



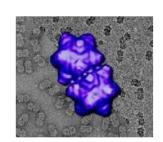
# Recovery Plan – Draft— Cotton leaf curl disease

(

): A whitefly-transmitted virus causing leaf curl disease of cotton, vegetable, and ornamental crops



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# **Economics of cotton**

- Cultivated cotton Gossypium species (L.) have been a major source of food, feed and fiber worldwide for over 7000 years.
- Globally ~32.6 million hectares are devoted to cotton cultivation with production estimated at 27.6 million tons for 2011/2012 (Anonymous, 2011).
- Cotton is placed sixth on the List of Economically Important Crops for the U.S.

#### Virus-like disorders of cotton

Disease	Vector/Transmission	Geographical Distribution
Acromania	unknown	USA (Arizona), China, Haiti,
Cotton (common) yellow mosaic	Bemisia tabaci	Sudan
	B. tabaci	Brazil
Cotton mosaic	unknown	
	B. tabaci	Central America
Cotton leaf curl		USA (Louisiana)
	grafting	0 %
Cotton leaf mottle	Aphis gossypii	Philippines
Cotton leaf roll	A. gossypii	Sudan, USSR
/leaf curliness		Thailand,
		Paraguay
Infectious	grafting,	India
variegation	B. tabaci	
Indian leaf crumple	grafting, B. tabaci	India
Murcha vermelha /purple wilt	B. tabaci	South America
Terminal stunt	A. gossypii	USA (Texas)
Tobacco streak	possibly thrips	Brazil



Caribbean



Central America



Diverse symptoms: whitefly-associated virus-like diseases











#### Three major cotton-infecting viruses identified and characterized

## **New World**:

: AZ, CA,

TX, Mexico, Guatemala, Caribbean, Brazil

















# <u>Asia</u>

complex (7+ species)







# **Africa**

Cotton leaf curl Gezira virus Nile Basin











# Specific cotton begomovirus genomes & satellites

## New World

- Cotton leaf crumple virus

## **Africa**

- Cotton leaf curl Gezira virus

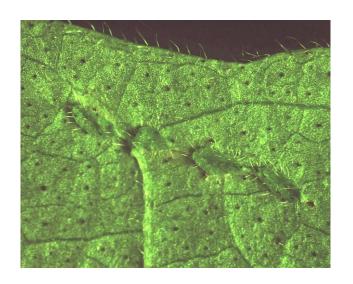
### <u>Asia</u>

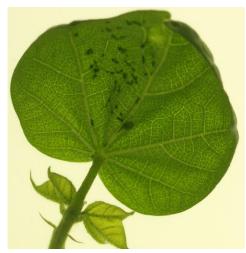
- Cotton leaf curl Allahabad virus
- Cotton leaf curl Bangalore virus
- Cotton leaf curl Burewala virus
- Cotton leaf curl Kokhran virus
- Cotton leaf curl Multan virus
- Cotton leaf curl Rajasthan virus
- Cotton leaf curl Shadadpur virus

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# Proliferation of spongy parenchyma and bundle sheath extension cells, and hyperplastic cells (transverse sections) = Enations are a unique feature of OW leaf curl begomoviruses

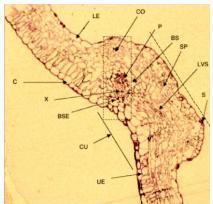
BS: bundle sheath, BSE: bundle sheath extension, C: cuticle, CO: collenchyma cells, CU: leaf curvature or vein groove, LE: lower epidermis, LVS: secondary vein, UE: upper epidermis, P: phloem, S: stoma, SP: spongy parenchyma, X: xylem.



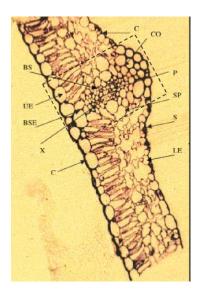


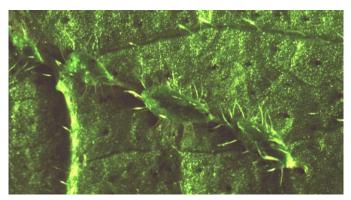


Box = main vein



healthy





**Cotton leaf curl** disease (CLCuD) is a debilitating disease of cotton that results in leaf curling, development of leaf-like enations on the undersides of leaves, overall stunting of the plant, and reduced yield and quality.

- -CLCuD occurs in Africa, Pakistan, and northwestern (Punjab region) India.
- -Caused by **one or more whitefly-transmitted geminiviruses** (family *Geminiviridae*, genus *Begomovirus*) that serve as the 'helper virus' for to different kinds of DNA satellite molecules of non-viral origin: **betasatellites and alphasatellites**.

The satellites contribute to host defense suppression, leading to increased virulence of the helper virus that leads to systemic infection and severe disease symptoms; the promiscuity of the beta/alpha satellites may confer versatility to helper viruses.

-The host range of the CLCuD virus-satellite complexes has been found to be astoundingly broad, spanning multiple plant families, albeit, it was initially thought to be restricted to cotton and several wild malvaceous hosts.

Since 1990 Pakistan and India have experienced two epidemics:

- the most recent is the result of an emergent, resistance breaking strain (2001-present)
- -overcame host-plant resistance in varieties developed to combat the first outbreak occurring in about 1990.

	Brief history: Leaf curl viruses of cotton
1912-1960	Nigeria (1912- <i>G. barbadense</i> ), Sudan (1924) –Leaf curl Gezira virus, Tanzania (1926), USA-CA/AZ leaf crumple virus (1951), Philippines (1959). In all instances - symptoms associated with whitefly <i>Bemisia tabaci</i> presence.
1967-1973	Leaf curl symptoms first reported in Pakistan in Multan, Punjab Provinces; by 1973 'Cotton leaf curl' disease; Cotton leaf crumple occurs annually in cotton, US/Mexico.
1983-1993	CLCD outbreak in cotton near Multan –resulting in the first epidemic (80% incidence), spreading throughout Pakistan and into western India. The disease emerged again during 1989 and 1992-1994 (entire cotton belt).
1994-1999	Common Fund-Cotton project initiated with NIBGE (Zafar-NIBGE and Brown-UA); first effort to characterize the causal agent; identified a monopartite begomoviral genome and beta satellite complex. Cotton viruses in Sudan and American Tropics studied; causality proven for two main species (Brown et al) + Leaf curl in Pakistan. Breeding yielded resistant cotton varieties—production returned to pre-epidemic levels.

Leaf curl symptoms reported in cotton fields in the Sindh Province.

First appearance of resistance breaking in cotton in western India.

and cotton India and Pakistan (Kirthi, Rehman, Shahid)

Evidence for more than one viral genome: Cotton leaf curl Alalabad virus (CLCuAV) and

Cotton leaf curl Khokhran virus (CLCuKV); Papaya leaf curl virus (Mansoor et al., 2003b) and Tomato leaf curl Bangalore virus (Kirthi et al 2004); Cotton leaf curl Rajasthan virus in tomato

All cotton varieties susceptible – tolerance in some lines; presence of a recombinant virus

+ Multan beta satellite: Cotton leaf curl Burewala virus (Amrao et al., 2010b) = parental

viruses: CLCuMuV and CLCuKoV; subsequently spread to India = Current Project

1997

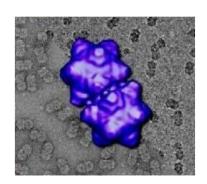
2001

2001-2004

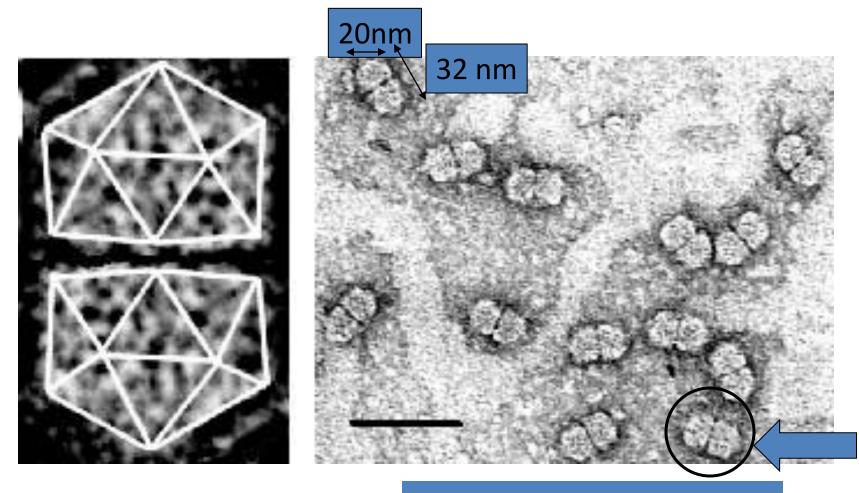
2009-12

# Primary causal agents: Whitefly-transmitted geminiviruses

- Family: Geminiviridae
- Genus: Begomovirus
- Distributed worldwide
- Infect dicotyledonous hosts
- Monopartite begomoviruses one circular ssDNA component ~2.8 kb
- Bipartite begomoviruses two ss circular DNA components ~2.6 kb each

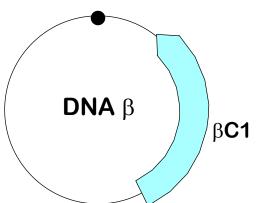


# Particle Morphology



One ssDNA molecule per particle

**DNA**  $\beta$  satellite

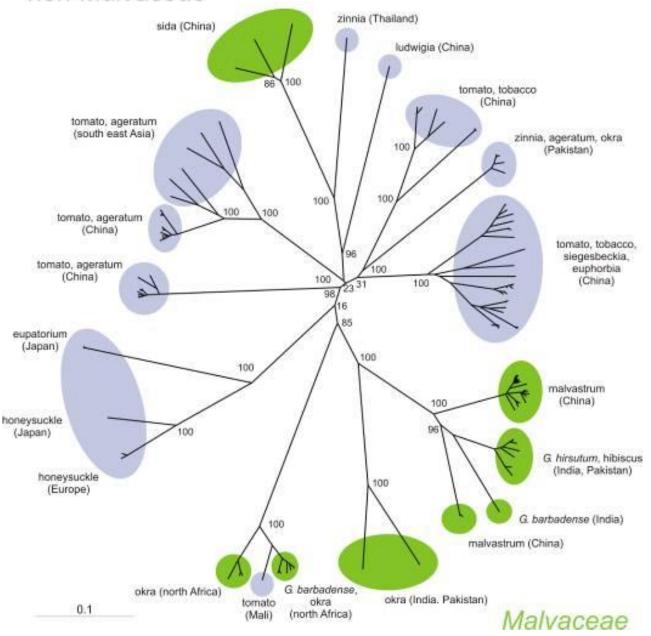


- Structure of DNA  $\beta$
- A single conserved ORF
- A region rich in adenine (approx. 300bp)
- A region conserved between all DNA βs (approx. 300bp)
- A hairpin structure (ori)

#### Properties of DNA $\beta$

- Trans-replicated by begomovirus encoded Rep
- Requires helper virus CP for transmission
- Essential for symptom development by helper virus
- Enhances helper virus DNA accumulation

#### non-Malvaceae



#### The vector

#### sibling species group



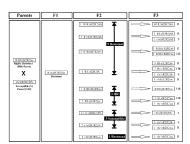
Kingdom:	Animalia
Phylum:	Arthropoda
Class:	Insecta
Order:	Hemiptera
Family:	Aleyrodidae
Genus:	Bemisia
Species:	tabaci

#### Characteristics

- Polyphagous insect (collectively, 500 host plant species)
- Haplodiploid reproduction
- Exclusively, phloem feeding
- Phylogenetically divergent mtCOI (0-26%)
- Variants referred to as haplotypes (mtCOI), or biotypes (mtCOI divergence + biological data)
- mtCOI phylogeographic distribution
- Causes billions of dollar losses annually worldwide
- Highly fecund
- Only insect vector, genus: Begomovirus (family, Geminiviridae)
- Recent examples invasive movement: TYLCV worldwide owing to dispersal via ornamentals, seedlings, whitefly; Cotton leaf curl virus-Pakistan, India, China

#### Cotton leaf curl virus timeline

Resistance attributed to 3 genes: two R genes and "suppressor"



First report

1988

1967

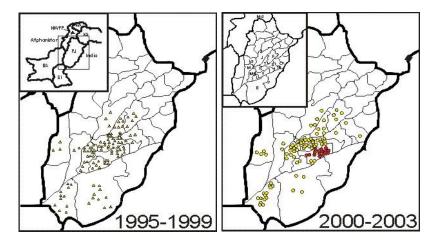
Sporadic isolated infections of cotton

Start of the CLCuV epidemic

Introduction of resistant varieties late 1990s

Introduction of an American cotton variety (S12)

\$5 billion losses from 1992-1997 due to CLCuD



2001

Resistance breaking

Second Epidemic:

recombinant Bur virus + CLCuMB

Courtesy R. Briddon)

#### The "Multan" strain of CLCuD

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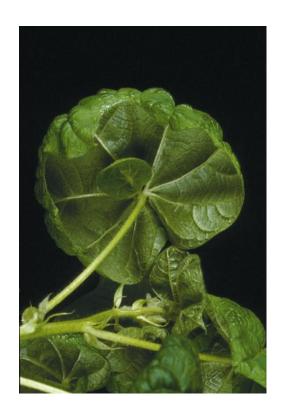
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One betasatellite (CLCuMB)



- Pre-resistance breaking 7 viruses one betasatellite
- Post -resistance breaking 1 virus one betasatellite

# **Disease Severity Ratings 0-4**



Brown Lab: SNP detection in CLCuV and associated satellites

# Estimated losses due to CLCVD

(Total)

1711.63

1435.29

1480.35

1956.62

1370.12

569.60

nancial

(Bill.PKR)

14.063

13.778

16.079

44.160

48.457

11.815

(Mill.Bales)

1.23

0.95

1.11

1.84

1.16

0.48

Year of infestation	Estim	ated crop dan	nage	Estimated Los	l Fin ses

(Complete)

25.21

2.50

40.25

43.39

28.33

0.00

(Partial)

1686.40

1432.80

1440.10

1693.50

1341.80

569.60

2006-07

2007-08

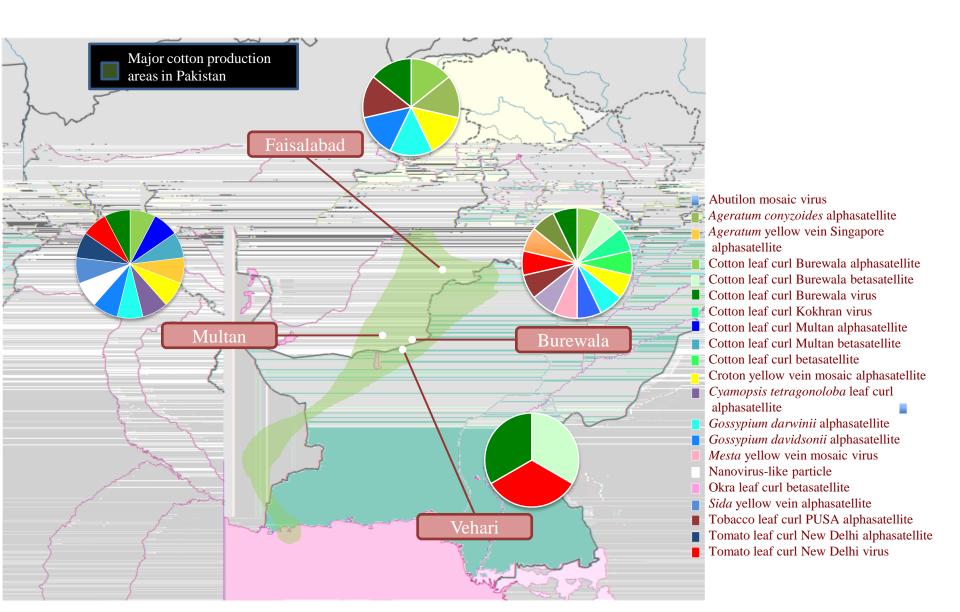
2008-09

2009-10

2010-11

2011-12

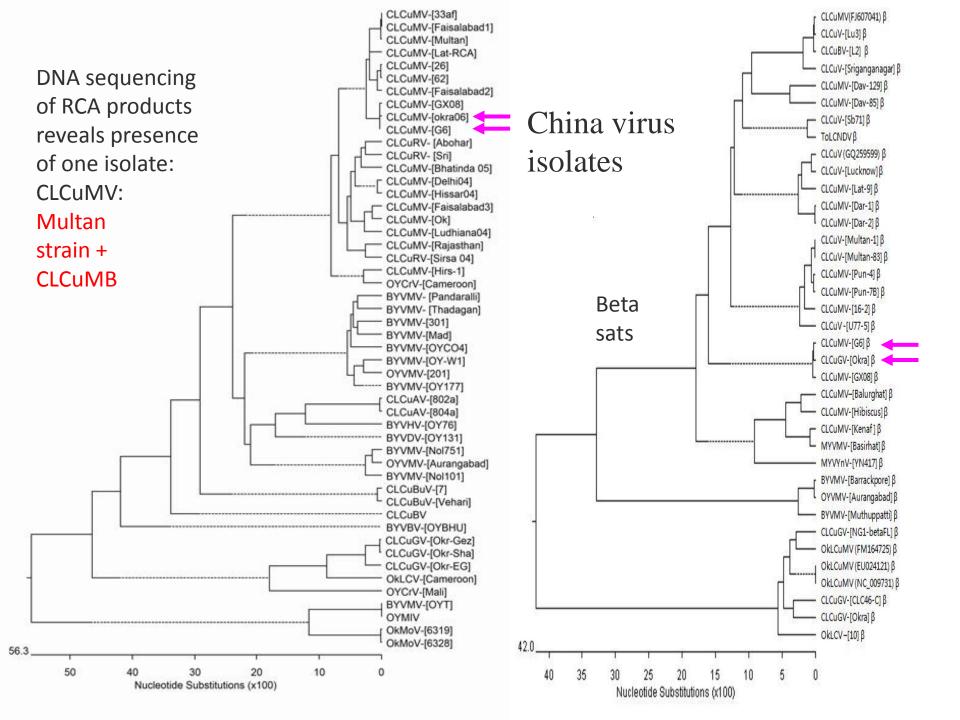
### Viruses and satellites in cotton, Pakistan 2012



Brown Lab: SNP detection in CLCuV and associated satellites

# **CLCuMV** distribution in China-2012





# Virus detection to populations.....

- Entire venue for virus detection and identification of species and variants has evolved from:
- No molecular techniques to -DNA hybridization to Polymerase chain reaction to - genome sequencing –Rolling circle amplification –to New Generation sequencing
- Enables high throughput vs single samples
- Facilitating views of diversification and population structure in a new light

# from....PCR strategies for ID & diversity studies

#### core Cp

quick ID: universal primers & determine

seq in a single run (576-579 bp)

#### full **CP**

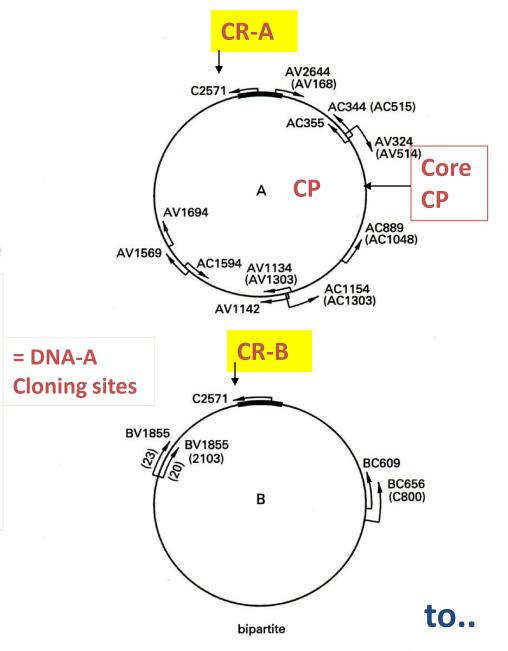
1.1 kbp fragment; two sequencing rxns; provisional ID and relationships = DNA-A)

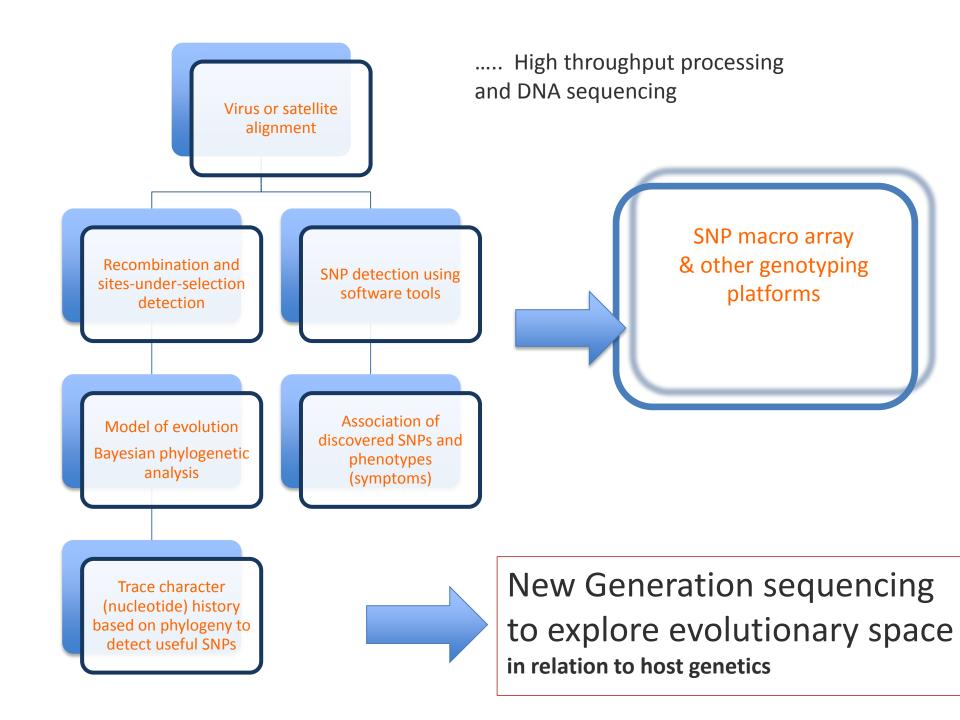
2/3 DNA-A overlap w/ CP CP & CR (2.1 kbp)

Top Half B

CR - cognate component

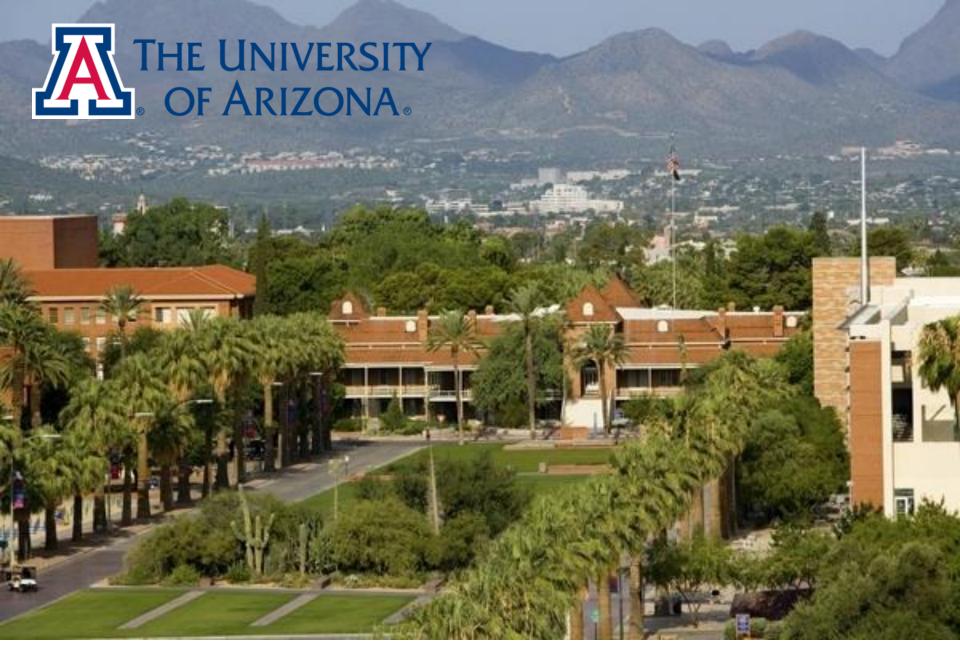
(1.5 kbp)





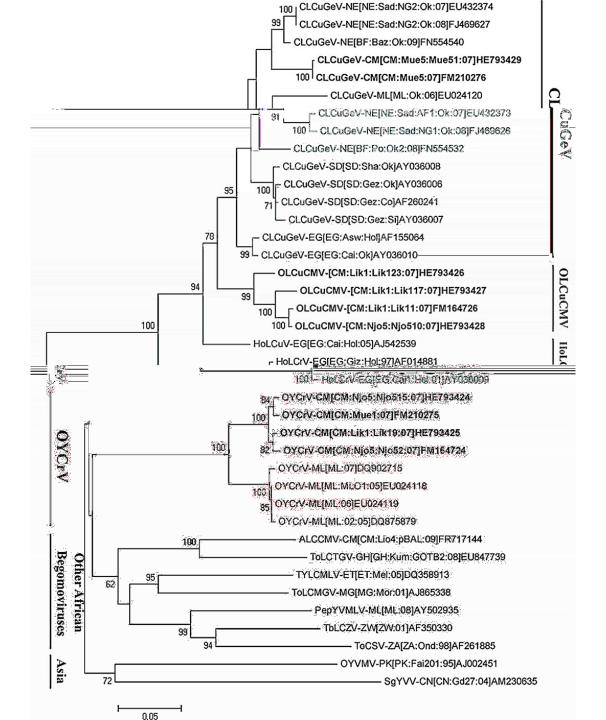


- Spread of diverse species to cotton-growing regions and widespread losses could create a global economic crisis and shortage of supply
- CLCuMV and satellite already found in China-moved on ornamental plants
- Invasive whitefly biotypes, also Re-distributed via trade
- Further threat due to <u>high diversity</u> / <u>broad host range</u>

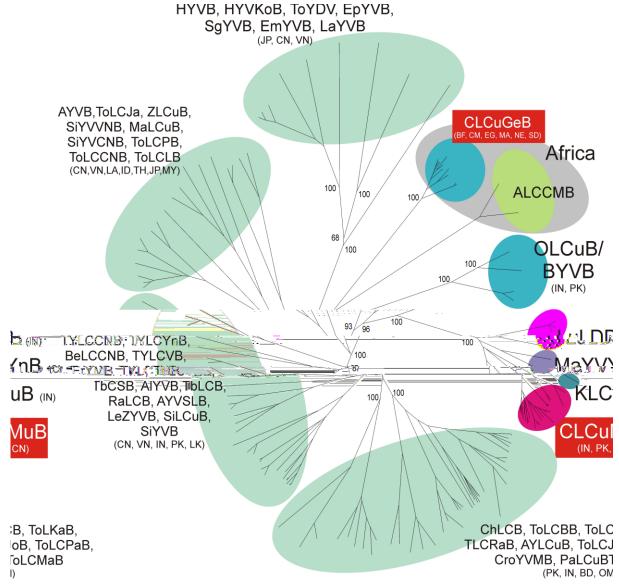


Thank you

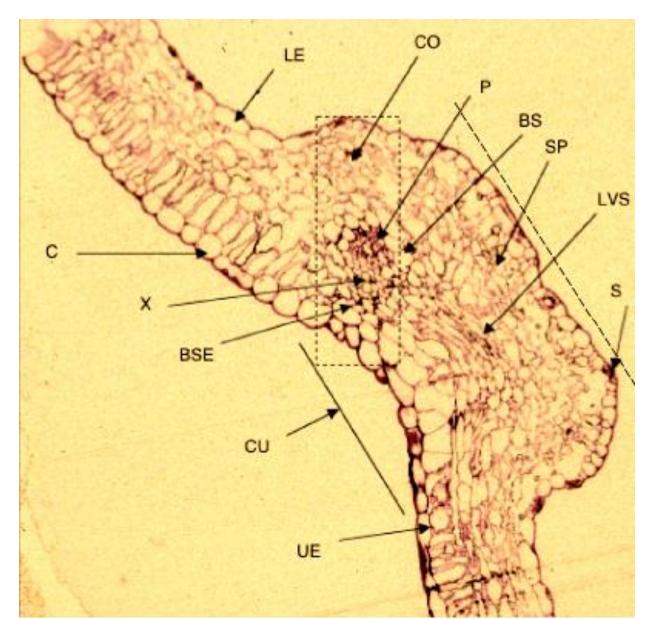
Phylogeny of OW begomoviruses infecting cotton, hollyhock, and okra



#### Alphasatellites



# Enations (box) developing on a <u>main vein</u> of cotton leaf inoculated with CLCuGV and sat DNA - 'vein thickening'



Swelling of veins results from proliferation of spongy parenchyma and bundle sheath cells, with SP being more swollen than bundle sheath cells.

This asymmetric proliferation of veins causes the formation of veinal grooves.

Transverse sections of cells associated with <u>enations</u> that form on CLCuGV-infected leaves contain parenchyma instead of collenchyma with very thin cell walls.