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*Supplement of*

## **Oceanic CO<sub>2</sub> outgassing and biological production hotspots induced by pre-industrial river loads of nutrients and carbon in a global modeling approach**

**Fabrice Lacroix et al.**

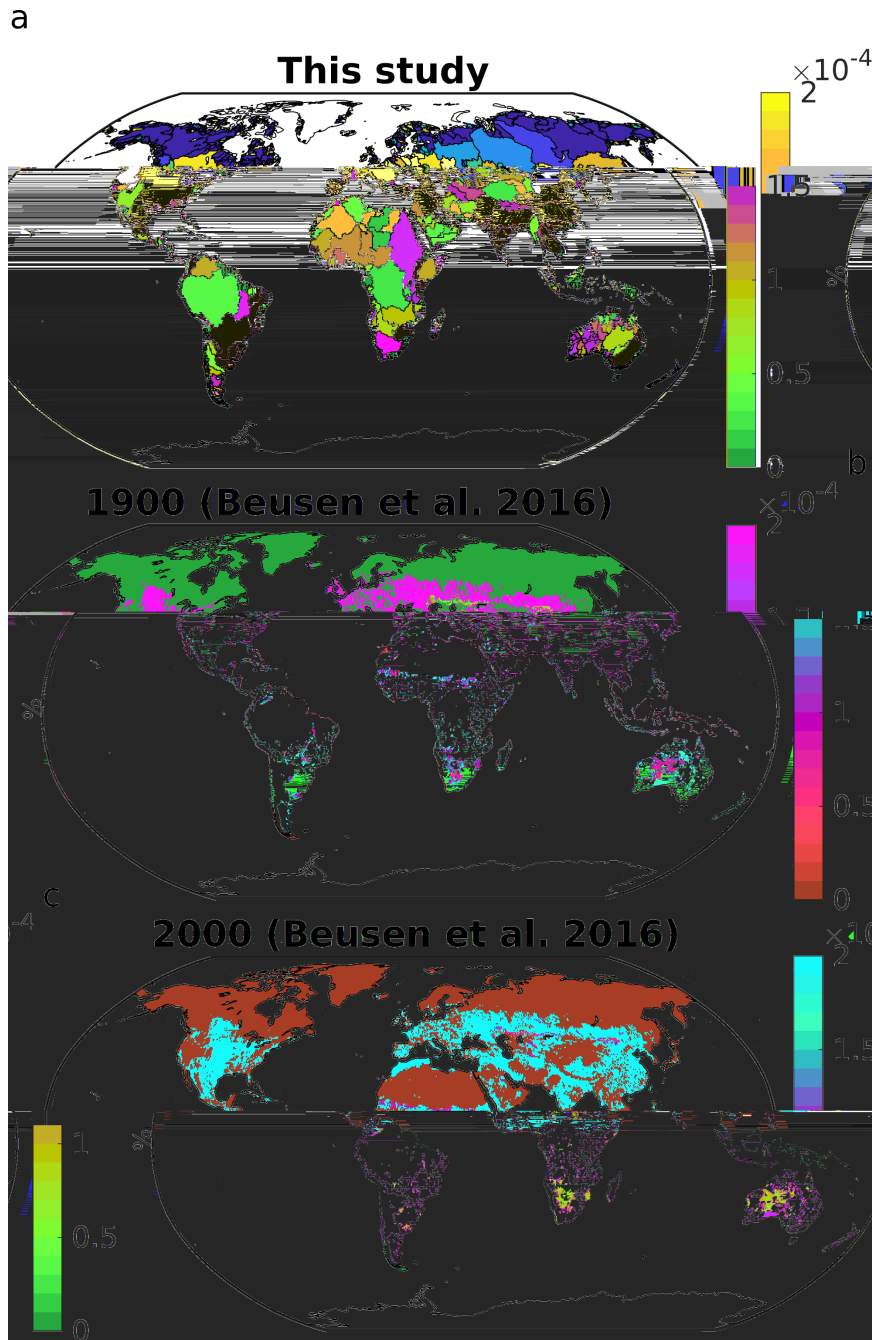
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## **S.1 Methods: Inputs to catchments**

### **S.1.1 Anthropogenic P inputs**

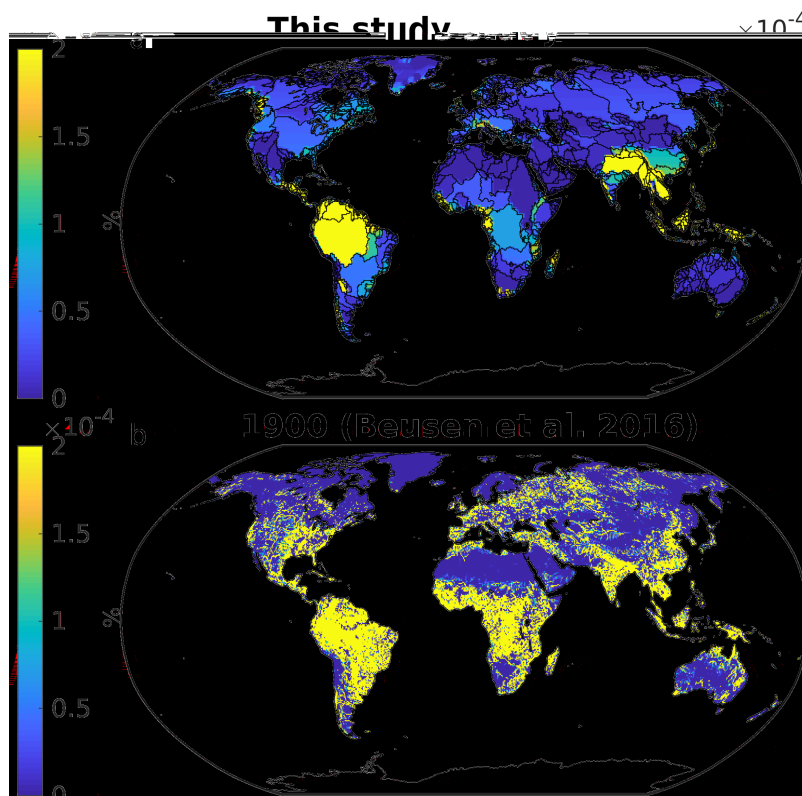
In this study, we assumed anthropogenic inputs from agricultural, sewage and allochthonous P sources. The anthropogenic sources were distributed according to anthropogenic exports of present-day NEWS2 DIP exports. To verify that this assumption is plausible, we compared our assumed distribution (Figure 1a) to the distributions of anthropogenic P inputs to catchments of the modelling study of Beusen et al. (2016), which account for 1900 (Figure 1b) to the year 2000 (Figure 1c). The results show that the present-day anthropogenic DIP distributions assumed in our study strongly agree with both the Beusen et al. (2016) for year 1900 and 2000. We furthermore observe very similar distributions of anthropogenic inputs to catchments for year 1900 and 2000 in the Beusen et al. (2016) study, although increasing contributions to P inputs in Southeast Asia can be observed. Nevertheless, we acknowledge that pre-industrial anthropogenic exports are likely strongly uncertain in their magnitudes as well as in their distributions due to the lack of data.



**Figure 1.** Relative contributions of anthropogenic P inputs to catchments [ $10^{-4}$  %] as assumed in our study (a), as well as for year 1900 (b) and 2000 as assumed in Beusen et al. (2016). This was calculated for every grid cell of a regular 720x360 grid.

### S.1.2 Allochthonous P inputs

The distributions of allochthonous P riverine exports was assumed to be the same as organic matter riverine exports, which were derived from the NEWS2 study. This was done primarily in order to avoid having strongly differing organic matter exports to total P exports, which would be problematic in our fractionation assumptions for P. For instance, if organic matter exports were very high and P inputs were very low, all P would be assumed to be transformed to organic matter, leaving DIP exports close to null. Comparing our assumed distribution with the distribution from Beusen et al. (2016) however reveals a strong level of agreement, signaling that the allochthonous inputs to catchments might be strongly correlated to net organic matter production within the catchments.



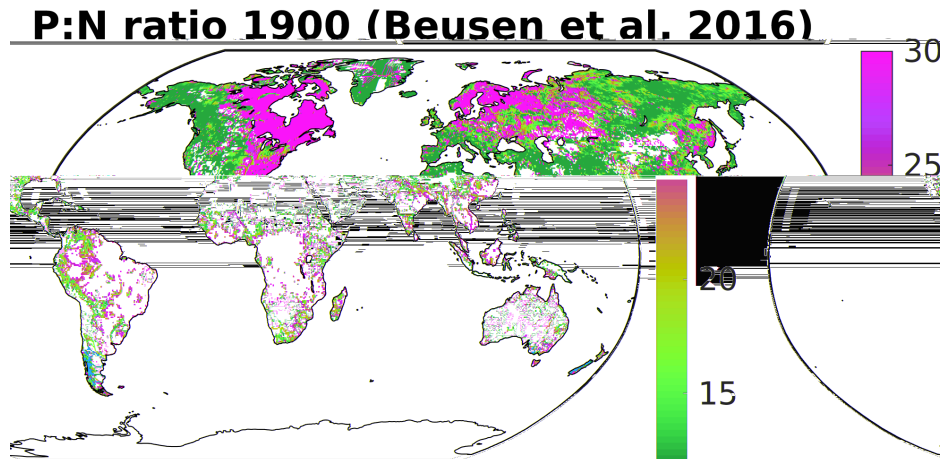
**Figure 2.** Relative contributions of allochthonous inputs to catchments as assumed in our study (a) and of 1900 inputs assumed in Beusen et al. (2016). This was calculated for every grid cell of a regular 720x360 grid.

### S.1.3 N:P ratios

10 In this study, we assume riverine N:P mole ratios of 16:1 for dissolved inorganic as well as organic species. In literature, higher values for this ratio can be found. For one, Beusen et al. (2016) simulate a mole N:P ratio for total exports to the ocean of around 21:1 both for 1900 as well as for present-day. Meybeck (1982) suggest a DIN:DIP mole ratio of 26:1 based on observational data from major rivers (contemporary) and Seitzinger et al. (2010) model a DIN:DIP ratio of present-day ratio of 29:1. DIN has however been reported to have undergone stronger increases than DIP, at least since 1970 (Seitzinger et al., 2010). Seitzinger et al. (2010) simulate a global DON:DOP mole ratio of 39:1. Meybeck (1982) also suggest a total organic N:P export mole ratio of around 9:1, albeit with acknowledgment of relatively little observational data.

While higher N:P values are reported in literature, denitrification in estuaries and in the coastal ocean likely compensate for the higher N:P ratio by eliminating DIN, with estimations of the denitrification flux being around 3-10 Tg N yr<sup>-1</sup>.

5 The total additional riverine N export (7 Tg N yr<sup>-1</sup>) when using for instance a ratio of 21:1 (Beusen et al., 2016) could therefore be completely offset by the loss of nitrogen in estuaries. When accounting for denitrification in estuaries, the difference in the global N export would be small (-14-10 %). Regionally however, differing N:P ratios could have strong impacts, especially when considering the anthropogenic of riverine exports. For instance, Beusen et al. (2016) show much higher N:P ratios on the South American continent, eastern North America, Northern Europe and Southeast Asia.



**Figure 3.** N:P ratio catchment inputs modelled in Beusen et al. (2016).

## S.2 Methods: HAMOCC Model parameters

### S.2.1 Comparison with standard parameters

**Table 1.** Comparison of main model parameters with Ilyina et al. (2013). Model equations can be found in Ilyina et al. (2013), as well as Paulsen et al. (2017) for the implementation of cyanobacteria in the model.

Symbol	Variable	This study	Ilyina et al., 2013	Units
<b>Phytoplankton</b>				
$\alpha_{\text{phy}}$	Initial slope of the photosynthesis versus the irradiance curve	0.03	0.03	$\text{m}^2 \text{W}^{-1} \text{d}^{-1}$
$\mu_{\text{phy}}$	Maximum growth rate	0.6	0.6	$\text{d}^{-1}$
$\text{Phy}_{\text{min}}$	Minimum Concentration of phytoplankton	1E-11	1E-11	$\text{kmol P m}^{-3}$
$\lambda_{\text{phy}}$	Mortality rate	0.008	0.008	$\text{d}^{-1}$
$\beta_{\text{phy}}$	Exudation rate	0.03	0.03	$\text{d}^{-1}$
<b>Cyanobacteria</b>				
$\alpha_{\text{cya}}$	Initial slope of the photosynthesis versus the irradiance curve	0.02	-	$\text{m}^2 \text{W}^{-1} \text{d}^{-1}$
$\mu_{\text{cya}}$	Maximum growth rate	0.2	-	$\text{d}^{-1}$
$T_{\text{opt}}$	Optimal growth temperature	28	-	$^{\circ}\text{C}$
$T_1, T_2$	Temperature distribution parameters	5.5, 1	-	$^{\circ}\text{C}$
$\text{Cya}_{\text{min}}$	Minimum Concentration of cyanobacteria	1E-11	-	$\text{kmol P m}^{-3}$
$\lambda_{\text{phy}}$	Mortality rate	0.1	-	$\text{d}^{-1}$
$\beta_{\text{cya}}$	Exudation rate	0.01	-	$\text{d}^{-1}$
<b>Zooplankton</b>				
$\mu_{\text{Zoo}}$	Max. Grazing Rate	1	1	$\text{d}^{-1}$
$K_{\text{Zoo}}$	Half-saturation constant for grazing	4E-8	4E-8	$\text{kmol P m}^{-3}$
$\text{Zoo}_{\text{min}}$	Min. concentration of zooplankton	1E-11	1E-11	$\text{kmol P m}^{-3}$
$1-\epsilon_{\text{Zoo}}$	Fraction of grazing egested	0.2	0.2	-
$\lambda_{\text{Zoo}}$	Mortality rate	3E6	3E6	$\text{d}^{-1}$
$\beta_{\text{Zoo}}$	Excretion rate	0.06	0.06	$\text{d}^{-1}$
$1-\epsilon_{\text{can}}$	Fraction of carnivores grazing egested	0.05	0.05	-
<b>Nutrients</b>				
$K_{\text{PO}_4}$	Half-saturation constant for DIP uptake	4E-8	4E-8	$\text{kmol P m}^{-3}$
$K_{\text{NO}_3}$	Half-saturation constant for DIN uptake	1.6E-7	1.6E-7	$\text{kmol N m}^{-3}$
$K_{\text{DFe}}$	Half-saturation constant for DFe uptake	0.0036	1.6E-7	$\text{kmol N m}^{-3}$
$R_{\text{dust}}$	Ratio of bioavailable iron in dust	6.26E-6	6.26E-6	$\text{kmol Fe kg}^{-1}$
$\text{Fe}_{\text{crit}}$	Critical Fe concentration of complexation	5E-10	6E-10	$\text{kmol m}^{-3}$

Symbol	Variable	This study	Ilyina et al., 2013	Units
<b>Particulate organic matter (POM)</b>				
$w_{det}$	Sinking speed of POM	5	5	$d^{-1}$
$O_{2crit}$	Aerob threshold	5E-8	5E-8	$kmol\ O_2\ m^{-3}$
$\lambda_{det}$	Remineralization rate	0.026	0.025	$d^{-1}$
$\lambda_N$	Denitrification rate to $N_2O$	0.01	0.01	$d^{-1}$
$R_{denit}$	Ratio of DIN consumed in denitrification	137.6	137.6	$mol\ N\ (mol\ P)^{-1}$
$\lambda_N$	Denitrification rate to N	0.005	0.005	$d^{-1}$
$R_{denit}$	Ratio of $N_2O$ consumed in denitrification to N	344	344	$mol\ N\ (mol\ P_2)^{-1}$
$R_{nit}$	DIN produced during nitrification	1E-4	1E-4	$mol\ N\ mol\ O_2^{-1}$
<b>Dissolved organic matter (DOM)</b>				
$\lambda_{DOM}$	(Bacterial) remineralization rate	0.008	0.004	$d^{-1}$
<b>Shell material</b>				
$K_{Si}$	Half saturation constant for $Si(OH)_4$ uptake	1E-6	1E-6	$kmol\ Si\ m^{-3}$
$R_{Si:P}$	DSi:P uptake ratio during opal production	25	25	$mol\ Si\ (mol\ P)^{-1}$
$R_{Ca:P}$	DIC:P uptake ratio during $CaCO_3$ production	35	20	$mol\ C\ (mol\ P)^{-1}$
$w_{opal}$	Sinking speed of opal	30	30	$d^{-1}$
$w_{calc}$	Sinking speed of $CaCO_3$	30	30	$d^{-1}$
$\lambda_{opal}$	Opal dissolution rate	0.01	0.01	$d^{-1}$
$\lambda_{calc}$	$CaCO_3$ dissolution rate	0.075	0.075	$d^{-1}$

### S.2.2 tDOM degradation validation

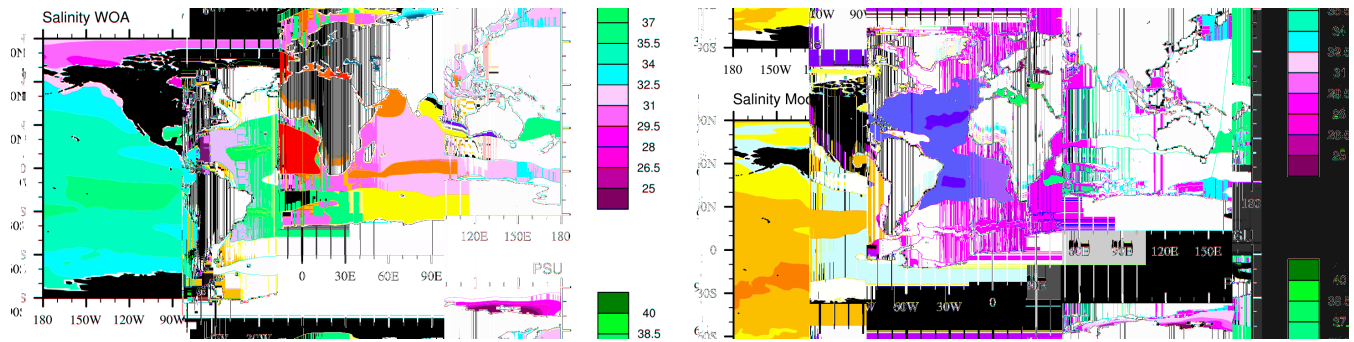
At a tDOM degradation rate of  $0.003 \text{ d}^{-1}$ ,  $0.39 \text{ Tg C yr}^{-1}$  of riverine tDOM is remineralized on the Louisiana Shelf, whereas  
5  $0.68$  is suggested to be degraded on the shelf in Fichot and Benner (2014).  $2.09 \text{ Tg C yr}^{-1}$  is simulated to be degraded in the  
Baltic Sea whereas  $1.75 \text{ Tg C yr}^{-1}$  is budgeted in Seidel et al. (2017).

The surface tDOM concentrations in the Pacific contribute to less than 1 % of total DOM concentrations in the entire basin,  
which is consistent with Lignin measurements in the surface ocean of the North Pacific that suggest tDOM contributions of  
around 1% to total DOM Hernes and Benner (2002). In the North Atlantic, the contribution of tDOM to the total DOM is  
10 suggested to be higher (around 2.4 %, Opsahl and Benner (1997)), which is also reflected in the model with contributions even  
exceeding 5% in some areas of the open ocean.



### S.3 Analysis: Freshwater inputs to the ocean

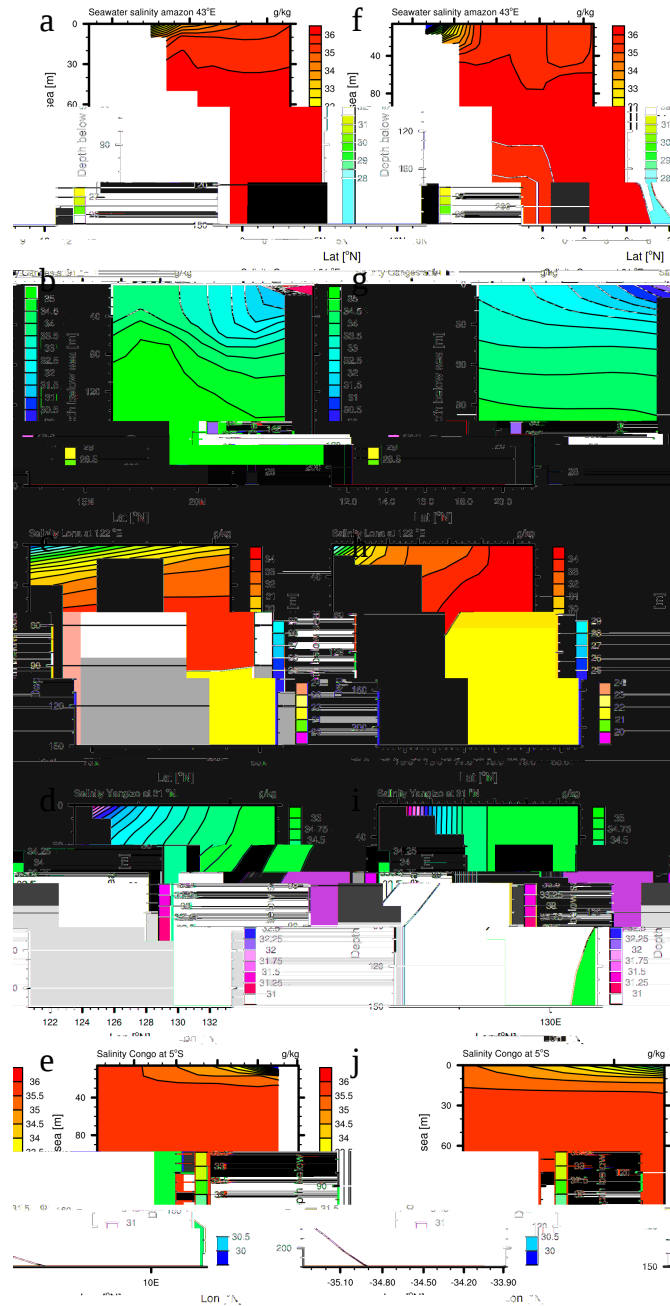
The freshwater loads from OMIP were already implemented in previous standard HAMOCC version, in order to close the hydrological cycle. Globally, they add  $32,542 \text{ km}^3 \text{ yr}^{-1}$  of freshwater to the ocean. These impact the salinity and the ocean stratification in the proximity of our biogeochemical riverine inputs, and therefore their advection within the ocean.



**Figure 4.** Comparison of salinity from observational data (WOA) and modelled salinity in RIV (Salinity Model).

#### **S.4 Analysis: Coastal Profiles**

Coastal vertical profiles show that despite the coarse resolution of our model, the features of salinity in selected coastal regions is comparable to WOA data, although vertical gradients are often not as strong as in the observational data (Figure A.2). For the Amazon Northward section, we see a vertical the salinity gradient in the WOA data, which is also reproduced albeit not extensively in the modelled data which induces stronger stratification of the water layers. In terms of absolute values the salinity is also well represented for the Ganges river, although the vertical stratification is also not quite as extensive further away from the coast. The bathymetry of the Laptev Sea at the Lena river mouth is poorly represented, as WOA data shows a height increase of the ocean floor at connection of the Sea with the Arctic Ocean, which is not represented in the model bathymetry. The salinity gradient in the East Chinese Sea is very strong due to currents parallel in the model which are also shown in observations (Ichikawa and Beardsley, 2002). Smaller currents in this region and in the Yellow Sea are however not well represented in the model. The Congo river has a very steep coastal shelf, and therefore the salinity relatively strong vertical horizontal gradient, due to little bathymetry induced vertical mixing, as well as heating of the surface waters.



**Figure 5.** Vertical profiles of the coastal bathymetry and salinity, along specific longitudes and longitudes for chosen river mouths. The left column (a-e) is made of WOA salinity profiles, whereas the right column (f-j) of profiles from model simulation RIV. a & f are shelf and salinity profiles at the Amazon river (TWA), b & g for the Ganges river, c & h Lena river, d & i Yangtze river, e & j Congo river.

## S.5 Analysis: Magnitudes of inputs to the open and coastal ocean in REF

In order to assess the impacts of open ocean biogeochemical inputs (as in the standard model simulation REF) on the dissolved inorganic nutrient concentrations, we calculated the P inputs to the surface layers and compared them to the surface ocean (first 12 meters) inventories of 3 regions of the open ocean in the model, as well as for the global coastal ocean (depths of less than 250m). In the productive regions of the equatorial Pacific and in the Southern Ocean, the P inputs are very small in comparison to the surface P inventories (Figure 2). This signals that the inputs are not strong contributors to maintaining the DIP concentrations here. In the tropical Atlantic, the inputs might cause a bias by artificially increasing the concentrations. We however only calculate the inventories in the first 12m inventories here, although primary can take place until up to around 100m in the model. Therefore, the inputs are still likely very small compared to the nutrient inventories that impact the NPP. Throughout our study, we also show that the ocean physics is a much stronger contributor to open ocean biogeochemical distributions than the locations of the biogeochemical inputs. For instance, the difference between RIV and REF in the open ocean NPP is relatively small, with the major features being conserved. Considering riverine inputs however impacts certain coastal regions substantially and could affect certain regions of the open ocean (tropical Atlantic).

The inputs to the coastal ocean are very small in REF (6%), meaning that the vast majority of inputs in the simulation REF happens in the open ocean. In the simulation RIV however, 100% of the inputs are added to the coastal ocean at the river mouths. Therefore, our simulations allow a comparison between an ocean state where inputs are added to the open ocean (REF), and one where inputs are added at their correct locations in the coastal ocean.

**Table 2.** Surface P inputs in REF in relation to surface DIP concentrations (first 12 m).

Region	P inputs (percentage of global input) [Tg P yr <sup>-1</sup> ]	Surface layer DIP inventories Tg P	Input to inventory ratio
Open equatorial Pacific	0.33 (9%)	5.15	6%
Open equatorial Atlantic	0.12 (3%)	0.41	29%
Southern Ocean	0.20 (6%)	9.82	2%
Coastal Ocean	0.19 (6%)	3.08	6%

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