mm from soil to cotyledons

## First true leaves

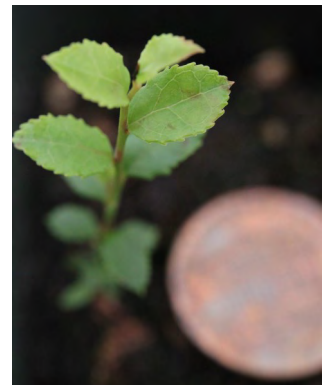
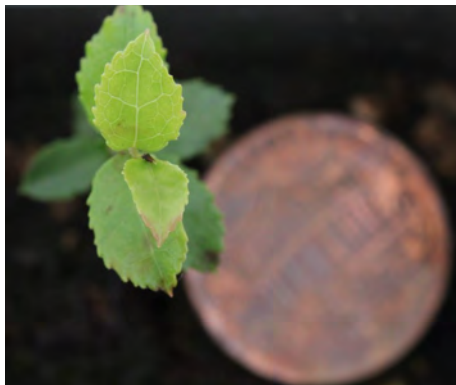
**Shape:** Ovate, elliptic, or obovate, slightly jagged margin, pointed tip, petiole absent or very short

**Color:** Light to medium green, may be yellowish or reddish, sometimes with a red leaf margin

**Texture:** White hairs on upper leaf surface, margins, and petiole



**Left:** Seedling with cotyledons and seed shell attached. **Right:** Seedling with cotyledons and first true leaves.



**Left:** Seedling with cotyledons and first true leaves. **Middle and right:** Older seedlings. Note that true leaf shape is usually ovate or elliptic for *Vaccinium parviflorum*, with the widest part of the leaf below the midpoint, but obovate for *Vaccinium deliciosum*, with the widest part of the leaf at or above the midpoint.



# Flowers

Flowers typically germinate and grow more rapidly than trees and shrubs. This is particularly true for flowers that are native to high elevation habitats such as subalpine meadows, because the growing season is short and provides a limited window for bloom time.

Although numerous resources focus on identifying flowers in bloom, the photographs in this section illustrate that there are many stages of life besides the flower itself. The challenge in identifying flowers as seedlings lies in training the eye to recognize subtle differences in leaf traits between species, particularly leaf shape. Traits will vary slightly between individual plants of the same species, so examining more than one plant of the same species is recommended.



# Pearly everlasting (*Anaphalis margaritacea*)

## Two cotyledons

**Shape:** Circular or slightly elliptic, 1 to 1.5 times as long as wide, smooth margin

**Color:** Medium green

**Texture:** Hairless

**Length:** Less than 5 mm from tip to tip

## Young stem

**Color:** Light green

**Texture:** Hairless when young, fuzzy white stem develops after first true leaves

**Length:** Less than 5 mm from soil to cotyledons, stem not easily visible when seedling is small

## First true leaves

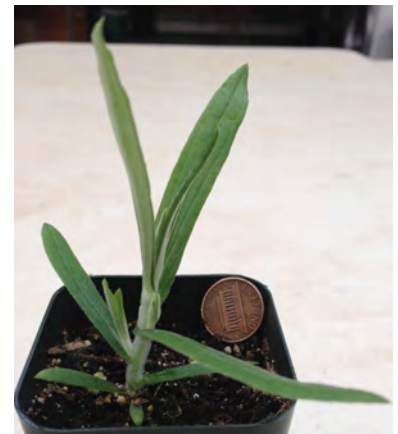
**Shape:** Elliptic, pointed or rounded tip, no petiole, thick and protruding vein on lower leaf surface

**Color:** Medium green, may have thin purplish vein lines on upper leaf surface

**Texture:** Soft, white hairs on upper and lower leaf surface



**Left and middle:** Seedlings with cotyledons. **Right:** Seedling with cotyledons and first true leaves. See how cotyledons are more circular than *Eriophyllum lanatum* even though first true leaves are similar. Note fuzzy hairs on true leaves.



Older plants. **Left:** First true leaves are elliptic. **Middle and right:** In contrast, older true leaves are linear.

# Western pasqueflower (*Anemone occidentalis*)

## Two cotyledons

**Shape:** Elliptic or oval, smooth margin

**Color:** Light to medium green

**Texture:** Hairless

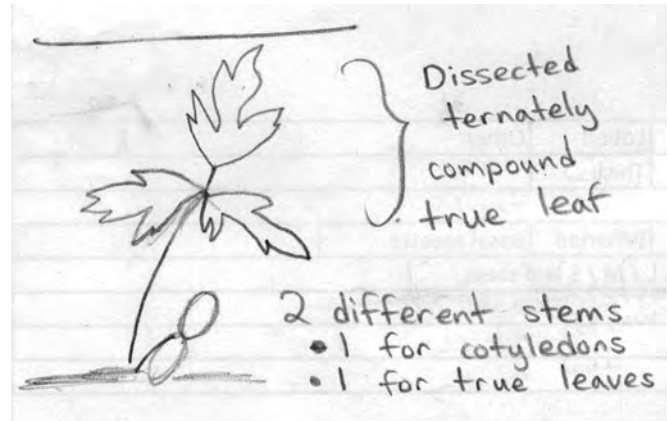
**Length:** 10-15 mm from tip to tip

## Young stem

**Color:** Light green, whitish or pinkish at base

**Texture:** Hairless

**Length:** 5-10 mm from soil to cotyledons



## First true leaves

**Shape:** Dissected, ternately compound (three leaflets)

**Color:** Lighter green on lower leaf surface, thin green branching vein lines

**Texture:** Hairless



**Left:** Seedling with cotyledons only. **Right:** Seedling with cotyledons and first true leaf. See how cotyledon and first true leaf stems are attached near soil level.



The first true leaf is formed on a second stem that is separate from the stem supporting the cotyledons. This is unusual.

# Broadleaf arnica (*Arnica latifolia*)

## Two cotyledons

**Shape:** Spatulate or oblong, smooth margin

**Color:** Medium green

**Texture:** Hairless

**Length:** 10-15 mm from tip to tip

## Young stem

**Color:** Whitish green or red

**Texture:** Hairless, but hairs along true leaf petioles

**Length:** 5-10 mm from soil to cotyledons

## First true leaves

**Shape:** Triangular, slightly jagged margin is more pronounced on older true leaves

**Color:** Similar to cotyledons, but prominent vein lines

**Texture:** White hairs on upper and lower leaf surface, leaf margin, and petiole



**Left:** Seedling with cotyledons only. **Middle:** Observe hairs on bud of leaf emerging between the cotyledons. **Right:** Seedling with cotyledons and pair of first true leaves. Note hairs and long petioles on true leaves.



Older plants. See how plant forms a rosette first rather than growing vertically.

# Mountain daisy (*Erigeron peregrinus*)

## Two cotyledons

**Shape:** Elliptic or oval, blunt or rounded tip, leaf as thin as paper, smooth margin

**Color:** Light to medium green, usually lighter than *Eriophyllum lanatum*

**Texture:** Surface looks and feels smooth, hairless

**Length:** 5-10 mm from tip to tip

## Young stem

**Color:** Light green

**Texture:** Hairless

**Length:** 5-10 mm from soil to cotyledons

## First true leaves

**Shape:** Elliptic or oval, curved or curled surface, smooth margin, long petiole, blunt or rounded tip

**Color:** Medium green

**Texture:** White hairs on leaf margin and petiole



**Left:** Seedlings with cotyledons and emerging true leaves. Note that cotyledons are rounder than *Arnica latifolia* cotyledons and smoother than *Eriophyllum lanatum* cotyledons. **Middle and right:** Seedlings with cotyledons and first true leaf.



Older plants. See how plant forms a rosette first rather than growing vertically.

# Common woolly sunflower (*Eriophyllum lanatum*)

## Two cotyledons

**Shape:** Spatulate, elliptic, oval, or oblong, leaf slightly thicker than paper and *Erigeron peregrinus*, smooth margin, rounded or notched apex

**Color:** Medium to dark green, usually darker than *Erigeron peregrinus*

**Texture:** Rougher surface than *Erigeron peregrinus*, hairless

**Length:** 5-10 mm from tip to tip

## Young stem

**Color:** Often reddish green when young and light green when older

**Texture:** Hairless when young, fuzzy white stem develops after first true leaves

**Length:** 5-10 mm from soil to cotyledons

## First true leaves

**Shape:** Elliptic, rounded or slightly pointed tip, smooth margin but older true leaves are dissected

**Color:** Medium green upper leaf surface, with whiter lower leaf surface

**Texture:** Long, white hairs form a dense horizontal mat on upper and lower leaf surface



**Left and middle:** Seedlings with cotyledons. Cotyledons have visual texture, while *Erigeron peregrinus* cotyledons appear smooth. **Right:** Seedling with cotyledons and first true leaves. See soft white hairs on true leaves, not found in *E. peregrinus*.



**Left:** Seedling with cotyledons and first true leaves. **Middle and right:** Older plants.

# Broadleaf lupine (*Lupinus latifolius*)

## Two cotyledons

**Shape:** Bean shaped, thick, and fleshy, smooth margin

**Color:** Semi-waxy, dark green upper leaf surface, with slightly lighter lower leaf surface

**Texture:** Hairless

**Length:** 5-10 mm from tip to tip

## Young stem

**Color:** Light green, red, or purplish red, with darker red coloring at base of stem

**Texture:** Hairless, but true leaf stem may have hairs

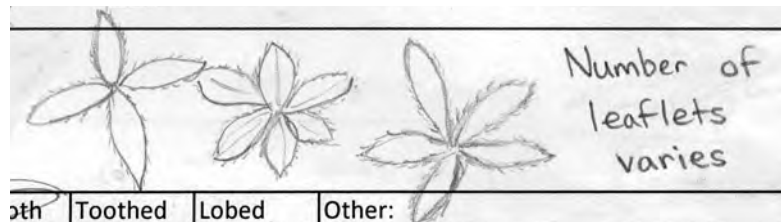
**Length:** 15-20 mm from soil to cotyledons

## First true leaves

**Shape:** Palmately compound with 3-7 leaflets (usually 4-6), leaflets elliptic or obelliptic

**Color:** Medium green

**Texture:** Hairs on margin



Seedlings with cotyledons only. See how cotyledons are bean shaped, the same shape as lupine seed.



Seedlings with cotyledons and first true leaves. Note that the number of leaflets on true leaves varies.

# Fringecup (*Tellima grandiflora*)

## Two cotyledons

**Shape:** Rounded but slightly triangular, widest at base, notched tip in older cotyledons

**Color:** Medium green, may have vein line down center of leaf surface

**Texture:** Small hairs on upper leaf surface

**Length:** Very tiny, less than 5 mm from tip to tip

## Young stem

**Color:** White

**Texture:** Hairless when young, white hairs develop after first true leaves

**Length:** Very short, less than 5 mm from soil to cotyledons

## First true leaves

**Shape:** Round with a scalloped margin, notched base, resemble miniature adult true leaves

**Color:** Medium green, with purplish veins

**Texture:** Small white hairs on petiole, leaf margin, and sometimes upper leaf surface



Seedlings with cotyledons. **Right:** Note notched tips on older cotyledons.



**Left:** Note seedling in bottom right photo corner with cotyledons and first true leaf. Practically impossible to distinguish from *Tolmiea menziesii* at this stage of development. **Middle and right:** Older plants. See how plant forms a basal rosette.



# Piggyback plant (*Tolmiea menziesii*)

## Two cotyledons

**Shape:** Rounded but slightly triangular, widest at base, notched tip in older cotyledons

**Color:** Medium green, may have vein line down center of leaf surface

**Texture:** Small white hairs, not obvious without a magnifying glass

**Length:** Very tiny, less than 5 mm from tip to tip

## Young stem

**Color:** White

**Texture:** White hairs

**Length:** Very short, less than 5 mm from soil to cotyledons

## First true leaves

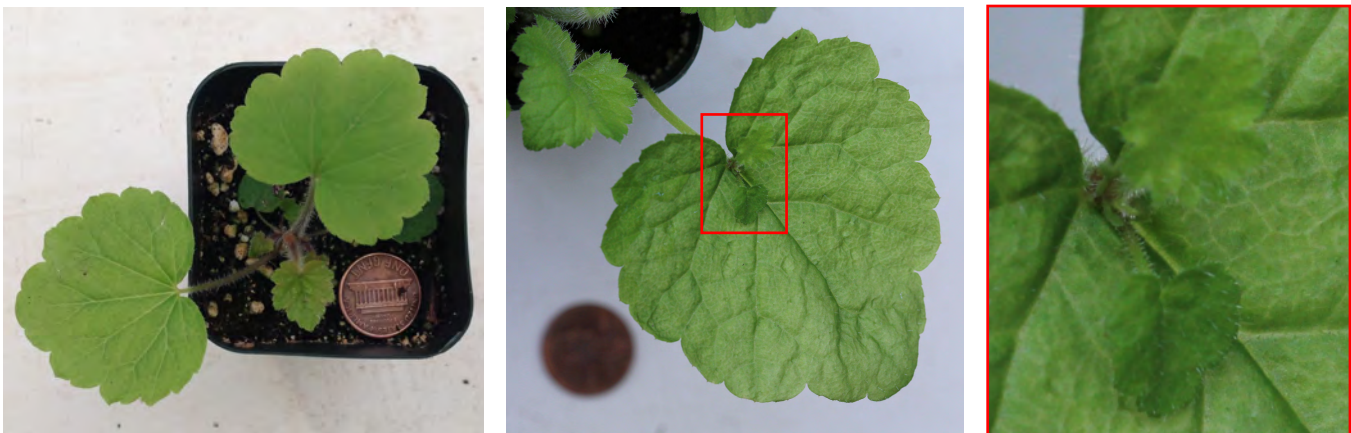
**Shape:** Round with a scalloped margin, notched base, resemble miniature adult true leaves

**Color:** Medium green, with palmate veins that are more distinct on older leaves

**Texture:** Dense white hairs on petiole, also hairs on margin and upper and lower leaf surface



**Left and middle:** Seedlings with cotyledons. **Middle:** Note notched tips on cotyledons. **Right:** Seedling with cotyledons and first true leaf. Indistinguishable from *Tellima grandiflora* at this stage.



Older plants. **Right:** Inset of middle photo. See tiny "piggyback" leaf at base of large leaf, hence the name piggyback plant. 41

# Beargrass (*Xerophyllum tenax*)

## One cotyledon

**Shape:** Linear, thick and flat, blunt or rounded tip, round cross section, seed often attached to tip

**Color:** Medium to dark green

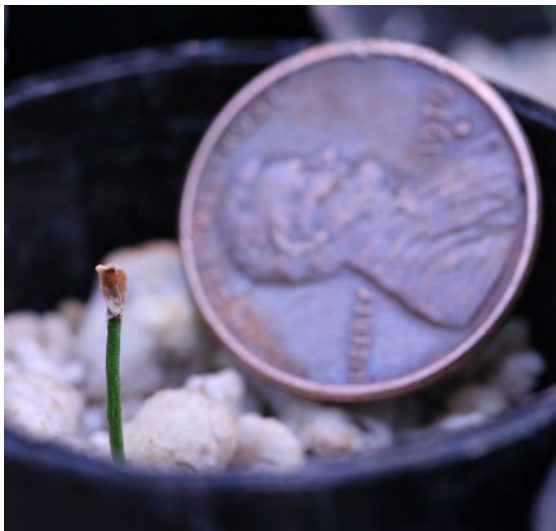
**Texture:** Hairless

## First true leaves

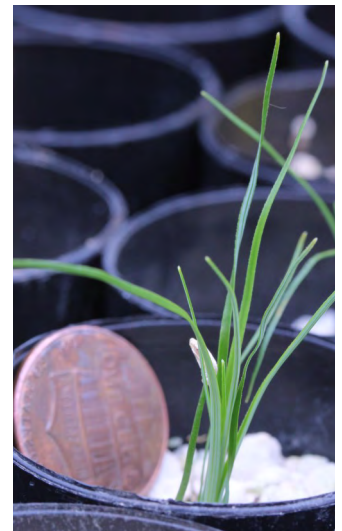
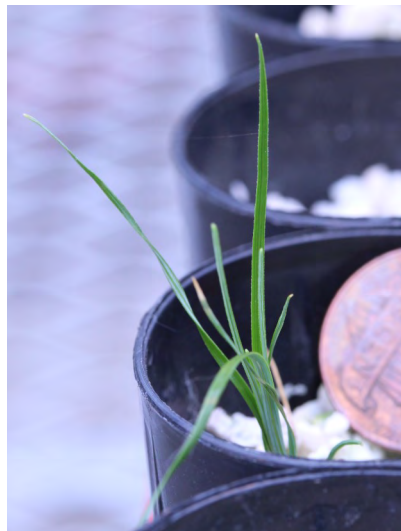
**Shape:** Linear, flat but substantial thickness, round or pointed tip

**Color:** Medium to dark green

**Texture:** Hairless



**Left:** Cotyledon with seed shell still attached. **Right:** Cotyledon after seed shell pops off cotyledon tip.



**Left:** Cotyledon with first true leaf. Note seed shell still attached to cotyledon tip. **Middle and right:** Older plants. Species resembles grass, despite being in the lily family.

# Sedges

Sedges can easily be confused with grasses and rushes, especially when young. An old rhyme can be used as a mnemonic to distinguish between these plant families. “Sedges have edges, rushes are round, grasses have knees that bend to the ground.” Grasses and rushes usually have round stems, while the stems of sedges are triangular, hence the edges.<sup>3</sup> However, sedge seedlings grow a round sheath at the base of their stalk before developing a triangular stem. This mnemonic is only useful for adult and older juvenile plants, after the clear development of the stem.



Another distinguishing feature of sedges is leaf arrangement. From a bird’s eye view, you can see leaves arranged in threes, with 120 degrees between each leaf.<sup>3</sup> This trait is visible in seedlings after the development of a few leaves.

# Showy sedge (*Carex spectabilis*)

## One cotyledon

**Shape:** Thin, flat leaf is vertical or slightly curved, round sheath at base visible with age, pointy tip

**Color:** Light green, whiter towards base

**Texture:** Hairless

## Stem

**Shape:** Cross section is triangular when older

**Color:** Whitish or light green at base

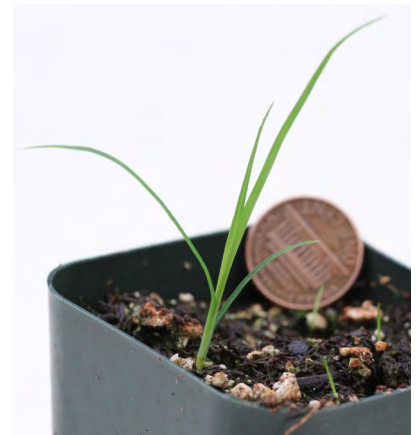
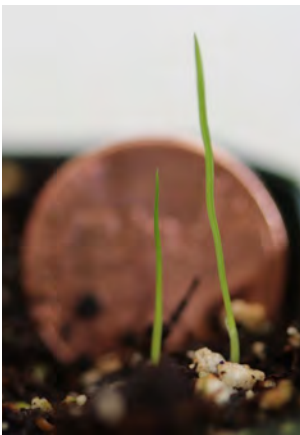
**Texture:** Striped pattern at base

## First true leaves

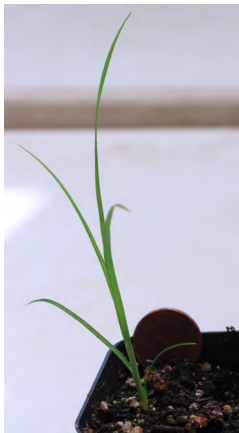
**Shape:** Linear tapering to a pointed tip, wide ridge down center of lower leaf surface develops with age, leaves narrower than *Carex stipata*

**Color:** Medium green

**Texture:** Hairless



**Left and middle:** Cotyledons. **Right:** Seedling with first true leaves. See how leaves are thinner than *Carex stipata*.



**Left:** Seedling with first true leaves. **Middle and right:** Older plants. **Middle:** Observe striped pattern on sheaths at stem base, in contrast to the cross hatched pattern in *Carex stipata*.

# Awlfruit sedge (*Carex stipata*)

## One cotyledon

**Shape:** Thin, flat leaf is vertical or slightly curved, round sheath at base visible with age, pointy tip

**Color:** Light green, whiter towards base

**Texture:** Hairless

## Stem

**Shape:** Cross section is triangular when older

**Color:** Whitish or light green at base

**Texture:** Cross hatching pattern at base

## First true leaves

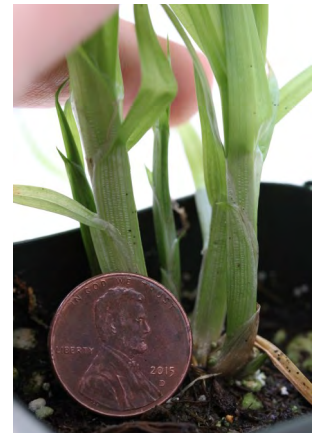
**Shape:** Linear tapering to a pointed tip, skinny ridge down center of lower leaf surface, wider leaves than *Carex spectabilis*

**Color:** Medium green

**Texture:** Hairless



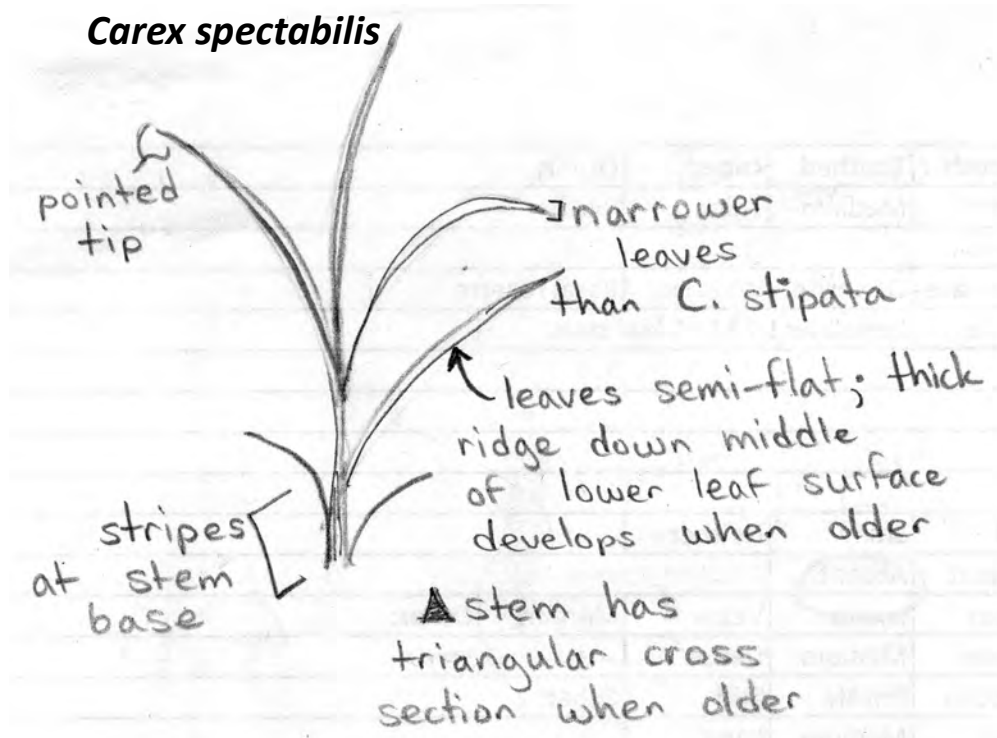
**Left:** Cotyledons. Visually impossible to identify at this stage of development. **Middle:** Seedlings with first true leaves. Note round whitish sheath at base of leaf, not found in *Xerophyllum tenax*. **Right:** Older seedling.



Older plants. See how leaves are wider than *Carex spectabilis*. **Right:** See cross hatched pattern on sheaths at stem base.

# Sedge (*Carex*) Comparison

*Carex spectabilis*



*Carex stipata*

There are many more species of sedges native to Washington than just these two species.

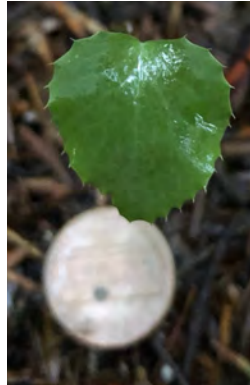
# Field Specimens



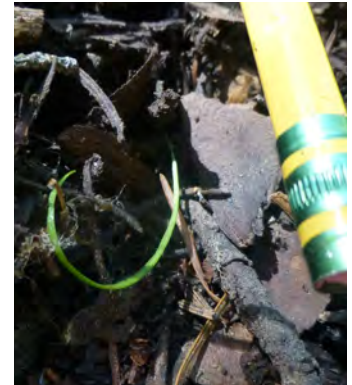
Subalpine fir  
(*Abies lasiocarpa*)



Western pasqueflower  
(*Anemone occidentalis*)



Tall Oregon grape  
(*Berberis aquifolium*)



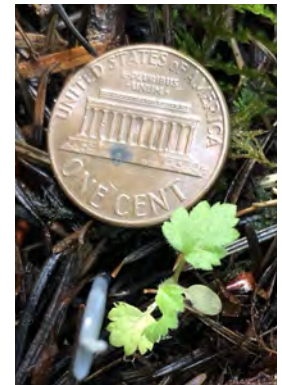
Showy sedge  
(*Carex spectabilis*)



Ponderosa pine  
(*Pinus ponderosa*)



Douglas fir  
(*Pseudotsuga menziesii*)



Trailing blackberry  
(*Rubus ursinus*)



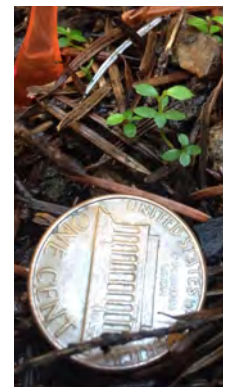
Blue elderberry  
(*Sambucus cerulea*)



Fringecup  
(*Tellima grandiflora*)



Piggyback plant  
(*Tolmiea menziesii*)



Cascade blueberry  
(*Vaccinium deliciosum*)

Photos from Hille Ris Lambers field crew members. Note that plants growing in the field may look slightly different than the same species growing in the greenhouse. Outdoor plants face higher levels of stress, such as drought, shading, and severe temperatures, which affect their appearance.

# Growing Native Plants From Seed<sup>4,5,6,7</sup>



Seeds, bundled and labelled for processing.

**Planning a seed propagation project.** Growing plants from seed usually takes longer than expected. Many seeds will not germinate if directly buried in soil! Instead, they require several months of pretreatment prior to sowing. Therefore, I researched the growing requirements for each species first. I also found a lab partner for my project since growing plants from seed is time-intensive and working with others makes the process more manageable. I collaborated with master's student Kimberly Ertel to germinate and grow the plants in this guide. Although we were using the plants for different projects, having multiple eyes on the plants proved invaluable. We created a calendar for different tasks, including pretreating and sowing seeds, to organize our propagation efforts.

**Buying and storing seeds.** After preliminary research, the next step was obtaining seeds. In this case, we used seeds that the Hille Ris Lambers Lab crew collected from the North Cascades with permit permission. The rest of the seeds were purchased from seed companies, namely, Inside Passage, Silvaseed Co., L&H Seeds, and Native Seed Foundation. The seeds were temporarily stored at room temperature in paper bags in a storage cabinet before we started working with them. However, seeds should be kept in a fridge or cold room if the duration of storage is longer than a few months. Over time, seeds lose their ability to germinate, but storing them in low temperature and low relative humidity conditions lengthens their lifetime.

**Sanitizing seeds.** Our work began with the crucial step of pretreating the seeds. Many seeds will not germinate and survive without pretreatment. First, we sterilized the seeds using bleach to eliminate any fungal spores or bacteria on the seeds that could kill seedlings. Note that we could have used hydrogen peroxide instead of bleach. To secure the seeds before sterilization, we placed each species in a square of mesh cloth, tied the cloth into a bundle with metal wire, and used a metal tag to label each bundle with the species name and date. Then, we filled a bucket with a solution of one part



# Growing Native Plants From Seed<sup>4,5,6,7</sup>



We soaked tiny seeds in cups (**above**) and the rest of the seeds in a tub of circulating water (**below**).

bleach to ten parts warm water, and soaked the seeds in the solution while stirring intermittently. After five minutes, the seeds were removed from the solution and rinsed with water. Secondly, we aerated the seeds in a tub of running water. This step promoted seed germination by softening the seed coats and leaching chemicals that inhibit germination from the seeds. For two days, we circulated the bundles of seeds in a tub full of water using aquarium air pumps. We placed a few species with tiny seeds, *Vaccinium deliciosum*, *Vaccinium parviflorum*, *Tellima grandiflora*, and *Tolmiea menziesii*, in their own cups of water, because these seeds were susceptible to escaping through the mesh cloth. The soaking water turned a yellowish color as chemicals leached from the seeds. To keep the water fresh, we dumped and refilled the water multiple times over the two days. After soaking, we rinsed the seeds again with water.

**Breaking seed dormancy.** Sanitizing and soaking are only the first part of pretreatment. Many species require further pretreatment to break seed dormancy, which equates to several months of patience. Seed dormancy occurs when a seed will not germinate in favorable environmental conditions. Seed dormancy is akin to hibernation in animals; basically, the seed needs to be woken up before germination can occur. In the lab, seed dormancy is broken by simulating environmental conditions that the seed would endure in nature. Dormancy breaking requirements, particularly the length of treatment, vary not only with species but sometimes with year and collection location.



# Growing Native Plants From Seed<sup>4,5,6,7</sup>



Seeds ready to be placed in the cold room for stratification.

There are multiple types of seed dormancy. Most species in this guide display physiological dormancy, which occurs due to high levels of hormones that inhibit seed germination. Physiological dormancy can be broken by cold stratification, where seeds are exposed to cold temperatures for a duration of weeks to months. Seeds that require cold stratification are native to places with cold winters and thus are genetically programmed to avoid germination in the middle of winter. We performed cold stratification by planting the bundles of seeds in shallow containers of moist peat moss and keeping the containers in a cold room at 5°C for 1-3 months. To prevent the seeds from drying out, we sprayed water on the peat moss every few weeks.

Another common type of seed dormancy is physical dormancy, where a hard seed coat prevents the embryo, the tiny plant inside, from busting out of the seed. Physical dormancy is broken through scarification where the seed coat is worn down with hot water, acid, or rubbing. The only species that we treated for physical dormancy was *Lupinus latifolius*. We used the hot water method because acid requires safety precautions and rubbing can easily damage the embryo. We heated up a cup of water, waited for the water to stop boiling, added the seeds to the water, and kept the seeds in the cup until the water reached room temperature. Then, we stratified the lupine seeds for thirty days.

Some species had little or no information on seed pretreatment requirements in the literature, namely, *Alnus rubra*, *Alnus viridis*, *Vaccinium deliciosum*, *Vaccinium parviflorum*, *Tellima grandiflora*, and *Tolmiea menziesii*. This information might be absent because these species have nondormant seeds, though this was not explicitly stated. It is also possible that these species require seed pretreatment, but few people have reported their methods. We treated these species with thirty days of stratification in case of seed dormancy. On the next page is a table of pretreatment methods that we used for each species. We obtained information on pretreatment methods primarily from the Native Plant Network, an online resource sponsored by the United States Department of Agriculture that maintains a Propagation Protocol Database for native plants.<sup>7</sup> We supplemented propagation information from the Native Plant Network with a few other sources, which are listed in the References section.<sup>4,5,6</sup>

## Seed Pretreatment Methods & Germination Notes<sup>7</sup>

Species	Pretreatment	Notes
<b>Conifers</b>		
Grand fir ( <i>Abies grandis</i> )	Cold strat. (30 days)	Very low germination
Subalpine fir ( <i>Abies lasiocarpa</i> )	Cold strat. (30 days)	
Engelmann spruce ( <i>Picea engelmanni</i> )	Cold strat. (30 days)	
Sitka spruce ( <i>Picea sitchensis</i> )	Cold strat. (30 days)	
Lodgepole pine ( <i>Pinus contorta</i> )	Cold strat. (30 days)	
Ponderosa pine ( <i>Pinus ponderosa</i> )	Cold strat. (30 days)	
Douglas fir ( <i>Pseudotsuga menziesii</i> )	Cold strat. (30 days)	
<b>Other Trees &amp; Shrubs</b>		
Vine maple ( <i>Acer circinatum</i> )	Cold strat. (90 days)	Very low germination
Rocky Mountain maple ( <i>Acer glabrum</i> )	Cold strat. (90 days)	Very low germination
Red alder ( <i>Alnus rubra</i> )	Cold strat. (30 days)	Seeds need light to germinate
Green alder ( <i>Alnus viridis</i> )	Cold strat. (30 days)	Seeds need light to germinate
Tall Oregon grape ( <i>Berberis aquifolium</i> )	Cold strat. (90 days)	Started germinating in strat.
Dwarf Oregon grape ( <i>Berberis nervosa</i> )	Cold strat. (90 days)	
Thimbleberry ( <i>Rubus parviflorus</i> )	Cold strat. (90 days)	
Trailing blackberry ( <i>Rubus ursinus</i> )	Cold strat. (90 days)	
Blue elderberry ( <i>Sambucus cerulea</i> )	Cold strat. (90 days)	
Red elderberry ( <i>Sambucus racemosa</i> )	Cold strat. (90 days)	
Cascade blueberry ( <i>Vaccinium deliciosum</i> )	Cold strat. (30 days)	
Red huckleberry ( <i>Vaccinium parviflorum</i> )	Cold strat. (30 days)	Very low germination
<b>Flowers</b>		
Pearly everlasting ( <i>Anaphalis margaritacea</i> )	Cold strat. (30 days)	Seeds need light to germinate
Western pasqueflower ( <i>Anemone occidentalis</i> )	Cold strat. (90 days)	
Broadleaf arnica ( <i>Arnica latifolia</i> )	Cold strat. (30 days)	
Mountain daisy ( <i>Erigeron peregrinus</i> )	Cold strat. (30 days)	Seeds need light to germinate
Common woolly sunflower ( <i>Eriophyllum lanatum</i> )	Cold strat. (90 days)	Started germinating in strat.
Broadleaf lupine ( <i>Lupinus latifolius</i> )	Scarification, then Cold strat. (30 days)	
False lily of the valley ( <i>Maianthemum dilatatum</i> )	Cold strat. (90 days)	No germination
False Solomon's seal ( <i>Maianthemum racemosum</i> )	Cold strat. (90 days)	No germination
Fringecup ( <i>Tellima grandiflora</i> )	Cold strat. (30 days)	
Piggyback plant ( <i>Tolmiea menziesii</i> )	Cold strat. (30 days)	
Beargrass ( <i>Xerophyllum tenax</i> )	Cold strat. (30 days)	No germination
<b>Sedges</b>		
Showy sedge ( <i>Carex spectabilis</i> )	Cold strat. (90 days)	
Awlfruit sedge ( <i>Carex stipata</i> )	Cold strat. (30 days)	

**Note:** For species with poor germination, longer stratification might yield higher germination results.

# Growing Native Plants From Seed<sup>4,5,6,7</sup>

**Avoiding problems during stratification.** Some of our seeds molded during stratification. In retrospect, peat moss was not the best medium choice. An inorganic medium such as sterilized sand, as opposed to an organic medium such as peat moss, might have reduced our mold problems. Additionally, it would have been better to keep the medium lightly moist, rather than heavily so. Fortunately, we were able to mitigate mold problems by regularly checking on the seeds. Once a week, we opened the mesh bags, rubbed mold off the seeds, and discarded the squishy seeds. We also soaked the seeds in a 1:10 diluted bleach solution during the middle of stratification, which temporarily reduced the amount of mold. If we had not closely monitored the seeds, they probably would not have survived.

Additionally, a few species started germinating in stratification before we were ready to sow seeds. Some of these seedlings died because they were held too long in stratification. When scheduling planting, remember that some species may need to be planted earlier than anticipated. Frequently monitoring the health and stage of development of the seeds is critical to stratification success!



Every week, we checked seeds in stratification for seed mold (**left**) and early germination (**right**).

**Planting seeds and transplanting seedlings.** To streamline our planting process, we scheduled stratification such that all seeds were ready to be planted at approximately the same time. We followed the general rule of thumb for seed sowing, that is, planting seeds as deep as their length. For example, we covered Douglas fir seeds with about 3/4 cm of soil, estimated by eye, because these seeds are about 3/4 cm long. All seeds were covered with soil except species with photodormant seeds, which need light to germinate. We planted the seeds in Sunshine 4 mix, but any seedling mix should do. We sowed tree seeds in cone-tainers, with a few seeds per cone-tainer, and sprinkled flower, sedge, and shrub seeds over flats. To speed up germination and seedling growth, the soil was covered with plastic lids to create miniature terrariums that maintained high humidity levels. After germination, we thinned tree seedlings to one per cone-tainer. We transplanted the rest of the seedlings from flats to 4-inch rose pots after they developed their first true leaves. Transplanting was time consuming, but starting non-tree seeds in flats conserved space. Note that we started the tree seedlings in cone-tainers rather than flats since they transplant poorly and develop long roots quickly.

# Growing Native Plants From Seed<sup>4,5,6,7</sup>



Layout on the greenhouse bench.

**Germination and growing conditions.** We grew our plants in spring in the greenhouse at the Center for Urban Horticulture at the University of Washington, where temperatures ranged from 68-75°F and the lights were on from 7 am to 5 pm daily. Germination took place over a span of a few days to weeks, with most germination occurring 1-2 weeks after planting. Trees and other large seeded plants typically took longer to germinate than herbs but had larger seedlings. After the plants were large enough, the greenhouse staff started watering our plants with diluted Dyna-Gro fertilizer instead of regular water. Fertilizing is important because most seedling soil mixes contain low levels of nutrients that cannot sustain a rapidly growing plant. Ideally, you should begin fertilizing after the first set of true leaves have emerged. We learned this the hard way and had to rescue our plants after their leaves started to change color due to nutrient deficiencies. Other dangers to seedling survival include overwatering, which causes yellow leaves and root rot, and underwatering, which causes drooping leaves. Seedlings need to be monitored closely since they are more sensitive to their environment than mature plants.

**Records.** I kept records so I could remember my propagation process, avoid making the same mistakes twice, and share knowledge with people interested in growing the same species. Along the way, I recorded pretreatment and planting methods, germination results, and observations. On the following page is a table of the identification traits that I recorded in my observation datasheets.



We thinned seedlings to avoid crowding (**left**). We sowed tree seeds in cone-tainers since seedlings grow long roots (**right**).



## Observation Datasheet for Seedling Identification Traits

Observation Date:		Species:		
Plant Organ	Trait	Classification	Definitions	
Leaves (Cotyledons, True Leaves)	Number	Monocot	One cotyledon	
		Dicot	Two cotyledons	
		Polycot	More than two cotyledons	
	Shape			Circular, cordate, elliptic, linear, obelliptic, oblong, obovate, oval, ovate, palmately compound, spatulate, ternately compound, triangular... also note shape of leaf tips and base
		Leaf outline		Compound, triangular... also note shape of leaf tips and base
		Leaf margin		Coarse or finely jagged, dissected, lobed, scalloped, smooth...
		Leaf thickness		Thin, medium, thick
		Leaf angle		Needles horizontal or cupped up, or leaf tilted up or down
	Color			Note color and any difference between upper and lower leaf surface; note if color is uniform, striped, or another pattern
		Leaf surface		Note if color is uniform, striped, or another pattern
		Leaf vein		Note if different color than leaf surface
		Leaf hairs		Note if different color than leaf surface
	Texture	Leaf shine		Dull, shiny, or regular leaf surface
		Leaf hairs		Present or absent
		Location of hairs		May be present on upper or lower leaf surface, vein, margin, petiole
		Density of hairs		Sparse, medium, dense
		Quality of hairs		Smooth, prickly, stiff; may secrete sticky substance from tip
Length of hairs			Short, medium, long	
Attachment	Arrangement		Stick straight up, angled, form interwoven mat	
	Surface quality		Waxy, smooth, rough, prickly, powdery, or bumpy surface; note any difference between upper and lower leaf surface	
	Sessile		No petiole	
Stem	Petiolate		Long, medium, or short petiole	
	Shape	Cross section	Round, flat, or triangular stem	
	Color	Stem	Note color and if color pattern is uniform, striped, or other	
Overall	Texture	Stem hairs	Look for same traits as leaf hairs	
	Size		Length and width of cotyledons, seedling height	
Notes	Growth habit		Upright, sprawling, semi-sprawling, basal rosette	
			Additional information	

I made a datasheet for efficiently collecting observations of the cotyledons, young stem, and true leaves of each species. Above is a condensed version of my datasheet, which I created after examining the traits included in the seedling identification guides listed under the References section. For the seedling photographs, I used a Canon EOS Rebel T6 DSLR camera with an EF-S 60mm f/2.8 fixed macro lens and a tripod for stabilization. I took photographs of each species from multiple angles, particularly top-down and side views, to capture both stem and leaf traits. I also photographed sequential stages of development over the course of several months in order to showcase the transition from cotyledons to first true leaves to older seedlings. While reviewing, reorganizing, and consolidating my notes, I selected photographs to emphasize the key traits of each species in my guide. Plant identification is a science and an art, and I hope that my guide will be appreciated as both.

# Elevation and Climate



Elevation matters to plants, mainly because the temperature drops as elevation increases. Since each plant species tolerates a limited temperature range, species will only be found in their native habitat at certain elevations. It is uncommon to discover high elevation species at low elevations, or low elevation species at high elevations. Furthermore, each species prefers different levels of rainfall and soil moisture. Some species are more drought tolerant than others. Any houseplant owner has observed this fact; certain species are more likely to survive if insufficiently watered. Thus, both elevation and climate can serve as a clue to the identity of a species. Additionally, species grown outdoors will perform poorly if planted outside of their preferred elevation range and climate. This is a word of caution to those who might otherwise be enticed into growing pretty alpine wildflowers at low elevations! On the following page is a table of the general elevation and climate preferences of species in this guide. This table is an oversimplification since species preferences are often nuanced.

# Elevation and Climate Preferences

8,9,10,11

	Lower Elevations	Higher Elevations	Low & High Elevations
Moist Climate	<b>Vine maple</b> <i>(Acer circinatum)</i> <b>Red alder</b> <i>(Alnus rubra)</i> <b>Dwarf Oregon grape</b> <i>(Berberis nervosa)</i> <b>Showy sedge</b> <i>(Carex stipata)</i> <b>Sitka spruce</b> <i>(Picea sitchensis)</i> <b>Thimbleberry</b> <i>(Rubus parviflorus)</i> <b>Red elderberry</b> <i>(Sambucus racemosa)</i> <b>Fringecup</b> <i>(Tellima grandiflora)</i> <b>Piggyback plant</b> <i>(Tolmiea menziesii)</i> <b>Red huckleberry</b> <i>(Vaccinium parvifolium)</i>	<b>Broadleaf arnica</b> <i>(Arnica latifolia)</i> <b>Awlfruit sedge</b> <i>(Carex spectabilis)</i> <b>Engelmann spruce</b> <i>(Picea engelmannii)</i>	
Dry Climate	<b>Pearly everlasting</b> <i>(Anaphalis margaritacea)</i> <b>Common wooly sunflower</b> <i>(Eriophyllum lanatum)</i> <b>Ponderosa pine</b> <i>(Pinus ponderosa)</i> <b>Blue elderberry</b> <i>(Sambucus cerulea)</i>	<b>Subalpine fir</b> <i>(Abies lasiocarpa)</i> <b>Western pasqueflower</b> <i>(Anemone occidentalis)</i> <b>Mountain daisy</b> <i>(Erigeron peregrinus)</i>	
Moist & Dry Climates	<b>Grand fir</b> <i>(Abies grandis)</i> <b>Rocky Mountain maple</b> <i>(Acer glabrum)</i> <b>Green alder</b> <i>(Alnus viridis)</i> <b>Tall Oregon grape</b> <i>(Berberis aquifolium)</i> <b>Douglas fir</b> <i>(Pseudotsuga menziesii)</i>	<b>Cascade blueberry</b> <i>(Vaccinium deliciosum)</i>	<b>Broadleaf lupine</b> <i>(Lupinus latifolius)</i> <b>Lodgepole pine</b> <i>(Pinus contorta)</i> <b>Beargrass</b> <i>(Xerophyllum tenax)</i>

Most of the species in this guide were grown to support field research conducted on Mt. Rainier and North Cascades by the Hille Ris Lambers Lab. To study the effects of climate change on plant communities, the lab selected pairs of native species in the same genus with different elevation and/or climate preferences. 56



# Acknowledgements

Thanks to master's student Kimberly Ertel, whom I collaborated with to germinate and grow the plants in this guide. Without her, I would not have been able to undertake a project of this scale. Additional thanks to Dr. Janneke Hille Ris Lambers for advising my project, and the Hille Ris Lambers Lab and field crew for support. A shout out to the University of Washington greenhouse staff who helped keep our plants alive. Also, thanks to research botanist Courtney Bobsin, who allowed me to take photographs of her *Xerophyllum tenax* seedlings when ours failed to germinate. Furthermore, I acknowledge a few people who were kind enough to answer my questions on plant propagation: Restoration Ecologist Brenda Clifton of the Skagit River System Cooperative, Horticulturist Josh Drown of Mount Rainier National Park, and George Macomber of the Washington Native Plant Society. Josh Drown also gave us an awesome tour of the Mount Rainier National Park greenhouse. Lastly, I am grateful for the University of Washington Biology Department for providing funding for my project.



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## Citations

### Conifer Seedling Identification

<sup>1</sup>Franklin, Jerry. *A Guide to Seedling Identification for 25 Conifers of the Pacific Northwest*. United States Department of Agriculture, 1961. <https://www.biodiversitylibrary.org/item/209039#page/10/mode/1up>.

### Leaf Shapes and Other Plant Terminology

<sup>2</sup>Harris, James and Melinda Harris. *Plant Identification Terminology: An Illustrated Glossary*, 2nd Edition. Spring Lake, Utah, Spring Lake Publishing, 2001.

### Pacific Northwest Plant Identification

<sup>3</sup>Giblin, David, Ben Legler, Peter Zika, and Richard Olmstead, eds. *Flora of the Pacific Northwest: An Illustrated Manual* by C. Leo Hitchcock and Arthur Cronquist, 2nd Edition. University of Washington Press, 2018.

### Plant Propagation Protocols

<sup>4</sup>Baskin, Carol, and Jerry Baskin. *Seeds: Ecology, Biogeography, and Evolution of Dormancy and Germination*. Academic Press, 1998.

<sup>5</sup>Dumroese, R. Kasten, Tara Luna, and Thomas Landis, eds. *Nursery Manual for Native Plants: A Guide for Tribal Nurseries— Volume 1: Nursery Management. Agriculture Handbook 730*. United States Department of Agriculture, Forest Service, 2009. [https://www.fs.fed.us/rm/pubs\\_series/wo/wo\\_ah730.pdf](https://www.fs.fed.us/rm/pubs_series/wo/wo_ah730.pdf).

<sup>6</sup>Hartmann, Hudson, Dale Kester, Fred Davies, and Robert Geneve. "Chapter Seven: Principles of Propagation from Seeds." *Hartmann and Kester's Plant Propagation: Principles and Practice*, 8th ed., Prentice Hall, 2011, pp. 200-249.

<sup>7</sup>Native Plant Network. "Propagation Protocols." *Native Plant Network — Reforestation, Nurseries and Genetics Resources*, United States Forest Service and Southern Regional Extension Forestry. <https://npn.rngr.net/npn/propagation>.

### Plant Ecology and Habitat Preferences

<sup>8</sup>Biek, David. *Flora of Mount Rainier National Park*. Oregon State University Press, 2000.

<sup>9</sup>Giblin, David, and Ben Legler, eds. *WTU Image Collection Website: Vascular Plants, Macrofungi, and Lichenized Fungi of Washington State*. University of Washington Herbarium, 2003+. Accessed Feb 2019. <http://biology.burke.washington.edu/herbarium/imagecollection.php>.

<sup>10</sup>Pojar, Jim, and Andy MacKinnon, eds. *Revised Plants of the Pacific Northwest Coast: Washington, Oregon, British Columbia & Alaska*. Lone Pine Publishing, 2004.

<sup>11</sup>USDA Forest Service. *Fire Effects Information System*. <https://www.fs.fed.us/database/feis/plants>.

## Other Helpful Resources

### How to Identify Plants

Mangold, Jane, and Hilary Parkinson. *Plant Identification Basics*. Montana State University Extension, 2013. <http://msuextension.org/publications/AgandNaturalResources/MT201304AG.pdf>.

### More Seedling Guides

Parkinson, Hilary, Jane Mangold, and Fabian Menalled. *Weed Seedling Identification Guide for Montana and the Northern Great Plains*. Montana State University Extension, 2013. <http://msuextension.org/publications/AgandNaturalResources/EB0215.pdf>.

Pavek, Pamela, Brenda Erhardt, Trish Heekin, and Richard Old. *Forb Seedling Identification Guide for the Inland Northwest: Native, Introduced, Invasive, and Noxious Species*. Natural Resources Conservation Service, 2012. [https://www.nrcs.usda.gov/Internet/FSE\\_PLANTMATERIALS/publications/wapmcpu11331.pdf](https://www.nrcs.usda.gov/Internet/FSE_PLANTMATERIALS/publications/wapmcpu11331.pdf).

Tveten, Richard, and Melissa Asher. *Seedling Identification Guide for Columbia Basin Upland Restoration Sites*. Washington Department of Fish and Wildlife, 2011. <https://wdfw.wa.gov/publications/01370/wdfw01370.pdf>.

