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The primary goal of this proposal is to develop improved enzymes for the inactivation and degradation of fungal mycotoxins associated with Fusarium head blight. This year it is planned to utilize the three-dimensional structure and kinetic properties of trichothecene 3-*O*-acetylase from *F. sporotrichioides* and *F. graminearum* (Tri101) to develop a modified enzyme with improved efficacy towards the inactivation of DON and nivalenol. This will be accomplished by directed evolution using the structure of Tri101 that was determined during the past year as a template for defining the components that need to be redesigned. The prospect of success in this first phase is high because the kinetic analysis of this enzyme suggest that there are already significant differences in specificity between isozymes from different fungi. A new component of the investigation this coming year will be to integrate the *in vitro* studies of Tri101 with the properties of the enzyme expressed in transgenic cereals to determine whether the limited performance of the transgenic cereals is due to low expression, inactive, or posttranslationally modified protein. In the final component of this project it is planned to develop new biodegradative agents based on the oxidative enzymes in the trichothecene biosynthetic pathway. Much less is known about these enzymes so it is planned to continue with their structural and biochemical characterization. This will establish a framework for expanding the repertoire of biodegradative agents in future years. Thus the specific aims of the project are to:

- 1. apply directed evolution to the trichothecene 3-*O*-acetylase (Tri101) from *F. sporotrichioides* and *F. graminearum* and screen for protection against DON and nivalenol in *Saccharomyces cerevisiae*. This is the first priority, since *tri101* has been shown to provide partial protection against the spread of *F. graminearum* in transgenic wheat.**
- 2. correlate the structure and function of the Tri101 protein produced in *E. coli* with that isolated directly from *F. sporotrichioides* and *F. graminearum* and from transgenic barley. This will ascertain the level of activity of the enzyme expressed in transgenic barley and establish a connection between the *in vitro* and *in vivo* studies of Tri101.**
- 3. express, purify, and initiate a structural and biochemical analysis of the oxidative biosynthetic enzymes in the trichothecene biosynthetic pathway.**

The work proposed here should lead to improved biological agents for inactivating mycotoxins and is expected to aid in the development of better methods for controlling FHB. At a fundamental level these studies will contribute to a greater understanding of how trichothecene mycotoxins are synthesized.