

References

- Ikuta KS, Swetschinski LR, Robles Aguilar G, Sharara F, Mestrovic T, Gray AP, et al.; GBD 2019 Antimicrobial Resistance Collaborators. Global mortality associated with 33 bacterial pathogens in 2019: a systematic analysis for the Global Burden of Disease Study 2019. *Lancet*. 2022;400:2221–48. [https://doi.org/10.1016/S0140-6736\(22\)02185-7](https://doi.org/10.1016/S0140-6736(22)02185-7)
- Katz TS, Harhay DM, Schmidt JW, Wheeler TL. Identifying a list of *Salmonella* serotypes of concern to target for reducing risk of salmonellosis. *Front Microbiol*. 2024; 15:1307563. <https://doi.org/10.3389/fmicb.2024.1307563>
- European Food Safety Authority (EFSA); European Centre for Disease Prevention and Control (ECDC). The European Union One Health 2022 zoonoses report. *EFSA J*. 2023;21:e8442. <https://doi.org/10.2903/j.efsa.2023.8442>
- World Data Centre for Microorganisms. *Salmonella enterica* serovar Abony WDCM 00029 [cited 2024 Jun 7]. https://refs.wdcm.org/browse/getinfo?sid=WDCM_00029
- American Type Culture Collection. *Salmonella enterica* serovar Abony ATCC BAA-2162 [cited 2024 Jun 7]. <https://www.atcc.org/products/baa-2162>
- Monte DFM, Nethery MA, Barrangou R, Landgraf M, Fedorka-Cray PJ. Whole-genome sequencing analysis and CRISPR genotyping of rare antibiotic-resistant *Salmonella enterica* serovars isolated from food and related sources. *Food Microbiol*. 2021;93:103601. <https://doi.org/10.1016/j.fm.2020.103601>
- Akinyemi KO, Fakorede CO, Linde J, Methner U, Wareth G, Tomaso H, et al. Whole genome sequencing of *Salmonella enterica* serovars isolated from humans, animals, and the environment in Lagos, Nigeria. *BMC Microbiol*. 2023;23:164. <https://doi.org/10.1186/s12866-023-02901-1>
- National Collection of Type Cultures. *Salmonella enterica* serovar Abony NCTC 6017 [cited 2024 Jun 7]. <https://www.culturecollections.org.uk/nop/product/salmonella-enterica-subsp-enterica-serotype-abony>
- International Council for Harmonization. Q4B evaluation and recommendation of pharmacopoeial texts for use in the ICH regions – annex 4B(R1) microbiological examination of nonsterile products: tests for specified microorganisms general chapter [cited 2024 Jun 7]. <https://www.fda.gov/media/71267/download>
- US Pharmacopoeia. <62> Microbiological examination of nonsterile products: tests for specified microorganisms [cited 2024 Jun 7]. https://www.usp.org/sites/default/files/usp/document/harmonization/gen-method/q05a_pf_ira_34_6_2008.pdf

Address for correspondence: Andrea I. Moreno-Switt, Pontificia Universidad Católica de Chile, Av. Vicuña Mackenna 4860, Santiago 7820436, Chile; email: andrea.moreno@uc.cl

Endogenous Endophthalmitis Caused by *Prototheca microalga* in Birman Cat, Spain

Laura Jimenez-Ramos,¹ Ana Ripolles-Garcia,¹ Gianvito Lanave, Francesco Pellegrini, Miriam Caro-Suarez, Almudena Latre-Moreno, Marta Ferruz-Fernandez, Maria Luisa Palmero-Colado, Vanessa Carballes-Perez, Antonio Melendez-Lazo, Carolina Naranjo, Fernando Laguna, Vito Martella, Manuel Villagrasa

Author affiliations: Puchol Veterinary Hospital, Madrid, Spain (L. Jimenez-Ramos, A. Ripolles-Garcia, M. Caro-Suarez, A. Latre-Moreno, M. Ferruz-Fernandez, F. Laguna, M. Villagrasa); Centro Oftalmológico Veterinario Goya, Madrid (L. Jimenez-Ramos, A. Ripolles-Garcia, M. Caro-Suarez, A. Latre-Moreno, M. Ferruz-Fernandez, F. Laguna, M. Villagrasa); University of Bari Aldo Moro, Bari, Italy (G. Lanave, F. Pellegrini, V. Martella); Gattos Veterinary Hospital, Madrid (M.L. Luisa Palmero-Colado, V. Carballes-Perez); T-Cito Laboratories, Barcelona, Spain (A. Melendez-Lazo); IDEXX Laboratories, Barcelona (C. Naranjo); University of Veterinary Medicine, Budapest, Hungary (V. Martella)

DOI: <https://doi.org/10.3201/eid3101.241198>

We identified *Prototheca* spp. microalga in ocular samples of a cat in Spain with nontreatable endogenous endophthalmitis. Within 2 years, the eye lesions progressively worsened and neurologic signs appeared, suggesting systemic spread of the infection. On multitarget sequence analysis, the feline pathogen could not be assigned to any known *Prototheca* species.

Protothecosis is an uncommon disease caused by the unicellular microalga *Prototheca* spp., described in humans and animals and associated with systemic disease, cutaneous lesions, or both (1,2). *Prototheca* spp. has been identified as the cause of cutaneous lesions and in 1 case of disseminated neurologic disease in cats (2–4). Diagnosis of protothecosis can be challenging and usually is based on observation of the organism in tissues and body fluids (5). Culturing or PCR is required for a definitive diagnosis and species identification (2,4).

A 5-year-old female Birman cat, spayed and maintained indoors, was referred to our veterinary hospital for a 1.5-month history of uveitis in the right eye. Neuro-ophthalmic evaluation revealed that the right eye was blind and had severe signs of uveitis,

¹These first authors contributed equally to this article.

whereas the left eye was unaffected. Ultrasound examination showed exudative retinal detachment in the right eye, confirming irreversible blindness. At 5.5 months, clinical signs of uveitis appeared also in the other eye (Figure, panel A). At the 6.5-month follow-up, the aqueous flare in the left eye worsened (Figure, panel B). We obtained an aqueous humor sample and a vitreous sample for cytologic examination, which revealed a mixed inflammatory process and the presence in the vitreous of structures morphologically compatible with algae of the genus *Prototheca* spp. (Appendix Figure 1, panel A, <https://wwwnc.cdc.gov/EID/article/31/1/24-1198-App1.pdf>). Antifungal therapy with itraconazole (5 mg/kg 2×/d) was initiated at 10.5 months and stopped voluntarily by the owner at 14.5 months. At the 16.5-month follow-up, the cat was completely blind, and the clinical signs had worsened with the development of corneal macro-deposits (Figure, panels C, D). At 17.5 months,

we observed 2 episodes of neurologic clinical signs, including vestibular signs, ataxia, and disorientation. At 19.5 months, the owner reported that the cat had lost her hearing. At 21.5 months, the cat's neurologic status further deteriorated, with the onset of seizures and prolonged anorexia. At this point, the owner opted for the humane euthanasia of the cat but did not give consent for a full-body necropsy.

The eyes of the cat were submitted for biopsy. The samples had been frozen before fixation and autolysis had occurred, so histopathologic investigations were challenging because of artifacts. Nevertheless, diffuse exudate was visible throughout all the ocular structures (Appendix Figure 1, panel B). We observed karyorrhectic remnants and microorganisms within the axial cornea (Appendix Figure 1, panel C). In addition, the lens capsule was ruptured, and we noted the presence of intra-lenticular microorganisms and hypermature cataract formation (Appendix Figure

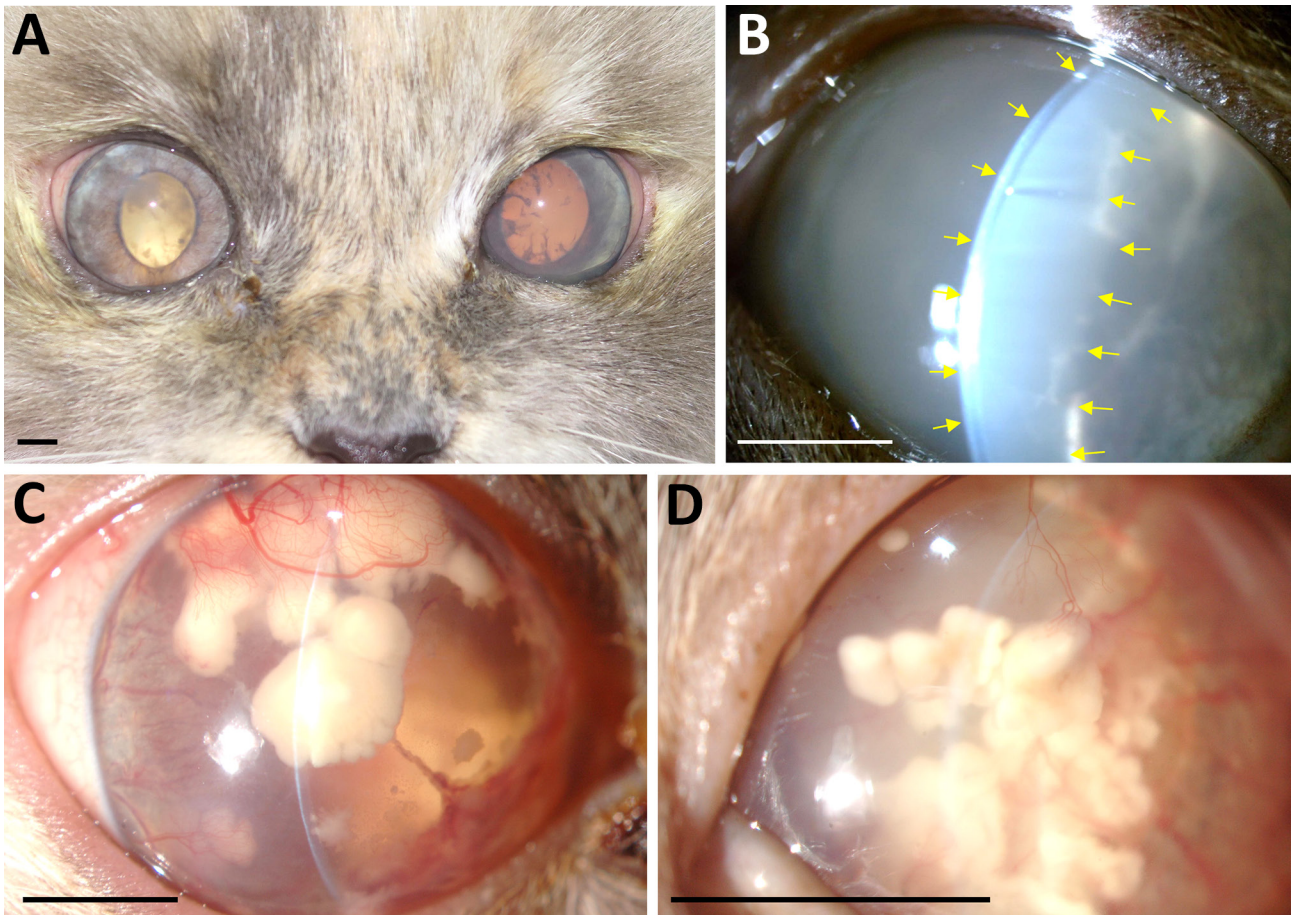


Figure. Clinical course of bilateral endogenous endophthalmitis in 5-year-old female Birman cat evaluated by slit-lamp biomicroscopic examination, Madrid, Spain. A) Digital photograph of both eyes demonstrating bilateral uveitis at 5.5 months after initial visit to clinic. B) Slit-lamp biomicroscopic image (original magnification $\times 10$) of the left eye, demonstrating a marked flare (yellow arrows) at 6.5 months after initial clinical signs. C, D) At 16.5 months, the right eye (C) (original magnification $\times 10$) and left eye (D) (original magnification $\times 16$) were imaged by slit-lamp biomicroscopic examination, revealing corneal endothelial macrodeposits of undefined origin, presumed to be the result of *Prototheca* spp. invasion. Scale bars indicate 5 mm.

1, panel D). The microorganisms exhibited a markedly periodic acid-Schiff–positive and Alcian blue–negative membrane. Results of PCR analysis for *Cryptococcus* spp. were negative and, on the basis of the morphology and staining characteristics, we suspected a diagnosis of *Prototheca* spp. infection.

The ocular samples tested positive for *Prototheca* spp. in PCR tests that used 3 primer sets (Appendix Table 1) and amplified a 1,800-bp sequence of the 18S rDNA, a 630-bp sequence of 28S rDNA, and a 650-bp sequence of the cytochrome B gene. We deposited the nucleotide sequences in GenBank (accession nos. PQ111814 [18S rDNA sequence], PQ122806 [28S rDNA sequence], and PQ115153 [cytochrome B gene sequence]). We conducted multitarget sequence and phylogenetic analysis by using the sequences generated in this study and cognate sequences retrieved from the National Center for Biotechnology Information database (Appendix Figure 2). The 18S rDNA, the 28S rDNA, and the cytochrome B gene sequences shared the highest nucleotide identity with *Prototheca lentescens* PK1 (GenBank accession nos. MZ198751 [86.0%], OK236514 [84.8%], and MW701399 [83.5%]) (Appendix Table 2). The feline *Prototheca* strain was segregated in a separate branch within the maximum-likelihood phylogenetic tree, diverging from other *Prototheca* species, thereby supporting the hypothesis of a distinct phylotaxonomic status for the strain SPA/2024/cat/259 (Appendix Figure 2).

Disseminated *Prototheca* infection already has been reported in a cat with central nervous system signs and a suspected diagnosis of multifocal lymphoma; however, in other reports, feline protothecosis has been associated with cutaneous or subcutaneous lesions (2–4). In our case, the cat had a history of chronic glucocorticoid administration for intestinal disease, which probably triggered immune suppression in the animal. In addition, the cat had received 2 fecal transplants, which might be a potential source of infection. Also, previous studies have indicated that the Birman breed is highly susceptible to certain infectious diseases, including chlamydia, cryptococcosis, feline infectious peritonitis, and *Tritrichomonas fetus* infection (6–9). However, we could not identify the primary source of the infection because this microalga can be found in multiple environmental sources. Another limitation of our study was that we could not isolate the *Prototheca* strain in vitro to assess its cultural properties.

In conclusion, we describe a novel candidate *Prototheca* species invading the ocular tissues of a cat, a rare clinical manifestation in felids. Our findings also extend the knowledge of the genetic diversity of

Prototheca spp. in animals, a piece of valuable information for pathogens with zoonotic potential.

Acknowledgments

The authors acknowledge the invaluable technical assistance provided by the staff of the Puchol Veterinary Hospital. In particular, Erica Cumpa, Patricia Leal, and Maria Peralta are to be commended for their role in coordinating the procedures and their exemplary care for the animals. Furthermore, we are grateful to Maria Soto for her invaluable assistance with sample management and study coordination.

This study was approved by the Ethics and Professional Integrity Committee of the College of Veterinary Surgeons of Madrid.

This research was funded in part by the National Laboratory for Infectious Animal Diseases, Antimicrobial Resistance, Veterinary Public Health and Food Chain Safety (grant no. RRF-2.3.1-21-2022-00001).

About the Author

Dr. Jimenez-Ramos is a veterinary ophthalmologist at Puchol Veterinary Hospital in Madrid, Spain.

Dr. Ripolles-Garcia also is a veterinary ophthalmologist at Puchol Veterinary Hospital. Her primary research interests are retinal diseases and ocular manifestations of systemic infections.

References

- Ahrholdt J, Murugaiyan J, Straubinger RK, Jagielski T, Roesler U. Epidemiological analysis of worldwide bovine, canine and human clinical *Prototheca* isolates by PCR genotyping and MALDI-TOF mass spectrometry proteomic phenotyping. *Med Mycol*. 2012;50:234–43. <https://doi.org/10.3109/13693786.2011.597445>
- Sykes JE. Protothecosis and chlorellosis [chapter 90]. In: Sykes JE, editor. *Greene's infectious diseases of the dog and cat*, 5th edition. St. Louis: Elsevier, Inc.; 2023. p. 1126–34.
- Ely VL, Espindola JP, Barasuol BM, Sangioni LA, Pereira DB, Botton SA. Protothecosis in veterinary medicine: a minireview. *Lett Appl Microbiol*. 2023;76:ovad066. <https://doi.org/10.1093/lambio/ovad066>
- Lanave G, Pellegrini F, Palermo G, Zini E, Mercuriali E, Zagarella P, et al. Identification of *Prototheca* from the cerebrospinal fluid of a cat with neurological signs. *Vet Sci*. 2023;10:681. <https://doi.org/10.3390/vetsci10120681>
- Webb AA, Cullen CL. Ocular manifestations of systemic disease, part 1: the dog [chapter 37.1]. In: Gelatt KN, editor. *Veterinary ophthalmology*, 6th edition. Hoboken (NJ): John Wiley & Sons; 2021. p. 2329–420.
- Wills J, Howard P, Gruffydd-Jones T, Wathes CM. Prevalence of *Chlamydia psittaci* in different cat populations in Britain. *J Small Anim Pract*. 1988;29:327–39. <https://doi.org/10.1111/j.1748-5827.1988.tb02293.x>
- Foley JE, Pedersen NC. The inheritance of susceptibility to feline infectious peritonitis in purebred catteries. *Feline Pract*. 1996;24:14–22.

8. Golovko L, Lyons LA, Liu H, Sørensen A, Wehnert S, Pedersen NC. Genetic susceptibility to feline infectious peritonitis in Birman cats. *Virus Res.* 2013;175:58–63. <https://doi.org/10.1016/j.virusres.2013.04.006>
9. Kuehner KA, Marks SL, Kass PH, Sauter-Louis C, Grahn RA, Barutzki D, et al. *Trichomonas foetus* infection in purebred cats in Germany: prevalence of clinical signs and the role of co-infection with other enteroparasites. *J Feline Med Surg.* 2011;13:251–8. <https://doi.org/10.1016/j.jfms.2010.12.002>

Address for correspondence: Fernando Laguna, Department of Ophthalmology, Puchol Veterinary Hospital, 8th Saucedá St, 28050, Madrid, Spain; email: ferlagu@gmail.com

Spread of Antifungal-Resistant *Trichophyton indotineae*, United Kingdom, 2017–2024

Alireza Abdolrasouli, Richard C. Barton, Andrew M. Borman

Author affiliations: King's College Hospital, London, UK (A. Abdolrasouli); Leeds Teaching Hospitals National Health Service Trust, Leeds, UK (R.C. Barton); Southmead Hospital, Bristol, UK (A.M. Borman); University of Exeter, Exeter, UK (A.M. Borman)

DOI: <https://doi.org/10.3201/eid3101.240923>

We describe 157 cases of *Trichophyton indotineae* infection in the United Kingdom, mostly in patients linked to southern Asia. *T. indotineae* is spreading in the United Kingdom and accounts for 38% of dermatophyte isolates referred to the UK National Mycology Reference Laboratory. Clinicians should suspect *T. indotineae* in tinea corporis cases.

Outbreaks of superficial skin infections caused by the emergent dermatophyte *Trichophyton indotineae* (*Trichophyton mentagrophytes* genotype VIII) were reported in southern Asia starting in 2014 (1–4). Typically, *T. indotineae* infections initially involve the groin (tinea cruris) and respond poorly to treatment, resulting in widespread lesions affecting multiple body sites. Many isolates exhibit in vitro

resistance to terbinafine, and most infections are clinically resistant to that drug (1–5). Infections spread easily from person to person (1–8), and some reports suggest sexual transmission (9).

T. indotineae is endemic across Asia, but cases have been reported worldwide (4), including in Europe (5–7), Canada (8), and the United States (9). Mounting evidence suggests infection acquisition and transmission outside original areas of endemicity (5,7,9,10). Occasional cases of *T. indotineae* infection have been reported from the United Kingdom (10). We describe all cases of *T. indotineae* identified at the UK National Mycology Reference Laboratory (MRL) during a 7-year period.

We reviewed laboratory records from August 2017–July 2024 for dermatophytes identified as *T. indotineae*. When available, we extracted clinical and epidemiologic data from requisition forms. Dermatophyte identification was determined by whole-genome sequencing (WGS) or internal transcribed spacer sequencing, combined with phenotypic identification (Appendix Table, <https://wwwnc.cdc.gov/EID/article/31/1/24-0923-App1.pdf>). Isolates received after 2021 were identified using phenotypic features alone. A key defining microscopic feature was abundant fusiform to clavate, thin smooth-walled macroconidia with an acute apical tip, as well as other macroscopic and microscopic characteristics (Appendix Figure 1). We performed susceptibility testing by broth microdilution according to Clinical and Laboratory Standards Institute standards (Appendix). In the absence of an established clinical breakpoint for terbinafine, we used an MIC of ≥ 0.5 mg/L to identify non-wild-type isolates.

The first WGS-confirmed case we noted was from October 2018. In nearly half (42.7%, 67/157) of identified cases, the groin, buttocks, and thighs were directly involved, and neighboring body sites (abdomen and back) were implicated in another 18 cases (Table 1). Most (84.7%) patients had links to endemic areas, including South Asian ethnic background (n = 97), recent travel to the Indian subcontinent or Middle East (n = 41), or both (n = 36). Household spread was noted in 5 cases (Appendix Table).

Before 2023, most (27/36) cases were identified in London, which was the most affected city according to total case numbers. Since 2023, increasing numbers of cases were found in an additional 27 cities in the United Kingdom and Ireland, and isolate numbers outside London exceed those in London (Appendix Figure 3). From 2018 to 2019, the prevalence of *T. indotineae* in the United Kingdom increased from 2% to