



Surface Water Monitoring Program for Pesticides in Salmonid-Bearing Streams, 2009 Data Summary

**A Cooperative Study by the Washington State
Departments of Ecology and Agriculture**



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A Cooperative Study by the Washington State Departments of Ecology and Agriculture

by
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Abstract

The Washington State Departments of Agriculture and Ecology are conducting a multi-year monitoring study to characterize pesticide concentrations in selected salmon-bearing streams during a typical pesticide-use period.

Monitoring is being conducting in six basins:

- Thornton Creek in the Cedar-Sammamish basin and Longfellow Creek in the Green-Duwamish basin, representing urban land use.
- Lower Skagit-Samish basin, representing western Washington agriculture.
- Lower Yakima basin, representing eastern Washington irrigated agriculture.
- Wenatchee and Entiat basins, representing central Washington tree fruit agriculture.

This report summarizes data collected during 2009 from four basins: Cedar-Sammamish, Green-Duwamish, Skagit-Samish, and lower Yakima. The report also provides a more intensive review of data collected during 2007-2009 from two basins: Wenatchee and Entiat.

In 2009, samples were collected for analysis of over 165 pesticides and pesticide degradates, as well as total suspended solids. Field data were collected for streamflow, temperature, pH, conductivity, and dissolved oxygen.

Changes to the monitoring study during 2009 include:

- Discontinuing monitoring at the upstream Thornton Creek site.
- Adding an urban land-use site in the Green-Duwamish basin (Longfellow Creek).
- Adding analysis for 18 pesticides and degradate pesticides.

An intensive review of pesticide results is conducted on a triennial basis. Year 2009 is the first in a three-year study cycle to investigate pesticides in the Green-Duwamish basin, the fourth in a six-year cycle in the Skagit-Samish basin, and the seventh in a nine-year cycle in the Cedar-Sammamish and Lower Yakima basins. Triennial results for the Wenatchee-Entiat basins are included in this report.

During 2007-09 few pesticides were detected at the Wenatchee-Entiat basin sites with the exception of Brender Creek. This is in part due to higher streamflows at some of the sites including Peshastin Creek and the Wenatchee and Entiat Rivers. Brender Creek endosulfan levels exceeded the endangered species level of concern for salmonids and the Washington State chronic water quality standard, indicating potential chronic health effects to aquatic life during mid-March through May. Pesticide concentrations at the Entiat River site met all U.S. Environmental Protection Agency (EPA) Pesticide Registration Toxicity Criteria and EPA National Recommended Water Quality Criteria as well as Washington State Water Quality Standards.

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Introduction

The Washington State Departments of Agriculture (WSDA) and Ecology (Ecology) are conducting a multi-year monitoring study to evaluate pesticide concentrations in surface water. The study assesses pesticide presence in salmon-bearing streams during a typical pesticide-use season (e.g., March through October).

WSDA, the U.S. Environmental Protection Agency (EPA), the National Atmospheric and Oceanic Administration (NOAA) National Marine Fisheries Service, and the U.S. Fish and Wildlife Service (USFWS) use the data collected from this study to refine exposure assessments for pesticides that are registered for use in Washington State. Understanding the fate and transport of pesticides allows regulators to assess the potential effects of pesticides on endangered salmon species while minimizing the economic impacts to agriculture.

The purpose of this data report is to provide results from monitoring conducted in 2009 in six basins and to document any changes that occurred in the program during the year. This report also includes an in-depth analysis of data collected during 2007-09 in the Wenatchee-Entiat basins. This three-year review includes an examination of trends and relationships and determines if water quality concentrations are healthy for aquatic life.

A triennial report for data collected during 2006-08 from Thornton Creek (Cedar-Sammamish basin), the Skagit-Samish basin, and the lower Yakima basin are included in Sargeant et al. (2010). A triennial review of the Wenatchee-Entiat sites was not included in the 2010 report because only two years of data were available at that time.

Study Area

This pesticide monitoring project has been ongoing since 2003. As the project has progressed, additional sampling areas have been added.

Basins Monitored During 2009

The six basins monitored in 2009 are presented in Figure 1: two urban and four agricultural basins. The urban basins were chosen due to land-use characteristics, history of pesticide detections, and habitat use by salmon. The agricultural basins were chosen because they support several salmonid populations, produce a variety of agricultural commodities, and have a high percentage of cultivated areas.

Monitoring areas and timeframes are:

- Thornton Creek, located in the Cedar-Sammamish basin (WRIA¹ 8), represents an urban land-use area. Two to four sites have been sampled on this creek from 2003-08. One site at the mouth of Thornton Creek was sampled in 2009.
- Longfellow Creek, located in the Green-Duwamish basin (WRIA 9), represents an urban land-use area. Sampling started on this creek at one site in 2009.
- Four sub-basins of the lower Skagit-Samish basin (WRIA 3) were selected to represent western Washington agricultural land-use practices. The Samish River, Big Ditch Slough, Browns Slough, and Indian Slough have been sampled since 2006.
- Three sub-basins of the Lower Yakima basin (WRIA 37) were selected to represent eastern Washington irrigated crop-land agricultural practices. Marion Drain, Sulphur Creek Wasteway, and Spring Creek have been sampled since the start of the project in 2003.
- Four sub-basins of the Wenatchee basin (WRIA 45) and Entiat basin (WRIA 46) were selected to represent central Washington agricultural tree fruit practices. Peshastin Creek, Mission Creek, Brender Creek, and the Wenatchee River (WRIA 45) and the Entiat River (WRIA 46) have been sampled since 2007.

Site locations and duration of sampling are described in Appendix A.

A full description of the Wenatchee-Entiat monitoring sites is included below. A detailed description of the Wenatchee-Entiat sites is provided in this report because of the triennial (2007-09) review. Descriptions of sites in the other four basins, including basin description, site map, climate, agricultural land-use, and the salmon fishery, are included in the last triennial report (Sargeant et al., 2010).

¹ Water Resource Inventory Area



Figure 1. State map showing the six urban and agricultural basins monitored during 2009.

Wenatchee-Entiat Basins, 2007-09

The Wenatchee River drains a portion of the east slopes of the Cascade Mountains in north-central Washington within Chelan County (Figure 1). The river flows generally in a southeasterly direction, emptying into the Columbia River at the City of Wenatchee. The Wenatchee River basin encompasses about 1,371 square miles. Lake Wenatchee is the source of the Wenatchee River. Major tributaries include the Chiwawa River and Icicle, Nason, Chumstick, Peshastin, and Mission Creeks. The primary land uses within the Wenatchee River basin are forestry, wilderness, agriculture, range, residential, and recreation.

The federal government is the largest landowner in the Wenatchee basin, with approximately 671,220 acres, 76% of the basin. Only 17% of the land is privately owned. Privately owned land occurs mostly in the low-lying valley bottoms and in the southern portion of the basin along the Wenatchee River and its major tributaries (Andonaegui, 2001).

The Wenatchee and Entiat watersheds (WRIs 45 and 46) support diverse salmon populations and produce a variety of agricultural commodities. Agriculture in the basins is dominated by orchard crops. Because previous studies showed pesticide detections in surface water, the Wenatchee-Entiat was added as an index watershed for evaluation of eastern Washington tree fruit agricultural practices. Sampling of the Wenatchee-Entiat began in 2007 as described in Dugger et al. (2007).

Sampling Sites

Sampling for this project is conducted at five sites in the Wenatchee-Entiat basins:

- Peshastin Creek at river mile 0.1
- Mission Creek at river mile 3.1
- Brender Creek at river mile 0.7
- Wenatchee River at river mile 2.8
- Entiat River at river mile 1.4

Figure 2 presents the locations of the five sampling sites. Appendix B describes sampling locations and duration of sampling for each site.

Figure 2. Location of sampling sites in the Wenatchee and Entiat basins.

The sampling sites are located to minimize the influence of residential areas. Brender Creek discharges to Mission Creek downstream of the confluence with Yaksum Creek. Peshastin and Mission Creeks discharge to the Wenatchee River, and the Wenatchee and Entiat Rivers discharge to the Columbia River.

Precipitation and Streamflow

In the Wenatchee basin, most precipitation occurs in late fall and winter. In the upper watershed, the Cascade Mountain area is characterized by heavy precipitation and snow, nearly 150 inches annually. Most of the precipitation occurs during the winter as snow. Temperatures at Wenatchee range from a January average of 26 °F to a July average of 73 °F. As air masses move east toward the Columbia Basin, moisture progressively decreases, resulting in arid conditions within the lowermost region of the watershed. In contrast to the mountainous areas, the City of Wenatchee receives only 8.5 inches or less of precipitation annually, with maximum summer temperatures averaging 95-100 °F (Andonaegui, 2001).

For the Wenatchee River at Monitor, the highest average monthly streamflows occur in May and June during spring snowmelt (Table 1) (USGS, 2009a).

Table 1. Average, maximum, and minimum streamflows (cfs) for the Wenatchee River at Monitor, 1963-2009 (USGS, 2009a).

	Oct	Nov	Dec	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Sep
Mean	1061	2194	1976	1839	1940	2375	3916	8032	8815	4301	1424	785
Max.	3095	9636	6983	4309	5447	6853	7260	12970	17020	9880	3985	1628
Min.	346	426	556	527	518	995	1634	3565	2273	1015	425	301

Peshastin Creek is a tributary to the Wenatchee River, originating at Blewett Pass and flowing in a northeasterly direction for 15.4 miles before entering the Wenatchee River at river mile 17.9, downstream of the town of Peshastin. Although it is one of the major subbasins in the Wenatchee basin in terms of size, Peshastin Creek contributes only 4% of the summer low flow in the Wenatchee River. The lower portion of the Peshastin Creek subbasin is more arid, with annual precipitation levels ranging from 80 inches in the upper elevations to 15 inches at the mouth of Peshastin Creek.

The Mission Creek subbasin is 93 square miles (59,609 acres). Mission Creek flows 9.4 miles before discharging to the Wenatchee River at RM 10.4 at the town of Cashmere. The average annual precipitation for the Mission Creek subbasin is 19 inches. Mission Creek contributes only 1% of the average annual flow of the Wenatchee River. Approximately 0.4% of the acreage in Mission Creek is in agricultural production. Brender Creek enters Mission Creek at RM 0.2, within the town of Cashmere, just upstream of the mouth of Mission Creek.

Flow characteristics in Mission and Brender Creeks are complicated by (1) diversions of surface water from Mission Creek, and (2) the influence of irrigation waters conveyed from Icicle and Peshastin Creeks into Mission and Brender Creeks. While reaches in both creeks have

historically gone dry, currently Brender Creek has year-round flow due to irrigation-return flows from the Peshastin Irrigation District (Andonaegui, 2001). At times Brender Creek receives more than 50% of its flow from the Peshastin Canal (Rickel, 2009).

The Entiat River basin is located in north-central Washington in Chelan County. It originates in a glaciated basin near the crest of the Cascade Mountains and flows southeasterly, meeting the Columbia River near the town of Entiat, about 20 miles upstream from Wenatchee. The drainage area is about 268,000 acres of which approximately 224,000 acres (84%) are in public ownership, primarily national forest. There are 1,300 acres of orchard land in the lower valley.

Mean annual precipitation in the Entiat basin ranges from 90 inches in the moist, alpine-type higher elevations to less than 10 inches in the arid shrub steppe of the lowest elevations. Most winter precipitation falls as snow; however, rain is not unusual. During the summer, mean temperatures in the lower Entiat watershed usually range from 60-70°F, decreasing to the 50s (°F) at higher elevations (Andonaegui, 1999).

As with the Wenatchee River, the highest average monthly streamflows seen in the Entiat River occur in May and June during spring snowmelt (Table 2) (USGS, 2009b).

Table 2. Average, maximum, and minimum streamflows (cfs) for the Entiat River near Entiat, 1996-2009 (USGS, 2009b).

	Oct	Nov	Dec	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Sep
Mean	147	216	166	182	174	263	559	1426	1600	675	233	135
Max.	301	508	316	330	290	623	1090	2277	2674	1682	655	232
Min.	89.4	99.7	101	108	92.1	125	165	673	497	213	93.5	71.2

Agricultural Land Use

The Wenatchee and Entiat basins produce a variety of agricultural products, with orchard crops (tree fruit) being the major agricultural commodity.

Table 3 has estimates of crop totals for the Wenatchee and Entiat basins (WSDA, 2009).

Approximately 0.6% of the Peshastin Creek subbasin is in agricultural production, with major crops being pears, apples, and cherries. For Mission Creek, 4% of the subbasin is in agricultural production, with pears as the major product. The Brender Creek subbasin has the greatest area in agricultural production (11%), with major crops being pears, apples, and cherries.

Approximately 1% of the Wenatchee basin is in production with pear and apple orchards.

Less than 1% of the Entiat basin is in agricultural production, with the major crops being pears and apples.

Table 3. Crop totals for the Wenatchee-Entiat sites (WSDA, 2009).

Site and Land Use	Area (acres)	Percent of Basin or Subbasin
Peshastin Creek		
Apple	33	0.038%
Cherry	10	0.012%
Fallow	14	0.016%
Pear	488	0.566%
Total Cropped Area	545	0.632%
Subbasin Area	86250	
Mission Creek		
Alfalfa/Grass, Hay	12	0.023%
Cherry	7	0.014%
Christmas Tree	5	0.009%
Pear	177	0.338%
Total Cropped Area	202	0.385%
Subbasin Area	52386	
Brender Creek		
Apple	112	1.640%
Cherry	59	0.865%
Fallow	23	0.335%
Golf Course	36	0.519%
Pear	525	7.640%
Total Cropped Area	755	11.00%
Subbasin Area	6866	
Wenatchee River		
Alfalfa/Grass, Hay	19	0.002%
Apple	1,018	0.120%
Apricot	1	< 0.001%
Cherry	326	0.038%
Christmas Tree	5	0.001%
Developed	284	0.033%
Fallow	166	0.020%
Golf Course	113	0.013%
Grape, Wine	10	0.001%
Grass, Hay	91	0.011%
Nectarine/Peach	10	0.001%
Nursery, Lavender	1	< 0.001%
Pear	6,509	0.766%
Total Cropped Area	8,554	1.010%
Basin Area	849905	
Entiat River		
Alfalfa/Grass, Hay	1	< 0.001%
Apple	170	0.064%
Cherry	31	0.012%
Fallow	66	0.025%
Grass, Hay	6	0.002%
Pasture	2	0.001%
Pear	529	0.199%
Unknown	3	0.001%
Total Cropped Area	808	0.304%
Basin Area	265434	

Salmonid Fishery Use

A summary of salmonid distribution and use is presented in Table 4. Salmonid distribution and habitat is classified according to the highest level of habitat supported. The greatest value is placed on spawning habitat, followed by rearing and migration. Habitat is classified for the stream reach where the sample station is located; higher quality habitat may be available in the upper watershed.

Table 4. Salmonid presence and use for the Wenatchee-Entiat sites.

(StreamNet, 2009; Burke et al., 2006.)

Species	Wenatchee River	Mission Creek	Brender Creek	Peshastin Creek	Entiat River
Spring chinook	Rearing	Rearing	--	Rearing	Rearing
Summer chinook	Spawning	Spawning	Presence	--	Presence
Coho	--	--	--	--	Spawning
Sockeye	Rearing	--	--	--	Presence
Bull trout	Rearing	--	--	Presence	Presence
Summer steelhead	Rearing	Spawning	Presence	Rearing	Spawning

Tables 5, 6, and 7 present the life phases and periods when salmonid species are present in Peshastin and Mission Creeks and the lower Wenatchee River (EES Consulting Inc. and Thomas R. Payne & Associates, 2005).

Entiat River salmonid life phases and periods of use are presented in Table 8 (Chelan County Conservation District, 2004).

Table 5. Timing of salmonid life phases in the Peshastin Creek subbasin.

Periods of heaviest fish use are in black, periods of moderate use are in gray, and periods of little or no use are in white. (EES Consulting, Inc. and Thomas R. Payne & Associates, 2005.)

Species	Life Stage	October	November	December	January	February	March	April	May	June	July	August	September
Spring	Spawning												
Chinook	Incubation												
	Rearing												
	In-migration												
Steelhead	Spawning												
	Incubation												
	Rearing												
	In-migration												
Bull Trout	Spawning												
	Incubation												
	Rearing												

Table 6. Timing of salmonid life phases in the Mission Creek subbasin.

Periods of heaviest fish use are in black, periods of moderate use are in gray, and periods of little or no use are in white. (EES Consulting, Inc. and Thomas R. Payne & Associates, 2005.)

Species	Life Stage	October	November	December	January	February	March	April	May	June	July	August	September
Spring	Spawning												
Chinook	Incubation												
	Rearing												
	In-migration												
Summer	Spawning												
Chinook	Incubation												
	Rearing												
	In-migration												
Steelhead	Spawning												
	Incubation												
	Rearing												
	In-migration												

Table 7. Timing of salmonid life phases in the lower Wenatchee River basin.

Periods of heaviest fish use are in black, periods of moderate use are in gray, and periods of little or no use are in white. (EES Consulting, Inc. and Thomas R. Payne & Associates, 2005.)

Species	Life Stage	October	November	December	January	February	March	April	May	June	July	August	September
Spring	Spawning												
Chinook	Incubation												
	Rearing												
	In-migration												
Summer	Spawning												
Chinook	Incubation												
	Rearing												
	In-migration												
Steelhead	Spawning												
	Incubation												
	Rearing												
	In-migration												
Bull Trout	Spawning												
	Incubation												
	Rearing												

Table 8. Timing of salmonid life phases in the Entiat River basin.

Periods of heaviest fish use are in black, periods of moderate use are in gray, and periods of little or no use are in white. (Chelan County Conservation District, 2004.)

Species	Life Stage	January	February	March	April	May	June	July	August	September	October	November	December
Late Run Chinook	Spawning												
	Incubation												
	Emergence												
	Fry Colonization												
	0-Age Active Rearing												
	0-Age Migrant												
	Prespawning migrant + Holding												
	Spring Chinook	Spawning											
Incubation													
Emergence													
Fry Colonization													
0-Age Active Rearing													
0-Age Migrant													
1-Age Transient Rearing													
Prespawning migrant + Holding													
Steelhead	Spawning												
	Incubation												
	Emergence												
	Fry Colonization												
	0-Age Active Rearing												
	0-Age Migrant												
	1-Age Resident Rearing												
	1-Age Transient Rearing												
2+-Age Transient Rearing													
Prespawning migrant + Holding													

Study Design and Methods

Sampling was designed to address pesticide presence in salmonid-bearing streams during a typical pesticide-use period (e.g., March through September). The focus of monitoring is on currently registered pesticides, but laboratory analysis also included some historically used pesticides. Conventional water quality parameters were measured: total suspended solids (TSS), pH, conductivity, temperature, dissolved oxygen, and streamflow. The conventional parameters provide information to help better determine the factors influencing pesticide toxicity, fate and transport, and general water quality.

Detailed information on study design and methods are described in the Quality Assurance (QA) Project Plan (Johnson and Cowles, 2003), subsequent addendums (Burke and Anderson, 2006; Dugger et al., 2007; Anderson and Sargeant, 2009), and the triennial reports (Burke et al., 2006; Sargeant et al., 2010).

During 2009, samples were collected for analysis of 169 pesticides and degradates: 70 insecticides, 58 herbicides, 28 degradate pesticides, 10 fungicides, two synergistic compounds, and one wood preservative. Due to concerns about false positive results, the following analytes were rejected and not included in the 2009 data set: 1-naphthol, aldicarb sulfone, aldicarb sulfoxide, and oxamyl.

Sampling Sites and Sampling Frequency

Changes to the 2009 sampling program include discontinuing the upstream Thornton Creek (TC-1) sampling site and adding a site on Longfellow Creek (LC-1) in the Green-Duwamish basin. The new site was selected collaboratively with the USFWS and the NOAA National Marine Fisheries Service. Figure 1 shows the location of the Longfellow Creek sampling site. The QA Project Plan Addendum (Anderson and Sargeant, 2009) describes this change.

In 2009, 27 sampling events were conducted. Sampling began the second week in March and continued through the second week in September at all sites except Marion Drain. As in previous years, Marion Drain sampling continued through the end of October (for a total of 34 sampling events) for organophosphate pesticides and TSS. The upstream Spring Creek site in the lower Yakima basin was sampled every other week for a total of 14 sampling events.

Field Procedures and Laboratory Analyses

A full description of field procedures and laboratory analysis is included in Sargeant et al. (2010). Field methods for grab sampling are a direct application or modification of United States Geological Survey (USGS) or EPA procedures. Surface water samples were collected by hand-compositing grab samples from quarter-point transects across each stream. In situations where streamflow was vertically integrated, a one-liter transfer container was used to dip and pour water from the stream into sample containers. Otherwise samples were collected using depth integrating equipment. Sample/transfer containers were delivered pre-cleaned by the

manufacturer to EPA specifications (EPA, 1990). After collection, all samples were labeled and preserved according to the QA Project Plan (Johnson and Cowles, 2003).

Field meters were calibrated at the beginning of the field day according to manufacturers' specifications, using Ecology standard operating procedures (SOPs) (Swanson, 2007). Meters were post-checked at the end of the field day using known standards. Conventional parameters measured in the field were replicated once per sample day. Dissolved oxygen meter results were compared to grab samples that were analyzed by Winkler Titration for dissolved oxygen following Ecology SOPs (Ward, 2007). Two to three Winkler grab samples were obtained during each sample day. Continuous, 30-minute interval, temperature data were collected year-round in 2009. Temperature instruments were calibrated against a National Institute of Standards and Technology (NIST) primary reference (Wagner et al., 2000). Data quality objectives for field meters are described in Anderson and Sargeant (2009).

Discharge for sites other than Sulphur Creek Wasteway, Wenatchee River, and Entiat River were measured using a Marsh-McBirney flow meter and top-setting wading rod, as described in the USGS method (Rantz et al., 1983). Discharge data for Sulphur Creek Wasteway were obtained from an adjacent U.S. Bureau of Reclamation gaging station, "SUCW – Sulphur Creek Wasteway at Holaday Road near Sunnyside". Wenatchee and Entiat River discharge data were obtained from USGS at the Wenatchee River at Monitor (Station 12462500) and Entiat River near Entiat (Station 12452990). Fifteen-minute discharges were available during the sampling period. The record closest to the actual sampling time was used in lieu of field measurements.

Ecology's Manchester Environmental Laboratory (MEL) analyzed all pesticide and TSS samples. Laboratory methods are presented in Table 9. A list of target analytes for this study is presented in Appendix C. Laboratory methods are also discussed in the QA Project Plan (Anderson and Sargeant, 2009); previous QA Project Plan (Johnson and Cowles, 2003) and QA Project Plan addendum (Burke and Anderson, 2006); and SOP for the *Pesticides in Salmonid Streams Project* (Anderson and Sargeant, 2010).

Table 9. Summary of 2009 laboratory methods.

Analyte	Analytical Methods ¹		
	Extraction	Analysis	Reference
Pesticides ²	3510	GC/MS	8270
Herbicides	8151	GC/MS	8270
Carbamates	3535M	HPLC	8321AM
Total Suspended Solids	n/a	Gravimetric	EPA 160.2

¹ All analytical methods refer to EPA SW 846, unless otherwise noted.

² Pesticides refers to all forms tested unless indicated otherwise.

GC: gas chromatograph.

MS: mass spectrometry.

HPLC: high performance liquid chromatography.

n/a: not applicable.

Data Quality

Laboratory Data Quality

Performance of laboratory analyses is governed by QA and quality control (QC) protocols. The QA/QC protocol employs application of blanks, replicates, surrogates, laboratory control samples; and matrix spike/matrix spike duplicates (MS/MSD). Laboratory surrogate, blank, replicate, and control samples are analyzed as the laboratory component of QA/QC. Field blanks, replicates, and MS/MSDs integrate field and laboratory components. A summary of laboratory and field data quality are presented below. For a detailed discussion of 2009 data quality, refer to Appendix C. Data quality results for 2007 and 2008 are presented in Sargeant et al. (2010).

Across the six study basins, field blank detections of certain carbamate compounds indicated problems with select carbamate parameters: 1-naphthol, aldicarb sulfone, aldicarb sulfoxide, and oxamyl. During 2009 an anomaly in the analytical method for carbamate pesticides was identified. This analytical anomaly caused false positive identification for 1-naphthol, aldicarb sulfone, and aldicarb sulfoxide in 2006-2009 and for oxamyl in 2009. Data for these parameters are not reported for this 2009 study. Although QA/QC criteria were met for all reported carbamate values, there is a possibility of some false positives.

In the initial phase of this multi-year project, a large portion of the budget was allocated toward QA/QC. Currently, QA/QC is approximately 17% of the budget. The large number of QA/QC samples necessitates the use of a pre-planned schedule. At a minimum, each week there is at least one replicate, one blank, and one MS/MSD covering at least one of the four laboratory analyses (PESTMS, HERBS, CARBAMLL, and TSS). QA/QC samples were concentrated during April, May, and June to cover the intensive application period for most pesticides. Sites were randomly selected for application of QA/QC samples.

Laboratory Blanks

In 2009 no laboratory blank detections occurred for the pesticide analysis, gas chromatography mass spectrometry (GCMS), or herbicide analysis. There were laboratory blank detections for the low level carbamate analysis (Appendix C, Table C-9). Five of the eight blank detections were for aldicarb sulfone, aldicarb sulfoxide, or oxamyl. Data for 2009 are not reported for these parameters due to concerns about false positives.

Field Blanks

Field blank detections indicate the potential for sample contamination in the field and laboratory as well as the potential for false detections due to analytical error. In 2009 there were two detections in field blanks: one of dichlobenil in Longfellow Creek and one of tricyclazole in Brender Creek.

Dichlobenil was not found in the associated sample at Longfellow Creek but was detected at other western Washington sites on the same day. Dichlobenil was detected in all the other 2009 samples for Longfellow Creek. Because the week in which the field blank detection occurred had no corresponding detection in the associated sample, it is likely that the field blank sample and the associated Longfellow Creek sample were unintentionally switched prior to analysis. However, all western Washington dichlobenil detections for this week (first week of sampling) are still qualified because it is impossible to be certain that this error occurred.

Tricyclazole was not detected in any of the associated samples, thus no sample detections were qualified.

Replicate Results

Replicate sampling tests the reproducibility or precision of sampling results. Field replicate sampling frequency was 8.7% during 2009, and 2.5% of the pairs had a detection in at least one replicate. For pesticides, 39 pesticides were detected in 113 replicate pairs. Of these, 81% were consistently identified in both samples. The average relative percent difference (RPD) of consistent field replicate pairs was low, 9.6%, and similarly the median pooled relative standard deviation (RSD) was 6.3%. The 2009, replicate variability is lower than 2006-2008 results (8.1% RSD) and the National Water Quality Assessment (NAWQA) median pooled RSD of 15% at concentrations < 0.01 ug/L and 12% at concentration near 0.01 ug/L (Martin, 2002).

TSS was detected in 32 replicate pairs. TSS replicate pairs were consistently identified, and 75% of the replicates were within the 20% RPD criterion. For the 25% of the replicates that did not meet the 20% RPD criterion, TSS concentrations were at or near the reporting limit. When replicates did not fall within the acceptable range, it is likely due to the high variability in detections near the minimum reporting limit (Mathieu, 2006).

Surrogates and Matrix Spikes

Surrogates are used to evaluate recovery for a group of compounds. The majority of surrogate recoveries fell within the control limits established by Manchester Laboratory for all compounds except chlorpyrifos-D10. Chlorpyrifos-D10 was used as a surrogate for organophosphate pesticides in GCMS analysis in late 2009. Recoveries were high. No sample results were qualified because all other GCMS surrogates, including the other organophosphate surrogate, triphenyl phosphate, were acceptably recovered in all samples.

MS/MSD provide an indication of bias due to interferences from components of the sample matrix. The duplicate spike can be used to estimate analytical precision at the concentration of the spiked samples. The average recovery of matrix-spiked compounds was 88.7%, and the average RPD between MS/MSD pairs was 12.2%. For most compounds, recovery and RPDs of MS/MSD pairs showed acceptable performance and were within defined limits for the project. Results with an average RPD outside of the $\pm 40\%$ criteria were qualified as estimates.

Field Data Quality

A detailed discussion of 2009 field data quality is included in Appendix C. In 2009 the field meter for the lower Yakima and Wenatchee-Entiat sites (eastside sites) met QC objectives including post-checks and Winkler comparisons except on March 18, 2009. Dissolved oxygen meter readings were biased high that day: meter and Winkler dissolved oxygen % RSD ranged from 11.1-13.6%. Only Winkler dissolved oxygen results will be reported for this day.

The field meter for the urban and lower Skagit-Samish sites (westside sites) did not meet post-check QC objectives for conductivity for the following dates: March 16 and 25, April 22 and 27, May 6, 20, and 26, 2009. Conductivity results for these days are rejected and not reported.

Data Analysis Methods

Field and laboratory data were compiled and organized using Excel[®] spreadsheet software and Access[®] database software (Microsoft Corporation, 2007). Water quality results from field and laboratory work were also entered into Ecology's Environmental Information Management (EIM) database (www.ecy.wa.gov/eim).

This report contains only 2009 data for Thornton and Longfellow Creeks, the Skagit-Samish sites, and the lower Yakima sites. For the Wenatchee-Entiat sites, this report contains a more in-depth analysis as part of a three-year review (2007-09). Data analysis methods for the three-year review are described below.

Protocols for Analysis of Pesticide Data

The following guidelines were used in reporting and analyzing data for this report.

Pesticide Detections

Laboratory data were qualified as needed, and qualifiers are described in Table 10. A positive pesticide detection included un-qualified values and values qualified with a J or E. Values qualified with NJ, U, or UJ were considered non-detects.

Table 10. Definitions of data qualifiers.

Qualifier	Definition
No qualifier	The analyte was detected at the reported concentration. Data are not qualified.
E	Reported result is an estimate because it exceeds the calibration range.
J	The analyte was positively identified; the associated numeric value is the approximate concentration of the analyte in the sample.
NJ	The analysis indicates the presence of an analyte that has been “tentatively identified,” and the associated numeric value represents its approximate concentration.
NAF	Not analyzed for.
NC	Not calculated.
REJ	The sample results are rejected due to serious deficiencies in the ability to analyze the sample and meet QC criteria. The presence or absence of the analyte cannot be verified.
U	The analyte was not detected at or above the reported sample quantitation limit.
UJ	The analyte was not detected at or above the reported sample quantitation limit. However, the reported quantitation limit is approximate and may or may not represent the actual limit of quantitation necessary to accurately measure the analyte in the sample.

MEL, 2000, 2008; EPA, 1999, 2007.

Comparison to Assessment Criteria and Water Quality Standards

Non-detect values (U, UJ, N, NJ) were not used for comparison to *assessment criteria* or *water quality standards*. When summing compound totals (such as total DDT, total endosulfan), the Toxic Studies Unit Guidance was used (Ecology, 2008). Non-detects (U, UJ) were assigned a value of zero (as in the guidance). Unlike the guidance, NJ values (tentatively identified compounds) were also assigned a value of zero.

Data Analysis

Graphs, plots, mass balance calculations, and some statistical analyses were made using Excel® software. For statistical trend analysis, WQHYDRO software (Aroner, 2007) was used. For correlation analysis, R statistical software was used (R Foundation for Statistical Computing, 2010).

Replicate Values

Field and laboratory replicates were obtained to determine data quality. Field and laboratory replicates were arithmetically averaged for comparisons to *assessment criteria* and *water quality standards*. For data analysis purposes, field and laboratory replicates were arithmetically averaged. If the sample or the replicate was a non-detect value while the other (either sample or replicate) was a detection, then the detected value was used.

When a laboratory replicate was performed on a field replicate, the laboratory replicate mean was calculated before the field replicate mean.

For select statistical analysis, NJ qualified data were used when detected pesticide values were not available. When this occurred, it is specified in the statistical test description.

Statistical Analysis

Summary Statistics

For 2007-09, the laboratory analyzed samples for over 160 pesticides and degradates. For a majority of compounds, concentrations were below the analytical reporting limit of the laboratory and were reported as “less than” the reporting limit. These “less-than” reporting limit values make it difficult to analyze data statistically. Substituting a value of zero or a value half the detection limit is not defensible, and results may vary depending on the substituted value selected.

Correlations

Correlation analysis was used to examine the association between pesticide concentrations and variables such as TSS, flow, and rainfall (day of rainfall, the sum of day of rainfall and previous 24-hour rainfall, previous 24-hour rainfall, and previous 48-hour rainfall). A two-tailed, Kendall’s tau-b, a non-parametric correlation coefficient, was used to test for correlation between parameters. NJ (analyte was tentatively identified) qualified data were used in this test. For pesticides and TSS, the data were first graphed and visually inspected to select data for analysis. Selected periods during the sampling season were tested where appropriate. Some pesticides are only detected during a select period; this minimizes the number of non-detect data in the analysis.

Assessment Criteria and Washington State Water Quality Standards

Assessment of pesticide effects on endangered salmonid species is evaluated by comparing detected pesticide concentrations against three criteria:

- EPA FIFRA Pesticide Registration Toxicity Criteria.
- EPA National Recommended Water Quality Criteria (NRWQC).
- Washington State Water Quality Standards for the Protection of Aquatic Life (WAC 173-201A).

The EPA and Washington State aquatic life criteria are based on evaluating the effects of a single chemical on a wide array of families, including at least one fish species from the family *Salmonidae* and a second species in the class *Osteichthys*. The criteria and standards do not account for the effects of multiple chemicals or pesticide mixtures on an organism. Many of the pesticides included in this study do not have EPA or Washington State criteria.

Aquatic life criteria, pesticide regulatory criteria, and toxicity (acute and chronic) results for fish, invertebrates, and aquatic plants are presented in Appendix D. Measured concentrations higher than criteria concentrations do not necessarily indicate that the water quality criteria have not been met. Numeric water quality criteria contain concentration values and duration of exposure components; both must be compared to the measured concentrations to assess compliance with the criteria.

In this report, EPA Pesticide Registration Toxicity Criteria and EPA NRWQC will be referred to as *assessment criteria*. Washington State numeric water quality standards for pesticides will be referred to as *water quality standards*. For a description of these criteria and standards for pesticides, refer to Appendix D.

EPA Pesticide Registration Toxicity Criteria

The EPA uses risk quotients (RQ) to assess the potential risk of a pesticide to non-target organisms. A RQ is calculated by dividing the environmental concentration by either an acute or chronic toxicity value, which gives an evaluation of exposure over toxicity. The resulting RQ is a unitless value that is compared to Levels of Concern (LOC). The LOCs set by EPA are presented in Table 11 and are used to assess the potential risk of a pesticide to non-target organisms.

The endangered species LOC (0.05 for aquatic species) is used as a comparative value to assess potential risk to threatened or endangered salmonids. The endangered species RQ can also be expressed as 1/20th of the acute Lethal Concentration 50 (LC₅₀) for aquatic organisms. To assess the potential risk of a pesticide to salmonids, the LC₅₀ for rainbow trout is commonly used as a surrogate species. Thus the endangered species LOC presented in subsequent tables are 1/20th of

the rainbow trout LC₅₀. When available, the endangered species LOC for specific salmonids is also presented.

Table 11. Risk quotient criteria for direct and indirect effects of pesticides on aquatic organisms.

Test Data	Risk Quotient	Presumption
Acute LC ₅₀	>0.5	Potentially high acute risk.
	>0.1	Risk that may be mitigated through restricted use classification.
	>0.05	Endangered species may be affected acutely, including sublethal effects.
Chronic NOEC	>1	Chronic risk; endangered species may be affected chronically, including reproduction and effects on progeny.
Acute invertebrate LC ₅₀	>0.5	May be indirect effects on T&E fish through food supply reduction.
Aquatic plant acute LC ₅₀	>1	May be indirect effects on aquatic vegetative cover for T&E fish.

(Turner, 2003).

NOEC – No observable effect concentration.

T&E – Threatened and endangered.

Acute toxicity is calculated by standardized toxicity tests using lethality as the measured criteria. A properly conducted test will use a sensitive (representative) species at a susceptible life stage (usually young, though not immature). The test also will subject the test species to a pesticide under a range of concentrations (minimum: no effect, 50% and 100% mortality). The dose response curve may be calculated, and the LC₅₀, lethal concentration to cause mortality in 50% of test species will be derived. For fish, the lethality test is conducted over 96 hours at a constant concentration. Acute invertebrate toxicity is normally calculated over 48 hours, with the criteria being mortality or immobility (LC₅₀, or Effective Concentration - EC₅₀ for immobility). Acute toxicity testing for aquatic plants is conducted over 96 hours; the criterion is based on reduction in aquatic plant growth (EC₅₀).

Chronic fish tests normally use reproductive effects, or effects to offspring, as the measured effect. The dose response curve is evaluated to determine a no observable effect concentration (NOEC). The chronic toxicity test is longer than the 96-hour acute test (21 day for fish, 14 days for invertebrates, 5 to 60 days for plants) to simulate exposure resulting from a persistent chemical or effect of repeated applications.

Toxicity values such as those used for pesticide registration are determined from continuous exposure over time (e.g., LC₅₀ freshwater fish acute toxicity tests are run for 96 hours at a constant concentration). When comparing the monitoring data to either the aquatic life criteria or directly to the toxicity criteria, one must consider the duration of exposure as well as the numeric toxicity value. For pesticide registration criteria, it is not possible to determine if an aquatic life criterion has been met based solely on an individual sample because the sampling frequency is usually weekly. Weekly sampling does not allow for assessment of the temporal component of the criteria.

EPA National Recommended Water Quality Criteria

The NRWQC are established by the EPA Office of Water for the protection of aquatic life, as established under the Clean Water Act (33 U.S.C. 1251 et. seq.). The pesticide criteria established under the Act are based on vertebrate and invertebrate acute and chronic toxicological data. States often adopt the NRWQC as their promulgated (legal) standards. The NRWQC was updated in 2006, and those criteria are used in this report (EPA, 2006).

Washington State Water Quality Standards

Pesticides

Washington State water quality standards are established in the Washington Administrative Code (WAC), Chapter 173-201A. These standards include numeric pesticide criteria for the protection of aquatic life.

The aquatic life criteria are designed to protect for both short-term (acute) and long-term (chronic) effects of chemical exposure. The criteria are primarily intended to avoid direct lethality, and growth and reproductive effects, to fish and other aquatic life within the specified exposure periods. The chronic criteria for a number of the chlorinated pesticides are based on protection of fish-eating wildlife from adverse effects due to bioaccumulation.

The exposure periods assigned to the acute criteria are expressed as: (1) an instantaneous concentration not to be exceeded at any time or (2) a one-hour average concentration not to be exceeded more than once every three years on average. The exposure periods for the chronic criteria are either: (1) a 24-hour average not to be exceeded at any time or (2) a four-day average concentration not to be exceeded more than once every three years on the average. For federal Clean Water Act section 303(d) listing purposes, measurements of instantaneous concentrations are assumed to represent the averaging periods specified in the water quality standards for both acute and chronic criteria, unless additional measurements are available to calculate averages (Ecology, 2006).

Because few water quality criteria for pesticides have been developed, the majority of comparisons to measured pesticide concentrations contained in this report are made using pesticide registration toxicity criteria.

Aquatic life criteria, pesticide regulatory criteria, and toxicity (acute and chronic) results for fish, invertebrates, and aquatic plants are presented in Appendix D.

Water Quality Standards for Temperature, pH, and Dissolved Oxygen

Washington State water quality standards for conventional water quality parameters are set forth in Chapter 173-201A of the WAC. Waterbodies are required to meet numeric water quality standards based on the beneficial uses of the waterbody. Conventional parameters including temperature, dissolved oxygen, and pH were measured in this study. Sargeant et al. (2010)

provides a full description of the water quality standards and also explains why parameters such as temperature, pH, and dissolved oxygen are important for fish health.

Numeric Water Quality Standards

Thornton Creek subbasin

Beneficial uses for Thornton Creek are *Core Summer Salmonid Habitat* and *Extraordinary Primary Contact Recreation*. The numeric water quality standards for temperature, dissolved oxygen, and pH in Thornton Creek are described in Table 12.

Table 12. Freshwater water quality standard for temperature, dissolved oxygen, and pH for *Core Summer Salmonid Habitat* use and *Extraordinary Primary Contact Recreation* use.

Parameter	Condition	Value
Temperature	Highest 7- DADMax	16° C. Thornton Creek also has <i>Supplemental Spawning and Incubation</i> criteria: during September 15 - May 15, highest 7-DADMax should not exceed 13° C.
Dissolved Oxygen	Lowest 1-day minimum	9.5 mg/L
pH	--	Range within 6.5 – 8.5, with a human-caused variation within the above range of < 0.2 units.

DADMax: Daily average of the daily maximum temperature.

Longfellow Creek subbasin

Beneficial uses for Longfellow Creek include Salmonid Spawning, Rearing, and Migration habitat and Primary Contact Recreation. The numeric water quality standards for temperature, dissolved oxygen, and pH in Longfellow Creek are described in Table 13.

Skagit-Samish basin

Beneficial uses for the Samish River, Indian Slough, Big Ditch, and Browns Slough are *Salmonid Spawning, Rearing, and Migration Habitat* and *Primary Contact Recreation*. The Samish River, Indian Slough, and Big Ditch sites are freshwater and must meet the water quality standards described in Table 13. The site on Browns Slough is marine water and must meet the water quality standards described in Table 14.

Lower Yakima basin

Beneficial uses for Marion Drain, Sulphur Creek Wasteway, and Spring Creek are *Salmonid Spawning, Rearing, and Migration Habitat*. The freshwater water quality standard described in Table 13 applies to these sites.

Wenatchee-Entiat basins

Beneficial uses for the Mission Creek, Brender Creek, Wenatchee River, and Entiat River are *Salmonid Spawning, Rearing, and Migration*. The water quality standard described in Table 13 applies to these sites.

Table 13. Freshwater water quality standard for temperature, dissolved oxygen, and pH for *Salmonid Spawning, Rearing, and Migration Habitat* use and *Primary Contact Recreation* use.

Parameter	Condition	Value
Temperature	Highest 7- DADMax	17.5° C. The Wenatchee River site also has <i>Supplemental Spawning and Incubation</i> criteria: during October 1 - May 15, highest 7-DADMax should not exceed 13° C.
Dissolved Oxygen	Lowest 1-day minimum	8 mg/L
pH	--	Range within 6.5 – 8.5, with a human-caused variation within the above range of < 0.5 units.

Table 14. Marine water quality standard for temperature, dissolved oxygen, and pH for *Aquatic Life Excellent* use.

Temperature (highest 7- DADMax)	Dissolved Oxygen (lowest 1-day minimum)	pH (must be within the range)
16°C (60.8°F).	6.0 mg/L.	7.0 – 8.5, with a human-caused variation within the above range of < 0.5 units.

Results

This study investigated pesticide occurrence in select salmon bearing surface waters. Results for Thornton Creek in the Cedar-Sammamish basin, Longfellow Creek in the Green-Duwamish basin, the lower Skagit-Samish basin, and the lower Yakima basin are presented as a data summary for 2009. Results for the Wenatchee and Entiat basins include a three-year triennial review (2007-09) with more in-depth data analysis. Monitoring results for all six basins are available through Ecology's EIM system, www.ecy.wa.gov/eim/.

Pesticide calendars are included in Appendix E. The calendars provide a chronological overview of concentrations and detections during 2009. The calendars also compare EPA Pesticide Registration Toxicity Criteria and EPA National Recommended Water Quality Criteria (*assessment criteria*) to numeric Washington State Water Quality Standards (*water quality standards*). Refer to Appendix D, Assessment Criteria and Water Quality Standards, in this report for information on assessment criteria development.

Cedar-Sammamish Basin (WRIA 8): Thornton Creek, 2009

Pesticide Detections and Concentrations

A total of 27 sampling events were conducted on Thornton Creek between March 11 and September 8, 2009. During 2009 there were 58 detections of 19 pesticides and degradates. The 19 pesticides and degradates included: five carbamate insecticides (6 detections), seven herbicides (42 detections), five degradates (6 detections), a wood preservative (3 detections), and a fungicide (1 detection).

The number and types of pesticide detections are presented in Figure 3. The maximum number of pesticides detected during a sampling event was six (Figure 3). The most frequently detected herbicides were dichlobenil and 2,4-D. Most pesticide detections occurred before June 3.

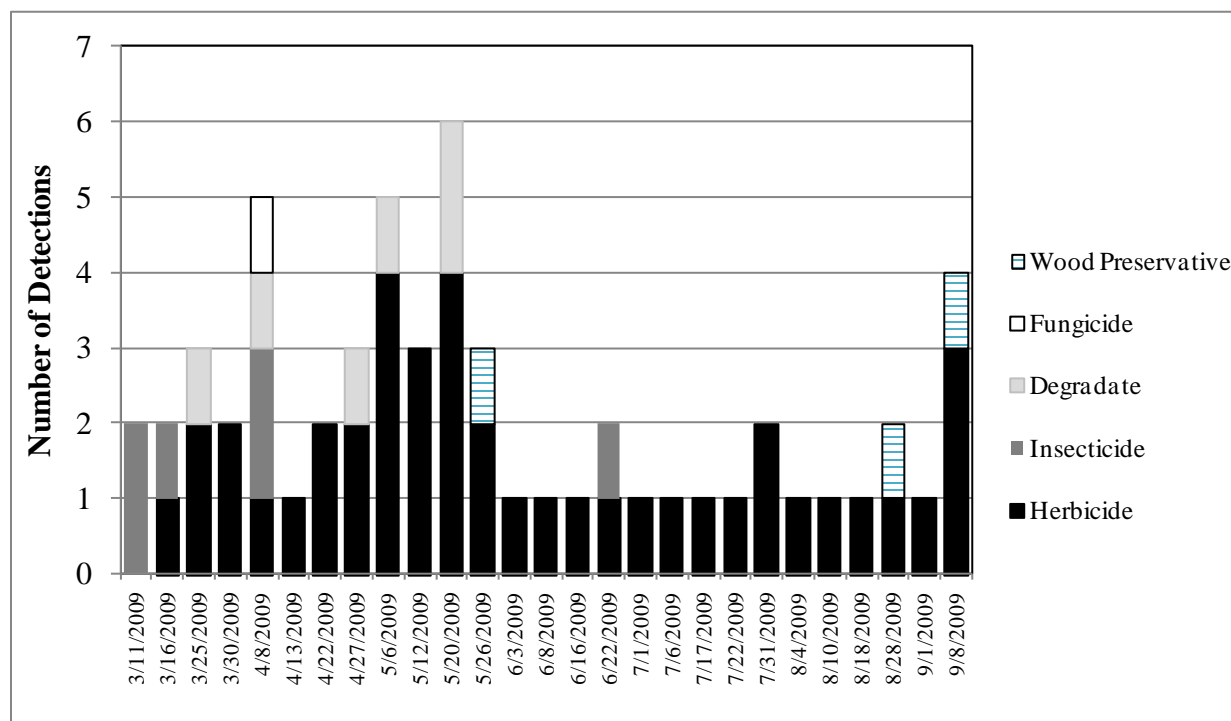


Figure 3. Pesticide detections by week and type in the Thornton Creek subbasin, 2009.

Table 15 presents a pesticide calendar for Thornton Creek. The calendar provides a chronological overview of concentrations and detections during 2009 and compares pesticide concentrations to *assessment criteria* and *water quality standards*. Appendix E, Table E-1 presents the color codes used to compare detected pesticide concentrations to assessment criteria. In 2009 pesticide concentrations in Thornton Creek met (did not exceed) any available *assessment criteria* or *water quality standard* (Appendix D).

Table 15. Thornton Creek pesticide detections, 2009.

Date		3/11	3/16	3/25	3/30	4/8	4/13	4/22	4/27	5/6	5/12	5/20	5/26	6/3	6/8	6/16	6/22	7/1	7/6	7/17	7/22	7/31	8/4	8/10	8/18	8/28	9/1	9/8	
Month		March				April				May				June				July				August				September			
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37	
2,4,6-Trichlorophenol	D-M								0.510																				
2,4-D	H									0.110	0.037	0.130	0.019									0.020						0.040	
3-Hydroxycarbofuran	D-C									0.054		0.076																	
4-Nitrophenol	D-M			0.120																									
Carbaryl	I-C					0.025																							
Carbofuran	I-C					0.031																							
Chlorothalonil	F					0.028																							
Dicamba I	H										0.010																		
Dichlobenil	H		0.023	0.046	0.017	0.010	0.025	0.014	0.012	0.053	0.017	0.049	0.015	0.020	0.020	0.011	0.012	0.014	0.037	0.018	0.014	0.027	0.024	0.030	0.028	0.024	0.026	0.051	
Diuron	H				0.057																								
MCPP	H			0.041						0.042		0.086																	
Methiocarb	I-C	0.099	0.215																										
Methomyl	I-C	0.065																											
Methomyl oxime	D-C											0.079																	
Oxamyl oxime	D-C					0.028																							
Pentachlorophenol	WP												0.007													0.015		0.024	
Prometon	H							0.075	0.039																				
Propoxur	I-C																0.053												
Triclopyr	H									0.080		0.040																	0.044
Total Suspended Solids		3	7	17	5	7	6	6	6	25	4	11	7	10	5	11	4	3	4	4	4	4	3	3	3	2	3	3	4

C – Carbamate, D – Degradate, H – Herbicide, I – Insecticide, M – Multiple, WP – Wood Preservative

Conventional Parameters

Conventional water quality parameters were measured in Thornton Creek. All summaries are based on point (discrete) measurements obtained during the time of sampling. Table 16 summarizes results for TSS, streamflow, pH, conductivity, temperature, and dissolved oxygen.

pH levels met water quality standards during 2009. Dissolved oxygen dropped below the 9.5 mg/L water quality standard seven times during June through August 2009.

Table 16. Mean, minimum, and maximum for discrete conventional parameter measurements for Thornton Creek, 2009.

Parameter	Number	Mean	Minimum	Maximum
Total Suspended Solids (mg/L)	27	6	2	25
Discharge (cfs)	27	6.8	2.6	28.2
pH (s.u.)	26	7.8	7.4	8.7
Conductivity (umhos/cm)	27	220	120	255
Temperature (°C)	27	13.9	5.7	20.2
Dissolved Oxygen (mg/L)	27	10.1	8.7	12.4

In addition to discrete measurements for temperature, continuous (30-minute interval) measurements were collected year-round. A temperature profile based on continuous temperature measurements is presented in Appendix F, Table F-1. During September 15 - May 15, the highest 7-Daily Average Daily Maximum (DADMax) should not exceed 13° C; during the rest of the year, the highest 7-DADMax should not exceed 16°C.

In 2009, temperatures did not meet (exceeded) the standard during the following periods:

- May 12-15, >13°C.
- May 29-June 21, >16°C.
- June 27-September 14, >16°C.
- September 15-30, >13°C.
- October 16-20, >13°C.

Green-Duwamish Basin (WRIA 9): Longfellow Creek, 2009

Pesticide Detections and Concentrations

A total of 27 sampling events were conducted on Longfellow Creek between March 11 and September 8, 2009. During this period, there were 58 detections of nine pesticide compounds. The nine compounds included one carbamate insecticide (2 detections), five herbicides (50 detections), two degradates (2 detections), and a wood preservative (4 detections).

The maximum number of pesticides detected during a sampling event was five, and most of the pesticides detected were herbicide compounds (Figure 4). The most frequently detected herbicides were dichlobenil, triclopyr, and 2,4-D.

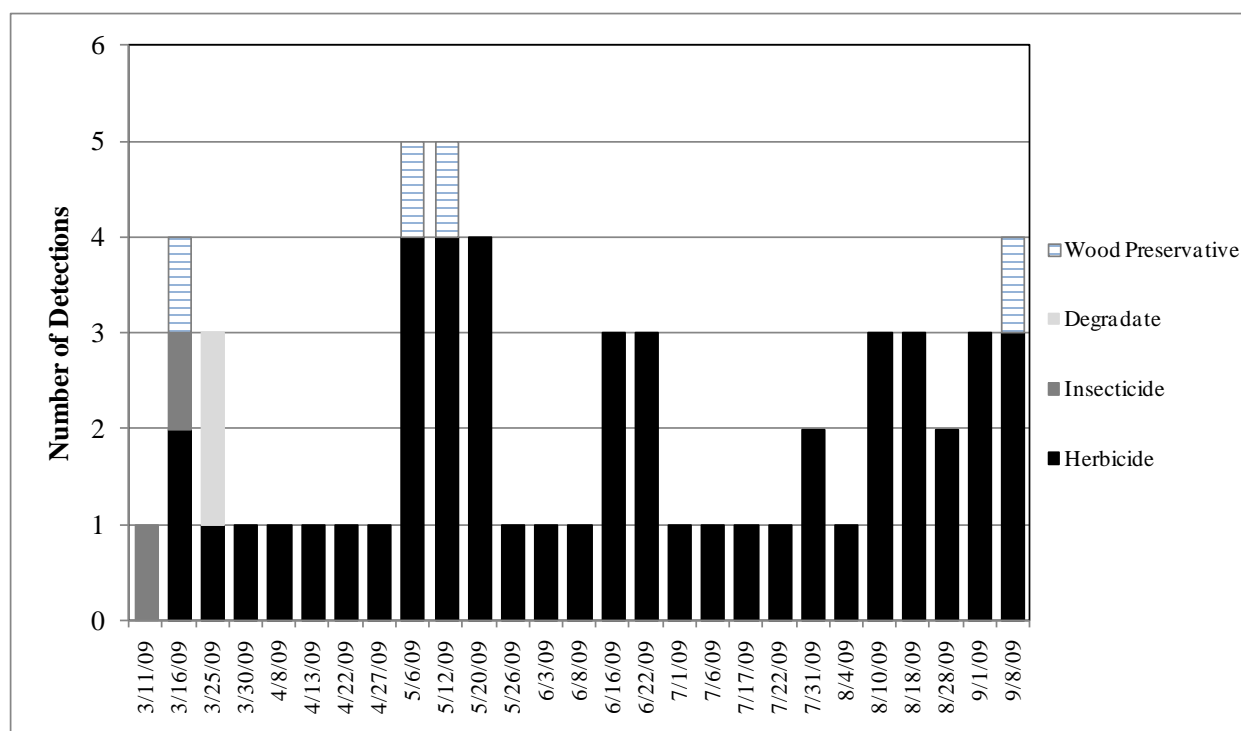


Figure 4. Pesticide detections by week and type in the Longfellow Creek subbasin, 2009.

Table 17 presents a pesticide calendar for Longfellow Creek. The calendar provides a chronological overview of concentrations and detections during 2009 and compares pesticide concentrations to *assessment criteria* and *water quality standards*. Appendix E, Table E-1 presents the color codes used to compare detected pesticide concentrations to *assessment criteria* and *water quality standards*. In 2009, pesticide concentrations in Longfellow Creek met (did not exceed) any available *assessment criteria* and *water quality standards* (Appendix D).

Table 17. Longfellow Creek pesticide detections, 2009.

Date		3/11	3/16	3/25	3/30	4/8	4/13	4/22	4/27	5/6	5/12	5/20	5/26	6/3	6/8	6/16	6/22	7/1	7/6	7/17	7/22	7/31	8/4	8/10	8/18	8/28	9/1	9/8
Month		March				April				May				June				July				August				September		
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
2,4,6-Trichlorophenol	D-M			0.510																								
2,4-D	H									0.110	0.038	0.085				0.058	0.110							0.042	0.022		0.027	0.035
3,5-Dichlorobenzoic Acid	D-H			0.520																								
Dichlobenil	H		0.046	0.010	0.016	0.013	0.047	0.014	0.014	0.130	0.019	0.025	0.013	0.012	0.011	0.008	0.010	0.009	0.022	0.011	0.011	0.023	0.021	0.021	0.030	0.025	0.030	0.033
MCPA	H											0.025																
MCPP	H									0.051	0.009																	
Methiocarb	I-C	0.117	0.200																									
Pentachlorophenol	WP		0.028							0.037	0.009																	0.020
Triclopyr	H		0.095							0.110	0.024	0.071				0.014	0.098					0.015		0.047	0.048	0.034	0.052	0.074
Total Suspended Solids		13	20	3	3	2	7	3	3	38	2	5	4	5	16	4	3	6	3	18	2	2	2	1		3	2	1

C – Carbamate, D – Degradate, H – Herbicide, I – Insecticide, M – Multiple, WP – Wood Preservative

Conventional Parameters

Conventional water quality parameters were measured in Longfellow Creek. Table 18 summarizes results for TSS, streamflow, pH, conductivity, temperature, and dissolved oxygen. All summaries are based on point (discrete) measurements obtained during the time of sampling.

On April 29, 2009, a pH value of 8.7 exceeded the water quality standard of 8.5 standard units (s.u.). During 2009 dissolved oxygen levels met the 8.0 mg/L minimum water quality standard.

Table 18. Mean, minimum, and maximum for discrete conventional parameter measurements for Longfellow Creek, 2009.

Parameter	Number	Mean	Minimum	Maximum
Total Suspended Solids (mg/L)	26	6	1	38
Discharge (cfs)	27	1.6	0.5	12.5
pH (s.u.)	26	7.9	7.1	8.7
Conductivity (umhos/cm)	27	271	110	318
Temperature (°C)	27	14	5.4	20.3
Dissolved Oxygen (mg/L)	27	10.2	8.8	14.3

In addition to discrete temperature measurements, continuous (30-minute interval) measurements were collected year-round. A temperature profile based on continuous temperature measurements is presented in Appendix F, Table F-2. The temperature standard for Longfellow Creek is: the 7-DADMax should not exceed 17.5° C. During 2009 temperature exceeded the standard during the following periods:

- June 1-6, >17.5 °C.
- June 10-13, >17.5 °C.
- July 1-6, >17.5 °C.
- July 13-August 5, >17.5 °C.
- August 17-21, >17.5 °C.

Lower Skagit-Samish Basin (WRIA 3), 2009

Pesticide Detections and Concentrations

The lower Skagit-Samish sites were sampled for 27 consecutive weeks from March 11 to September 8, 2009. The lower Skagit-Samish sites are: upstream and downstream Big Ditch, Indian Slough, Browns Slough, and Samish River. Browns Slough is classified as marine water while the other four sites are classified freshwater.

For the five Skagit-Samish sites combined, there were a total of 33 pesticides including degradates detected: eight insecticides, 20 herbicides, two fungicides, two degradates, and a wood preservative. For the Skagit-Samish sites, Indian Slough and upstream Big Ditch had the greatest number of pesticide detections, with each having 134 detections. The downstream Big Ditch site had 89 detections, Browns Slough had 46 detections, and the Samish River had 20 detections. The downstream Big Ditch site had the greatest number of pesticides detected in one sample: on May 20, 13 pesticides were detected.

Big Ditch

Two sites on Big Ditch were sampled in 2009. Water quality at the upstream site is influenced by industrial land use and stormwater, while the downstream site is influenced by agricultural land use. In 2009, 27 pesticides and degradates were detected in Big Ditch. Eighteen of these were found at the upstream site, and 23 were found at the lower Big Ditch site.

The maximum number of pesticides detected during a sampling event was 10 at the upstream site (Figure 5) and 13 at the downstream site (Figure 6). Most of the pesticides detected were herbicide compounds. At the upstream site, pesticides were detected during every sampling event; at the downstream site, the greatest number of detections occurred from March through May.

The most frequently detected pesticides at the upstream site were:

- Bromacil (herbicide), 25 detections.
- Dichlobenil (herbicide), 24 detections.
- Picloram (herbicide), 16 detections.
- Tebuthiuron (herbicide), 12 detections.
- Imidacloprid (neonicotinoid insecticide), 12 detections.

The most frequently detected pesticides at the downstream site were:

- Metolachlor (herbicide), 14 detections.
- Dichlobenil (herbicide), 13 detections.
- Bromacil (herbicide), 9 detections.
- 2,4-D (herbicide), 8 detections.

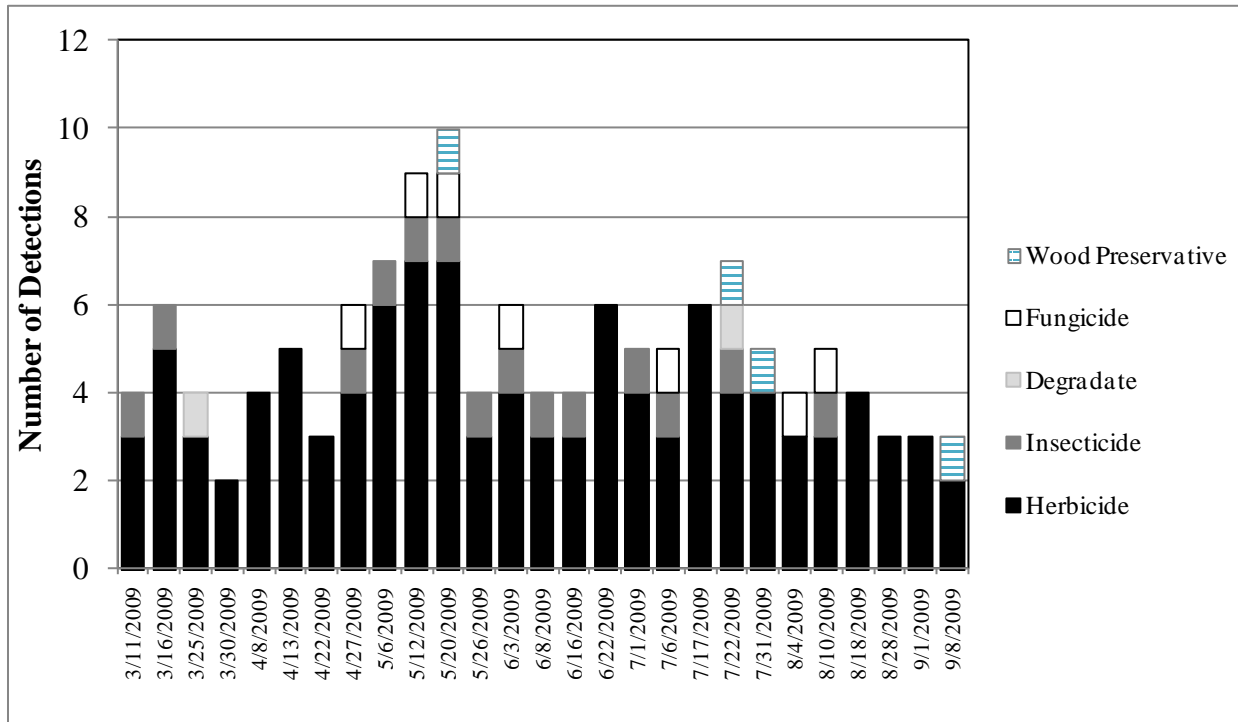


Figure 5. Pesticide detections by week and type for the upstream Big Ditch site, 2009.

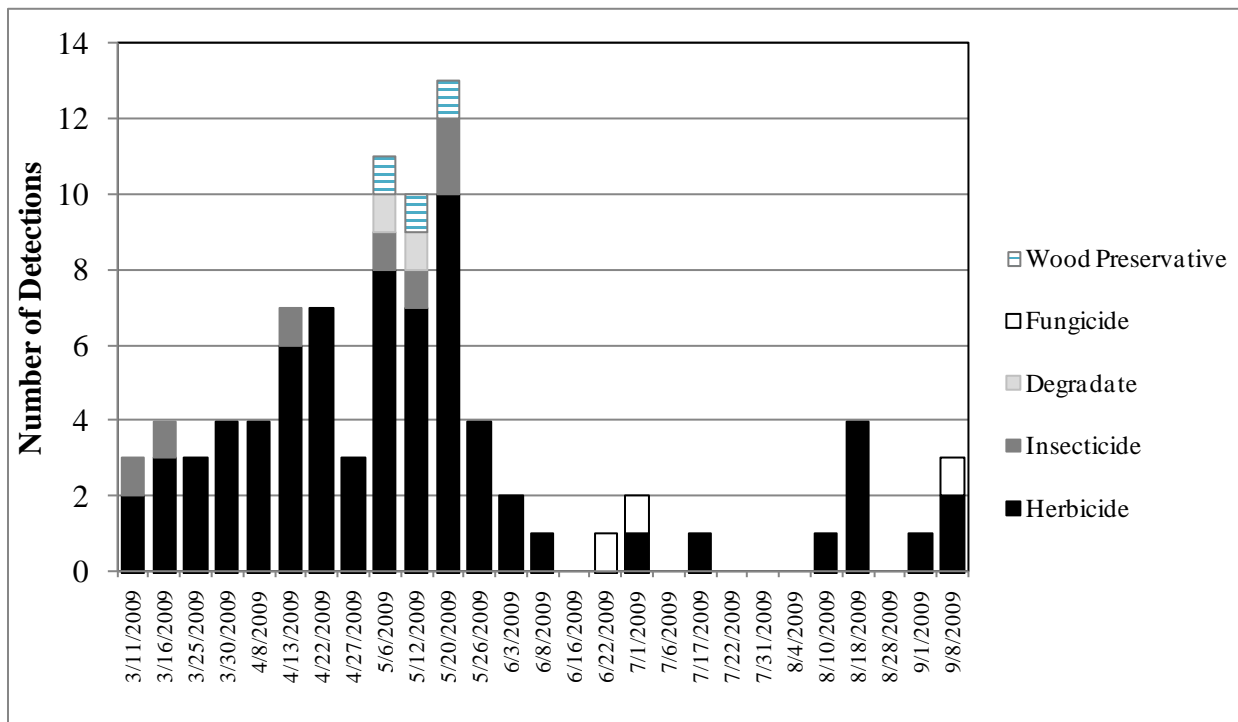


Figure 6. Pesticide detections by week and type for the downstream Big Ditch site, 2009.

Tables E-4 and E-5 in Appendix E present the 2009 pesticide calendars for the upstream and downstream Big Ditch sites, respectively. The calendars provide a chronological overview of concentrations and detections during 2009 and compare pesticide concentrations to *assessment criteria* and *water quality standards*.

The downstream Big Ditch site did not exceed any available *assessment criteria* and *water quality standards* (Appendix D). At the upstream Big Ditch site on May 20, there was one detection of malathion, an organophosphate insecticide, that exceeded the endangered species level of concern (ESLOC) for freshwater fish and the EPA chronic invertebrate criteria. No other detections exceeded any available *assessment criteria* or *water quality standards*.

Indian Slough

During 2009 there were 134 detections of 21 pesticides and a degradate. These 21 compounds were 13 herbicides, five insecticides, one fungicide, one degradate, and a wood preservative. The number and types of pesticide detections are presented in Figure 7. The maximum number of pesticides detected during a sampling event was 10 (Figure 7). Of the 134 pesticide detections, 122 were herbicides.

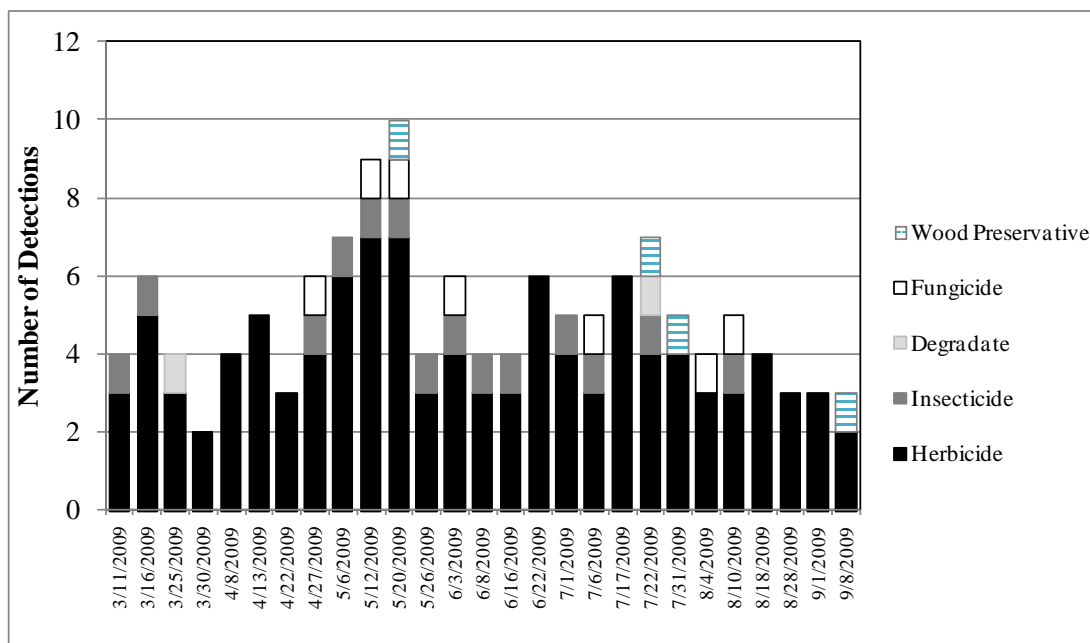


Figure 7. Pesticide detections by week and type for Indian Slough, 2009.

The most frequently detected compounds were the following herbicides:

- Tebuthiuron (19 detections).
- Bromacil (19 detections).
- Dichlobenil (17 detections).
- Diphenamid (16 detections).

During the 2006-08 monitoring, the herbicide diphenamid was detected 52 times in Indian Slough. Diphenamid has not been registered for use by EPA since 1991 (EPA, 2002). It is not known why diphenamid is detected so frequently in Indian Slough. Data quality for herbicide parameters is excellent, and detections are not likely due to field or laboratory error.

Appendix E, Table E-6 presents the 2009 pesticide calendar for Indian Slough. The calendar provides a chronological overview of concentrations and detections during 2009 and compares pesticide concentrations to *assessment criteria* and *water quality standards* (Appendix D).

One detection of malathion at Indian Slough exceeded the ESLOC for freshwater fish and the EPA chronic invertebrate criteria in March 2009. No other pesticide concentrations exceeded available *assessment criteria* or *water quality standards*.

Browns Slough

Browns Slough is sampled downstream of a tidegate. Due to higher salinity at this site marine *assessment criteria* and *water quality standards* are used for evaluating water quality. During 2009 there were 46 detections of 12 pesticides. The 12 pesticides were 10 herbicides, one insecticide, and one wood preservative. The number and types of pesticide detections are presented in Figure 8. The maximum number of pesticides detected during a sampling event was six (Figure 8). Most of the pesticides detected were herbicides (44 out of 46). There were only two pesticide detections after mid-June.

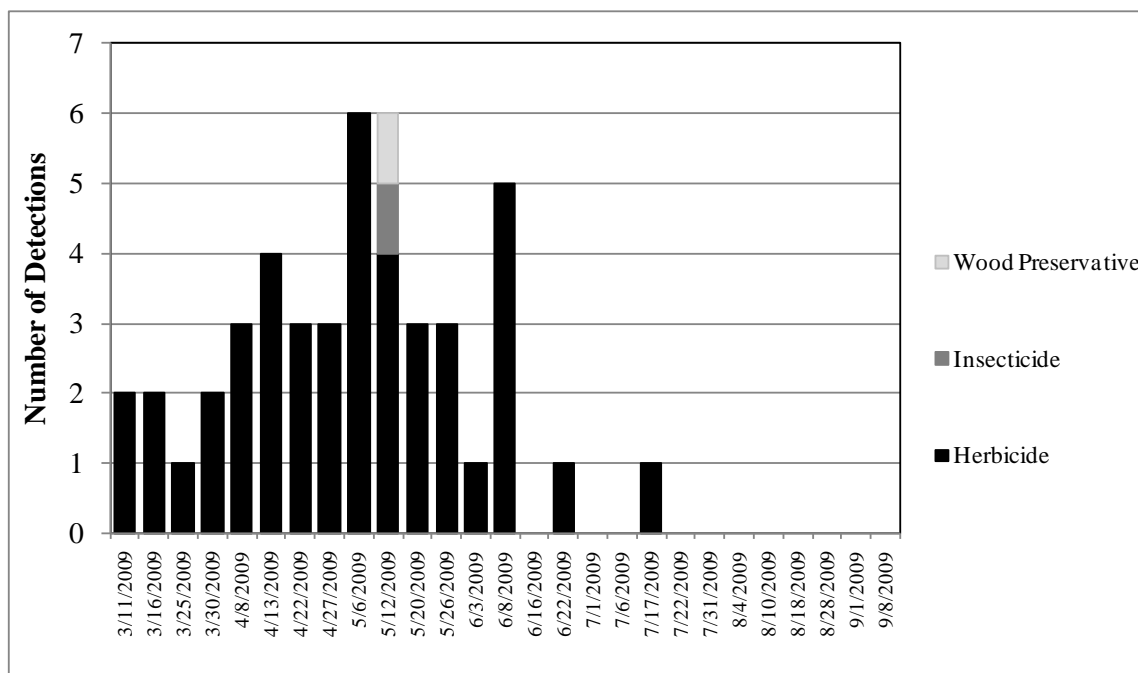


Figure 8. Pesticide detections by week and type for Browns Slough, 2009.

The most frequently detected pesticides were the following herbicides:

- DCPA (dacthal) (13 detections).
- Metolachlor (7 detections).
- Simazine (7 detections).

Appendix E, Table E-7 presents the 2009 pesticide calendar for Browns Slough. The calendar provides a chronological overview of concentrations and detections during 2009 and compares pesticide concentrations to *assessment criteria* and *water quality standards*. In 2009 pesticide concentrations in Browns Slough did not exceed any available marine *assessment criteria* or *water quality standards* (Appendix D).

Samish River

A total of 27 sampling events were conducted on the Samish River between March 11 and September 8, 2009. There were very few pesticide detections (20 detections); these were seven herbicides and a wood preservative. The number and types of pesticide detections are presented in Figure 9. The maximum number of pesticides detected during a sampling event was six (Figure 9). The most commonly detected pesticide was the herbicide dichlobenil, with six detections.

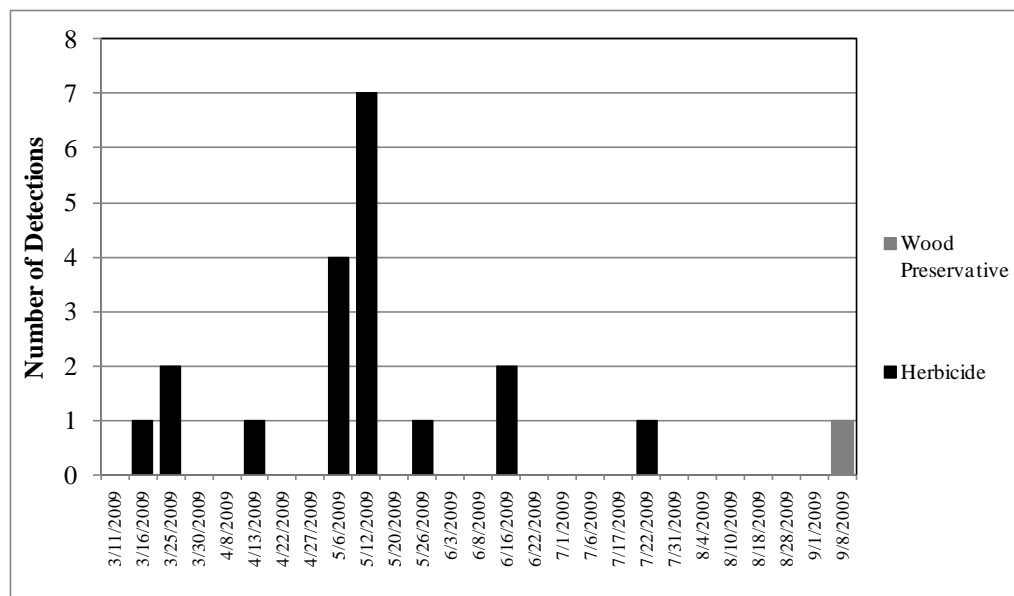


Figure 9. Pesticide detections by week and type for the Samish River, 2009.

Appendix E, Table E-8 presents a pesticide calendar for the Samish River. The calendar provides a chronological overview of concentrations and detections during 2009 and compares pesticide concentrations to *assessment criteria* and *water quality standards*. Samish River did not exceed any available *assessment criteria* or *water quality standards* (Appendix D).

Conventional Parameters

Conventional water quality parameters were measured at the Skagit-Samish sites. Table 19 summarizes results for TSS, streamflow, pH, conductivity, and dissolved oxygen. All summaries are based on point (discrete) measurements obtained during the time of sampling. Browns Slough is a marine site and must meet marine water quality standards; all the other Skagit-Samish sites must meet freshwater quality standards.

Table 19. Mean, minimum, and maximum for discrete conventional parameter measurements for the Skagit-Samish sites, 2009.

Site	Total Suspended Solids (mg/L)	Flow (cfs)	pH (s.u.)	Conductivity (umhos/cm)	Dissolved Oxygen (mg/L)
Big Ditch (upstream)					
number	27	25	26	27	27
mean	17	2.6	7.0	315	8.8
minimum	3	0.4	6.7	135	6.1
maximum	118	13	7.5	426	11.6
Big Ditch (downstream)					
number	24	27	26	27	27
mean	12	11.8	7.3	344	9.5
minimum	1	1.7	6.7	44	6.3
maximum	38	23.6	9.4	933	14.3
Indian Slough					
number	26	25	26	27	27
mean	8	15.1	7.0	937	7.1
minimum	2	1.3	6.7	261	5.2
maximum	23	31.0	7.7	2296	9.6
Brown Slough					
number	27	16	26	27	27
mean	9	4.3	7.5	13245	10.1
minimum	4	0.5	7.0	918	2.9
maximum	18	6.6	8.7	30450	16.4
Samish River					
number	27	26	26	27	27
mean	13	166.5	7.2	122	10.3
minimum	2	26.5	6.7	53	8.8
maximum	89	699.4	7.6	442	12.8

During 2009 dissolved oxygen levels did not meet the 8.0 mg/L minimum fresh water quality standard in upper Big Ditch (9 times), lower Big Ditch (7 times), and Indian Slough (22 times). The Samish River met dissolved oxygen water quality standards during all sampling events. Browns Slough did not meet the 6.0 mg/L minimum marine water quality standard for two sampling events.

Upper Big Ditch, Indian Slough, and Samish River met pH water quality standards. Both Browns Slough (marine) and lower Big Ditch (freshwater) did not meet the pH standard once during the sample period.

In addition to the discrete temperature measurements, continuous (30-minute interval) measurements were collected year-round. A temperature profile based on continuous temperature measurements is presented in Appendix F, Figures F-3 – F-7. The temperature standard for the freshwater sites is the 7-DADMax should not exceed 17.5° C; and for the marine water it should not exceed 16.0° C. Table 20 describes the periods that temperature exceeded the standard.

Table 20. Periods of water temperature exceedance for the Skagit-Samish basin sites, 2009.

Site	Periods When Temperature Did Not Meet Standards
Big Ditch (upstream) >17.5°C	May 29-June 18, June 20-21, June 24-July 8, July 11-Sept 20.
Big Ditch (downstream) >17.5°C	April 28-May 1, May 18-June 4, June 10-12, June 28-Sept 25.
Indian Slough >17.5°C	May 25-Sept 23.
Browns Slough >16.0°C	April 18-May 3, May 5-Sept 25, Oct 2-3.
Samish River >17.5°C	June 15-20, June 29-July 4, July 13-Aug 6, Aug 28-Sept 2.

Lower Yakima Basin (WRIA 37), 2009

Pesticide Detections and Concentrations

In the lower Yakima River basin, downstream Spring Creek, Marion Drain, and Sulphur Creek Wasteway were sampled for 27 consecutive weeks from March 11 to September 9, 2009. The upstream Spring Creek site was sampled every other week during the same period for a total of 14 sampling events. In Marion Drain, weekly sampling for organophosphates continued from September 16 through October 26, 2009. Historically, Marion Drain sampling for organophosphates has continued through the end of October.

For the four sites combined, there were a total of 35 types of pesticides and degradates found. These 35 compounds were 11 insecticides, 20 herbicides, three degradates, and one wood preservative. Marion Drain had the greatest number of detections, 150 (and greatest number of sampling events). Sulphur Creek Wasteway had 124 pesticide detections followed by the downstream Spring Creek site (88 detections) then the upstream Spring Creek site had 42 detections (and the least number of sampling events).

Spring Creek

Two sites on Spring Creek were sampled in 2009. The upstream site was sampled every two weeks, and the downstream site weekly. A total of 19 pesticide and degradate types were detected in Spring Creek: 11 herbicides, five insecticides, two degradates, and one wood preservative. Similar pesticides were detected at both sites with 15 of the pesticides found upstream and 18 found at the downstream site. The number and types of pesticide detections are presented in Figure 10 for the upstream site and Figure 11 for the downstream site. The maximum number of pesticides detected during a sampling event at the upstream site was six (Figure 10) and at the downstream site was eight (Figure 11).

The most frequently detected pesticides at the upstream site were the following herbicides:

- 2,4-D (5 detections).
- Atrazine (5 detections).
- Bentazon (5 detections).

The most frequently detected pesticides at the downstream site were the following herbicides:

- 2,4-D (18 detections).
- Bromacil (15 detections).
- Pendimethalin (9 detections).

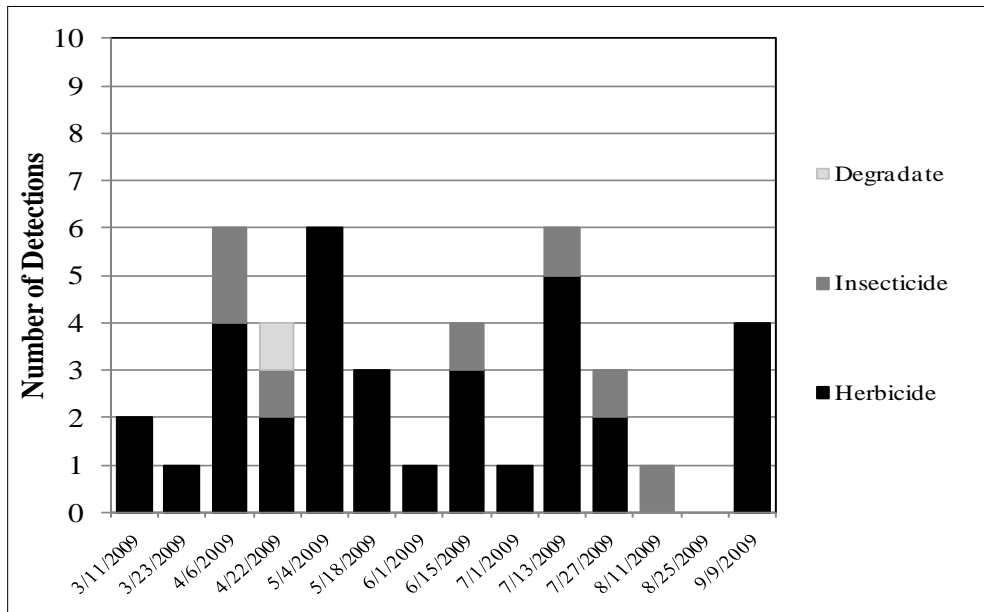


Figure 10. Pesticide detections by week and type for upstream Spring Creek, 2009.

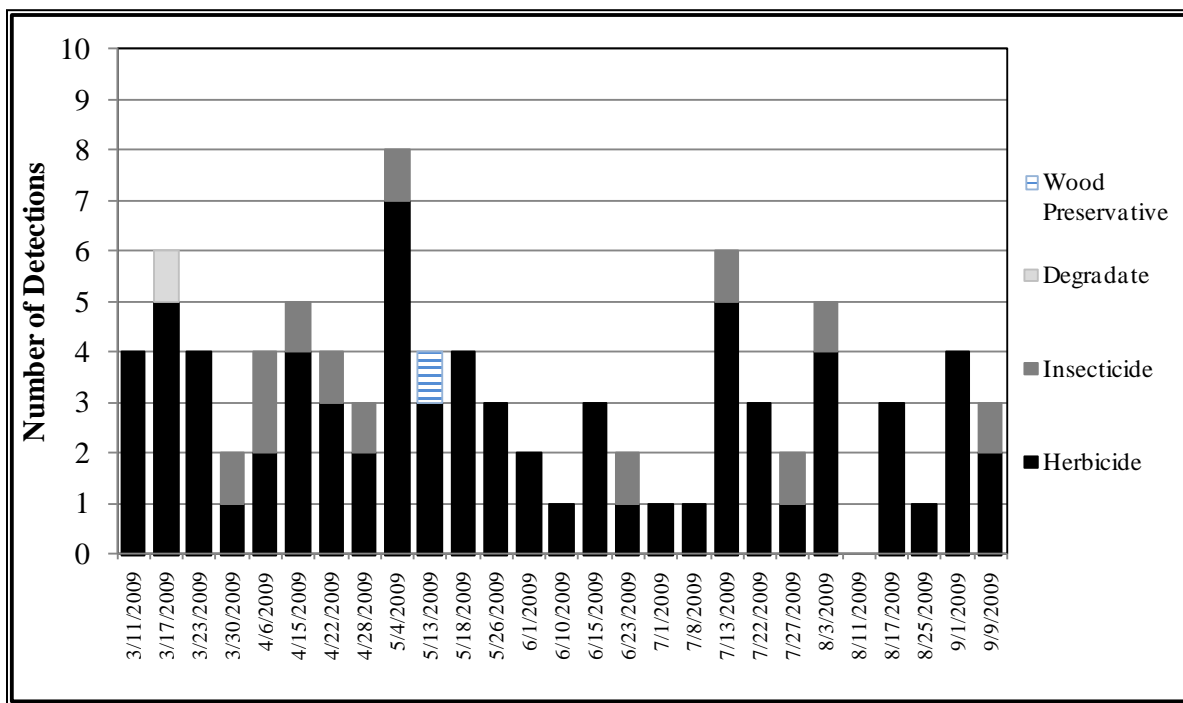


Figure 11. Pesticide detections by week and type for downstream Spring Creek, 2009.

Appendix E, Tables E-9 and E-10 present the 2009 pesticide calendars for the upstream and downstream Spring Creek sites, respectively. The calendars provide a chronological overview of concentrations and detections during 2009 and compare pesticide concentrations to *assessment criteria* and *water quality standards* (Appendix D).

At the upstream site, one April sample of 4,4'-DDE did not meet (was above) chronic water quality standards.

At the downstream site, three consecutive detections of chlorpyrifos from March to April were above the chronic *water quality standard* and EPA's chronic invertebrate *criteria*. Three consecutive weeks of detections mean that the four-day time component of the *water quality standard* was not met.

Marion Drain

An additional seven weeks of sampling for organophosphates was conducted on Marion Drain after September 9, 2009. No pesticides were detected during the last two sampling events at the end of October. During 2009 there were 150 detections of 19 pesticides and a degradate. These 19 compounds were 14 herbicides, five insecticides, and one insecticide degradate. The number and types of pesticide detections are presented in Figure 12. The maximum number of pesticides detected during a sampling event was nine (Figure 12). A total of 75% of the detections were herbicides; the rest were insecticides or insecticide degradates.

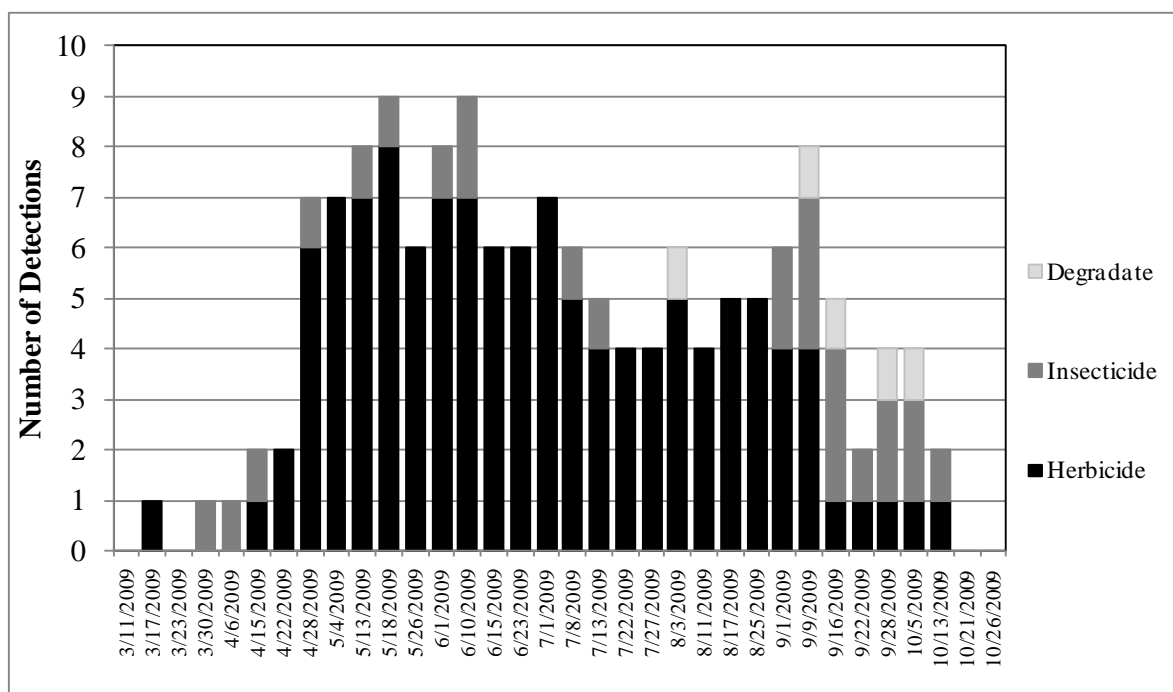


Figure 12. Pesticide detections by week and type for Marion Drain, 2009.

The most frequently detected compounds were herbicides:

- Terbacil (26 detections).
- 2,4-D (19 detections).
- Dicamba I (18 detections).
- Bentazon (15 detections).

The most frequently detected insecticide was chlorpyrifos with 10 detections during the March through October sampling period.

The 2009 pesticide calendar in Appendix E, Table E-11 provides a chronological overview of concentrations and detections during 2009 and compares pesticide concentrations to *assessment criteria* and *water quality standards* (Appendix D).

One detection of chlorpyrifos was found at the EPA chronic invertebrate criteria once in 2009. This single event did not exceed the 21-day time component of the chronic invertebrate criteria.

Sulphur Creek Wasteway

During 2009 there were 124 detections of 21 pesticides and degradates. These 21 compounds were 15 herbicides, five insecticides, and one insecticide degradate. The number and types of pesticide detections are presented in Figure 13. The maximum number of pesticides detected during a sampling event was eight (Figure 13).

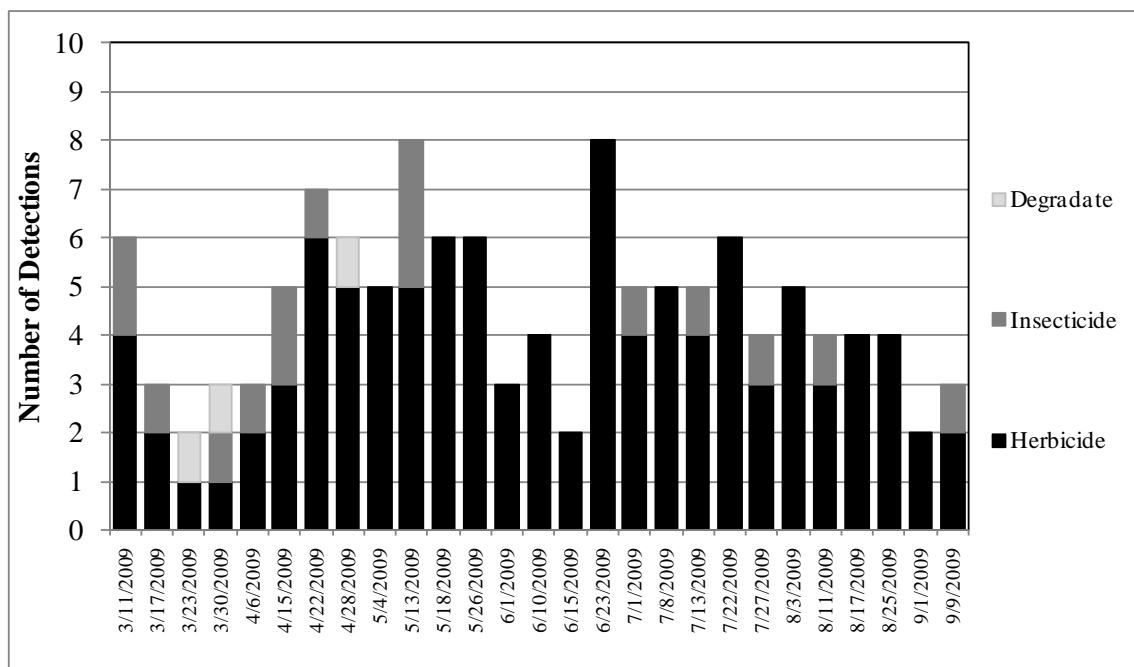


Figure 13. Pesticide detections by week and type for Sulphur Creek Wasteway, 2009.

The most frequently detected compounds were the following herbicides:

- 2,4-D (21 detections).
- Bromacil (18 detections).
- Dicamba I (18 detections).
- Dichlobenil (9 detections).

The 2009 pesticide calendar in Appendix E, Table E-12 provides a chronological overview of concentrations and detections during 2009 and compares pesticide concentrations to *assessment criteria* and *water quality standards* (Appendix D).

4,4'-DDE did not meet the chronic *water quality standard* in March and April. The chronic *water quality standard* for DDT and its metabolites were exceeded due to consistent weekly detections.

There were three consecutive detections of chlorpyrifos above the chronic *water quality standard*; one detection was also above the ESLOC for fish. Because chlorpyrifos was detected in three consecutive weeks, the chronic *water quality standard* was not met.

Conventional Parameters

Conventional water quality parameters were measured at the four lower Yakima sites. Table 21 summarizes results for TSS, streamflow, pH, conductivity, and dissolved oxygen. All summaries are based on point (discrete) measurements obtained during the time of sampling. All sites must meet freshwater quality standards.

With the exception of the upstream Spring Creek site, all sites exceeded the pH water quality standard of 8.5 s.u. The downstream Spring Creek site exceeded the standard 11 times, Marion Drain four times, and Sulphur Creek Wasteway seven times. Maximum pH values are described in Table 21.

All sites met the dissolved oxygen standard with the exception of the upstream Spring Creek site. On July 13, a dissolved oxygen value of 7.9 fell below the minimum water quality standard of 8.0 mg/L.

In addition to the discrete temperature measurements, continuous (30-minute interval) measurements were collected year-round. A temperature profile based on continuous temperature measurements is presented in Appendix F, Figures F-8 – F-11. The temperature standard for the freshwater sites is that the 7-DADMax should not exceed 17.5°C. None of the sites met temperature standards during all periods. Table 22 describes the periods that temperature did not meet the standard.

Table 21. Mean, minimum, and maximum for discrete conventional parameter measurements for the lower Yakima sites, 2009.

Site	Total Suspended Solids (mg/L)	Flow (cfs)	pH (s.u.)	Conductivity (umhos/cm)	Dissolved Oxygen (mg/L)
Spring Creek (upstream)					
number	14	14	14	14	14
mean	22	6.4	8.0	366	9.4
minimum	4	1.9	7.5	266	7.9
maximum	68	11.9	8.5	502	12.1
Spring Creek (downstream)					
number	27	27	27	27	27
mean	14	8.7	8.6	338	9.9
minimum	1	1	7.9	180	8.1
maximum	50	17	9.4	478	13.6
Marion Drain					
number	34	32	34	34	34
mean	13	116	8.1	226	12.5
minimum	2	12.7	7.5	138	9.3
maximum	40	265	9.3	341	17.6
Sulphur Creek Wasteway					
number	27	27	27	27	27
mean	40	260	8.4	264	10.3
minimum	7	49	7.8	164	9.1
maximum	98	641	8.8	535	12.1

Table 22. Periods of water temperature exceedance for the lower Yakima sites, 2009.

Site	Periods When Temperature Did Not Meet Standards
Spring Creek (upstream) >17.5°C	May 20-Sept 6, Sept 11-17
Spring Creek (downstream) >17.5°C	April 18-23, May 13-Sept 23
Marion Drain >17.5°C	May 25-Aug 13
Sulphur Creek Wasteway >17.5°C	May 17-Sept 26

Wenatchee-Entiat Basin (WRIAs 45 and 46), 2007-09

This report includes the triennial (2007-09) monitoring summary for the Wenatchee and Entiat basins. In the most recent three-year (2006-08) summary for all other project areas (Sargeant et al., 2010), the Wenatchee and Entiat basins were not included due to having only two years of monitoring data for that three-year period. With the additional data obtained in 2009, the Wenatchee-Entiat three-year summary is instead included within this report.

Sampling of the Wenatchee-Entiat basins began in 2007 as described in Dugger et al. (2007). Sampling is conducted at five sites in the Wenatchee-Entiat:

- Peshastin Creek at river mile 0.1.
- Mission Creek at river mile 3.1.
- Brender Creek at river mile 0.7.
- Wenatchee River at river mile 2.8.
- Entiat River at river mile 1.4.

Figure 1 presents the locations of the sampling sites. Appendix B describes sampling locations and duration of sampling for each site.

Peshastin Creek

Pesticide Detections and Concentrations

During 2007-09 very few pesticides were detected: 14 detections of 12 pesticides and degradates (Table 23).

Comparison to Assessment Criteria and Water Quality Standards

Pesticide calendars in Appendix E, Tables E-13-15, provide a chronological overview of concentrations and detections during 2007-09 and compare pesticide concentrations to *assessment criteria* and *water quality standards*.

During 2007-09 there was one detection of endosulfan that did not meet (was above) the ESLOC criteria for fish in 2008 and again in 2009. A single detection of azinphos-methyl was above the chronic NRWQC in 2007.

Table 23. Summary of pesticide detections (ug/L) in Peshastin Creek, 2007-09.

Pesticide Name and Type	ALPQL	2007 n = 31			2008 n = 27			2009 n = 27		
		#Det	Freq	Max	#Det	Freq	Max	#Det	Freq	Max
Insecticides										
Azinphos Methyl (Organophosphate)	0.038	1	3.2%	0.024	ND			ND		
Carbaryl (Carbamate)	0.019	1	3.2%	0.019	ND			ND		
Endosulfan I (Organochlorine)	0.050	ND			1	3.7%	0.130	2	7.4%	0.040
Endosulfan II (Organochlorine)	0.050	ND			1	3.7%	0.046	ND		
Methomyl (Carbamate)	0.046	1	3.2%	0.023	ND			ND		
Oxamyl (Carbamate)	0.047	1	3.2%	0.026	1	3.7%	0.010	ND		
Degradates										
Fipronil Sulfide (Pyrethroid)	0.100	MEL added analysis in 2009						1	3.7%	0.015
Fipronil Sulfone (Pyrethroid)	0.101	MEL added analysis in 2009						1	3.7%	0.016
Oxamyl Oxime (Carbamate)	0.019	1	3.2%	0.012	ND			ND		
Herbicides										
Simazine	0.033	ND			ND			1	3.7%	0.014
Simetryn	0.100	ND			ND			1	3.7%	0.055

ALPQL: Average lower practical quantitation limit. ND: Not detected.

Pesticide Distribution

As with the majority of the Wenatchee-Entiat sites, insecticides were the most frequently detected pesticide in Peshastin Creek. Figure 14 presents the distribution of detections by pesticide group for 2007-09 detections. Seven types of insecticides were detected. Endosulfan was most frequently observed with three detections during the three-year period.

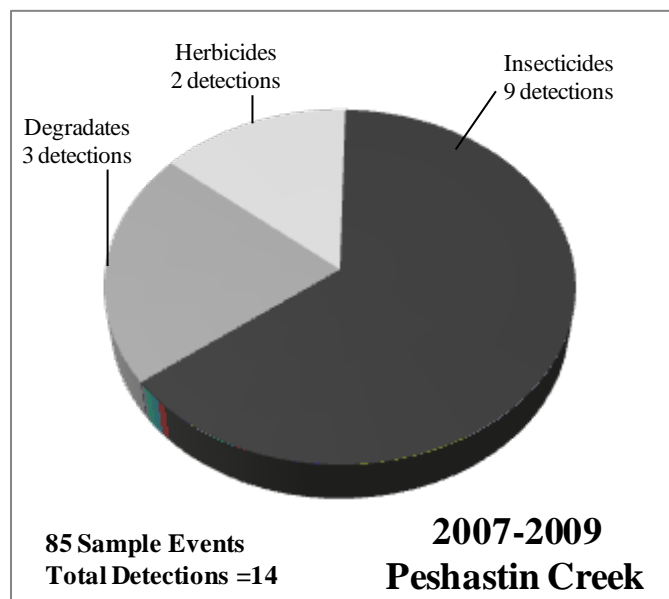


Figure 14. Pesticide distribution in Peshastin Creek, 2007-09.

During 2007-09 no co-occurrence of insecticides with the same mode of action (acetylcholinesterase inhibition) occurred during a sampling event.

Mission Creek

Pesticide Detections and Concentrations

During 2007-09 very few pesticides were detected: 19 detections of 11 pesticides and degradates (Table 24).

Table 24. Summary of pesticide detections (ug/L) in Mission Creek, 2007-09.

Pesticide Name and Type	ALPQL	2007 n =31			2008 n =27			2009 n =27			
		#Det	Freq	Max	#Det	Freq	Max	#Det	Freq	Max	
Insecticides											
Carbaryl (Carbamate)	0.019	ND			1	3.7%	0.014	ND			
Chlorpyrifos (Organophosphate)	0.033	1	3.2%	0.024	ND			ND			
Endosulfan I (Organochlorine)	0.050	1	3.2%	0.017	1	3.7%	0.047	1	3.7%	0.024	
Endosulfan II (Organochlorine)	0.050	1	3.2%	0.022	ND			ND			
Methiocarb (Carbamate)	0.019	2	6.5%	0.034	ND			ND			
Methomyl (Carbamate)	0.046	1	3.2%	0.019	ND			ND			
Degradates											
3-Hydroxycarbofuran (Carbamate)	0.047	ND			ND			1	3.7%	0.051	
Oxamyl Oxime (Carbamate)	0.019	2	6.5%	0.018	ND			ND			
Synergist											
Piperonyl Butoxide	0.100	MEL added analysis in 2009					1	3.7%	0.095		
Herbicides											
Norflurazon	0.033	2	6.5%	0.041	3	11%	0.034	ND			
Simazine	0.033	ND			1	3.7%	0.019	ND			

ALPQL: Average lower practical quantitation limit. ND: Not detected.

Comparison to Assessment Criteria and Water Quality Standards

Pesticide calendars in Appendix E, Tables E-16-18, provide a chronological overview of concentrations and detections during 2007-09 and compare pesticide concentrations to *assessment criteria* and *water quality standards* (Appendix D).

During 2007-09 there was one detection of endosulfan not meeting (above) the ESLOC criteria for fish in 2008. No other detections were above *assessment criteria* or *water quality standards*.

Pesticide Distribution

As with the majority of the Wenatchee-Entiat sites, *insecticide* was the most frequently detected type of pesticide at Mission Creek. Figure 15 presents the distribution of detections by pesticide group. Six types of insecticides were detected. Endosulfan was detected most frequently with three detections during the three-year period.

During 2007-09 no co-occurrence of insecticides with the same mode of action (acetylcholinesterase inhibition) occurred during a sampling event.

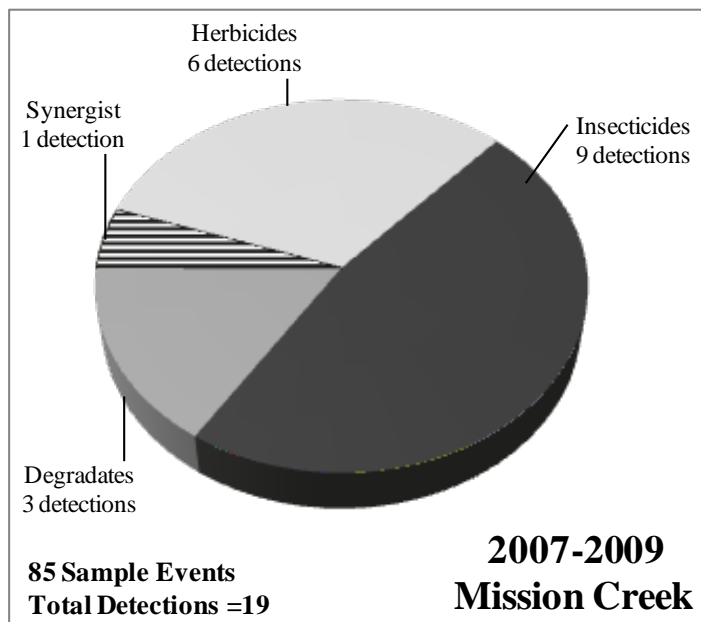


Figure 15. Pesticide distribution in Mission Creek, 2007-09.

Brender Creek

Pesticide Detections and Concentrations

During 2007-09 Brender Creek had the greatest number of pesticide detections of the Wenatchee-Entiat sites. Brender Creek also has the lowest streamflows of any of the sites; this makes detection of pollutants more likely. Table 25 summarizes pesticide detections. During the three years of sampling, there were 401 detections of 28 pesticides, degradates, and a synergistic compound.

Comparison to Assessment Criteria and Water Quality Standards

Pesticide calendars in Appendix E, Tables E-19-21, provide a chronological overview of concentrations and detections during 2007-09 and compare pesticide concentrations to *assessment criteria* and *water quality standards* (Appendix D).

Endosulfan was detected above the ESLOC for rainbow trout between March and May in six samples in 2007, eight samples in 2008, and four samples in 2009. Each year showed consecutive detections in two or more weeks indicating chronic exceedance.

Table 25. Summary of pesticide detections (ug/L) in Brender Creek, 2007-09.

Pesticide Name and Type	ALPQL	2007			2008			2009		
		n = 30			n = 27			n = 27		
		#Det	Freq	Max	#Det	Freq	Max	#Det	Freq	Max
Insecticides										
2,4'-DDT (Organochlorine)	0.033	7	23%	0.017	2	7.4%	0.053	2	7.4%	0.019
4,4'-DDT (Organochlorine)	0.033	27	90%	0.050	26	96.3%	0.300	20	74%	0.037
Azinphos Methyl	0.038	3	10%	0.530	ND			ND		
Carbaryl (Carbamate)	0.019	4	13%	0.040	1	3.7%	0.024	ND		
Chlorpyrifos (Organophosphate)	0.033	9	30%	0.110	5	18.5%	0.028	6	22%	0.083
Diazinon (Organophosphate)	0.033	1	3.3%	0.021	ND			ND		
Endosulfan I (Organochlorine)	0.050	7	23%	0.100	5	18.5%	0.089	5	19%	0.100
Endosulfan II (Organochlorine)	0.050	7	23%	0.074	8	29.6%	0.120	6	22%	0.058
Imidacloprid (Neonicotinoid)	0.020	MEL added analysis in 2008			2	7.4%	0.060	1	3.7%	0.022
Methiocarb (Carbamate)	0.019	ND			ND			1	3.7%	0.033
Methomyl (Carbamate)	0.046	1	3.3%	0.017	ND			ND		
Oxamyl (Carbamate)	0.047	1	3.3%	0.027	ND			ND		
Degradates										
2,4'-DDD (Organochlorine)	0.033	2	6.7%	0.018	1	3.7%	0.015	ND		
2,4'-DDE (Organochlorine)	0.033	ND			ND			2	7.4%	0.009
3-Hydroxycarbofuran (Carbamate)	0.047	ND			ND			1	3.7%	0.106
4,4'-DDD (Organochlorine)	0.033	16	53%	0.025	20	74.1%	0.025	13	48%	0.030
4,4'-DDE (Organochlorine)	0.033	29	97%	0.071	22	81.5%	0.045	25	93%	0.047
Endosulfan Sulfate (Organochlorine)	0.033	17	57%	0.100	24	88.9%	0.160	21	78%	0.098
Oxamyl Oxime (Carbamate)	0.019	ND			1	3.7%	0.140	ND		
Fungicide										
Triadimefon	0.033	1	3.3%	0.015	ND			ND		
Synergist										
Piperonyl Butoxide	0.100	MEL added analysis in 2009						1	3.7%	0.070
Herbicides										
Dicamba I	0.063	ND			ND			1	3.7%	0.012
Dichlobenil	0.032	ND			1	3.7%	0.008	10	37%	0.030
Diuron	0.056	1	3.3%	0.120	2	7.4%	0.220	ND		
MCPA	0.063	1	3.3%	0.072	ND			ND		
Norflurazon	0.033	10	33%	0.160	10	37.0%	0.250	7	26%	0.048
Prometon	0.033	1	3.3%	0.009	ND			ND		
Simazine	0.033	2	6.7%	0.028	1	3.7%	0.012	1	3.7%	0.096

ALPQL: Average lower practical quantitation limit. ND: Not detected.

Three detections of chlorpyrifos in March and April of 2007 and 2009 did not meet (were above) the chronic *water quality standard*, and in 2009 detections in two consecutive weeks were above the chronic *water quality standard*. Consecutive weekly detections above numeric criteria indicate the four-day exposure criteria for chronic *water quality standard* was exceeded.

DDT and DDT degradates were found consistently throughout all three years, except for one week in April 2008 and two weeks in May and June 2009. All total DDT concentrations were above the chronic *water quality standard*.

Pesticide Distribution

Unlike the majority of the Wenatchee-Entiat sites, insecticide degradates were the most frequently detected compound in Brender Creek. Figure 16 presents the distribution of detections by pesticide type.

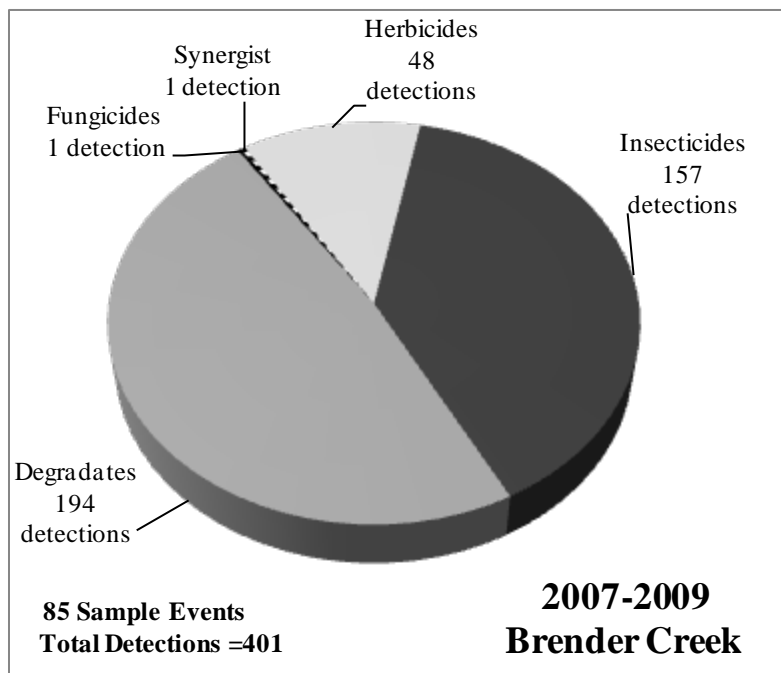


Figure 16. Pesticide distribution in Brender Creek, 2007-09.

The most commonly detected insecticide degradates were:

- 4,4'-DDE (76 detections), a degradate of the legacy pesticide DDT.
- Endosulfan sulfate (62 detections), a degradate of endosulfan.
- 4,4'-DDD (49 detections), degradates of the legacy pesticide DDT.

After degradates, insecticides were the most commonly observed pesticide. Six types of insecticides were detected. The most commonly detected insecticides were:

- 4,4'-DDT (73 detections), a legacy pesticide DDT.
- Endosulfan II (21 detections).
- Chlorpyrifos (20 detections).
- Endosulfan I (17 detections).

The most commonly detected herbicides were:

- Norflurazon (27 detections).
- Dichlobenil (11 detections).

Figure 17 presents Brender Creek carbamate and organophosphate insecticide detections for 2007-09. Carbamate and organophosphate insecticides are both acetylcholinesterase-inhibiting insecticides; they have the same mode of action. When these insecticides co-occur, the effect on aquatic life may be additive or in some cases synergistic (Laetz et al., 2009). In Figure 17 when two pesticides co-occur, the cumulative totals are presented.

During 2007-09 co-occurrence of acetylcholinesterase-inhibiting insecticides occurred four times in 2007, but there was no co-occurrence in 2008 and 2009 (Figure 17.). The greatest cumulative sum of co-occurring acetylcholinesterase-inhibiting insecticides was 0.047 ug/L on May 21, 2007 for a carbamate (carbaryl) and an organophosphate (chlorpyrifos).

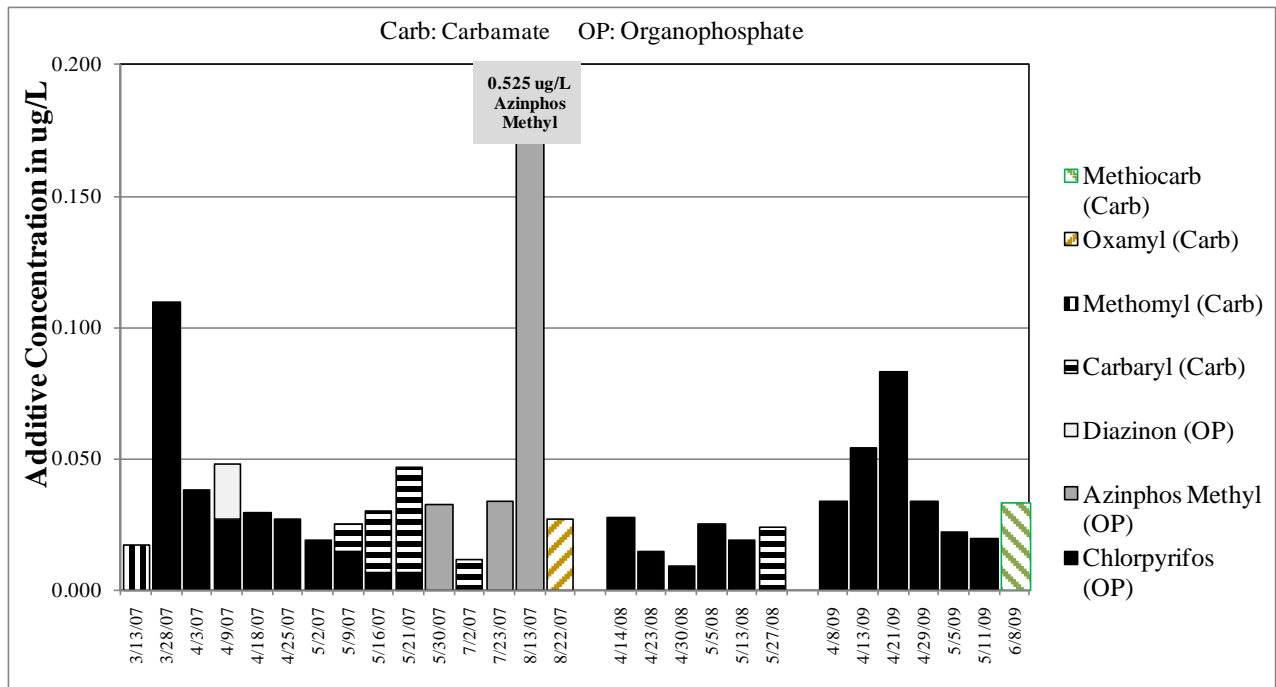


Figure 17. Cumulative amount for acetylcholinesterase-inhibiting insecticides (same mode of action) in Brender Creek, 2007-09.

Due to the consistent presence of the legacy DDT and degradate compounds, co-occurrence of organochlorine insecticides occurred frequently. Figure 18 presents cumulative concentrations for total DDT and total endosulfan, both organochlorines.

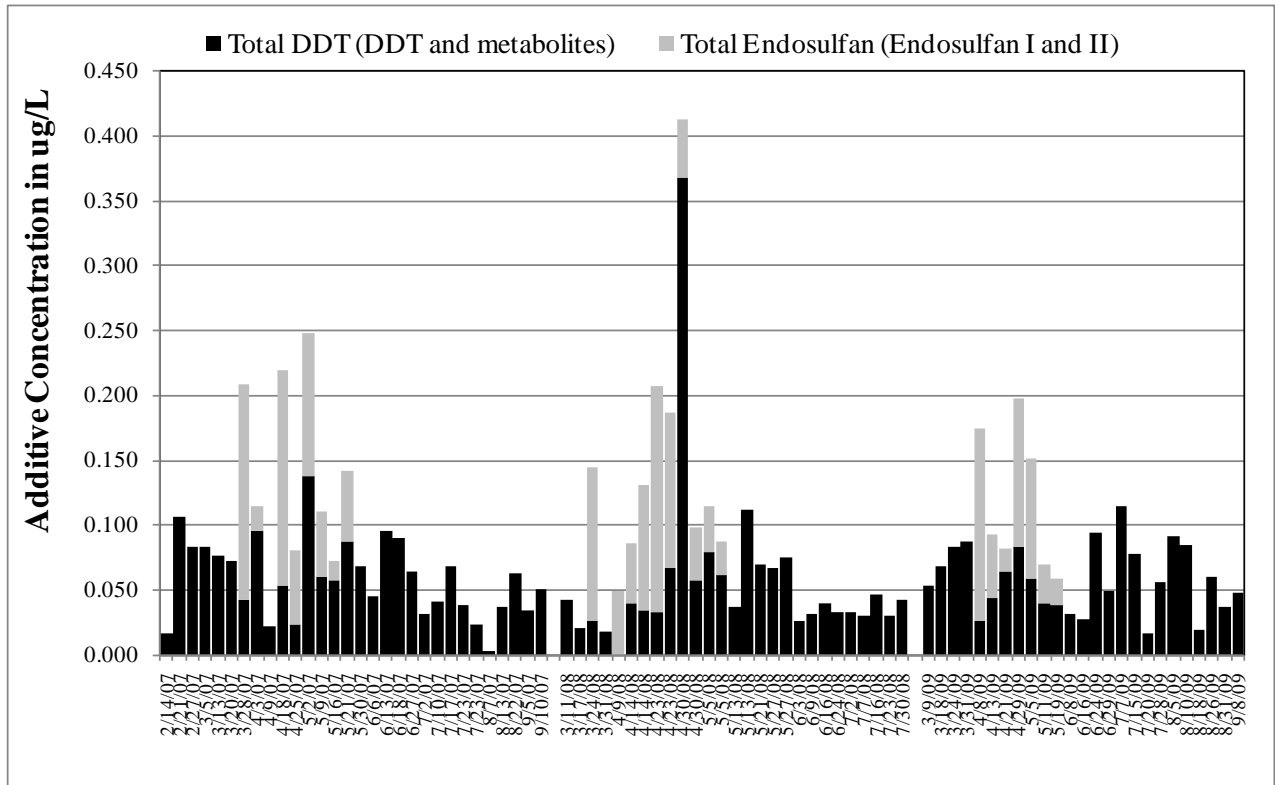


Figure 18. Cumulative amount for organochlorine insecticide detections in Brender Creek, 2007-09.

Wenatchee River

Pesticide Detections and Concentrations

During 2007-09 there were only 13 detections of eight types of pesticides (Table 26).

Comparison to Assessment Criteria and Water Quality Standards

Pesticide calendars in Appendix E, Tables E-22-24, provide a chronological overview of concentrations and detections during 2007-09 and compare pesticide concentrations to *assessment criteria* and *water quality standards* (Appendix D).

During 2007-09 there was one detection of endosulfan that did not meet (was above) the ESLOC criteria for fish once in 2008 and once in 2009. No other detections were above *assessment criteria* or *water quality standards*.

Table 26. Summary of pesticide detections (ug/L) in the Wenatchee River, 2007-09.

Pesticide Name and Type	ALPQL	2007 n =30			2008 n =27			2009 n =27		
		#Det	Freq	Max	#Det	Freq	Max	#Det	Freq	Max
Insecticides										
Chlorpyrifos (Organophosphate)	0.033	1	3.3%	0.035	ND			1	3.7%	0.038
Endosulfan I (Organochlorine)	0.050	1	3.3%	0.014	2	7.4%	0.079	1	3.7%	0.061
Endosulfan II (Organochlorine)	0.050	ND			2	7.4%	0.076	ND		
Imidacloprid (Neonicotinoid)	0.020	ND			1	3.7%	0.028	ND		
Methomyl (Carbamate)	0.046	1	3.3%	0.016	ND			ND		
Oxamyl (Carbamate)	0.047	1	3.3%	0.016	ND			ND		
Wood Preservative										
Pentachlorophenol	0.063	ND			ND			1	3.7%	0.014
Herbicides										
2,4-D	0.063	ND			ND			1	3.7%	0.018

ALPQL: Average lower practical quantitation limit. ND: Not detected.

Pesticide Distribution

As with the majority of the Wenatchee-Entiat sites, insecticides were the most frequently detected type of pesticide in the Wenatchee River. Figure 19 presents the distribution of pesticide types found. Six types of insecticides were detected. Endosulfan I or II were the most frequently observed with six detections during the three-year period.

During 2007-09 no co-occurrence of insecticides with the same mode of action (acetylcholinesterase inhibition) occurred during a sampling event.

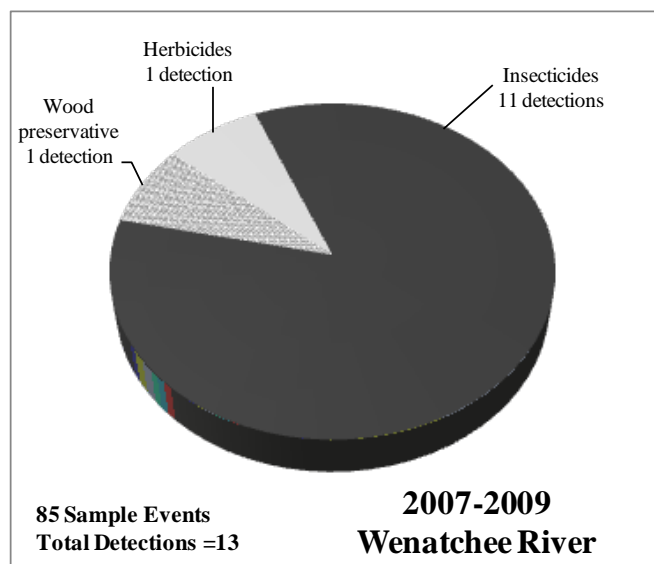


Figure 19. Pesticide distribution in the Wenatchee River, 2007-09.

Entiat River

Pesticide Detections and Concentrations

During 2007-09 the Entiat River had the fewest pesticide detections of any of the sites sampled in the Wenatchee-Entiat basin. Table 27 summarizes pesticide detections. During the three years of sampling, there were nine detections of four pesticides, a degradate, and a synergistic compound.

Table 27. Summary of pesticide detections (ug/L) in the Entiat River for 2007-09.

Pesticide Name and Type	ALPQL	2007 n = 30			2008 n = 27			2009 n = 27		
		#Det	Freq	Max	#Det	Freq	Max	#Det	Freq	Max
Insecticides										
Carbaryl (Carbamate)	0.019	1	3.3%	0.016	ND			ND		
Chlorpyrifos (Organophosphate)	0.033	1	3.3%	0.034	ND			1	3.7%	0.023
Endosulfan I (Organochlorine)	0.050	ND			ND			1	3.7%	0.024
Degradates										
3-Hydroxycarbofuran (Carbamate)	0.047	ND			1	3.7%	0.014	ND		
Synergist										
Piperonyl Butoxide	0.100	MEL added analysis in 2009						3	11.1%	0.100
Herbicides										
Dichlobenil	0.032	1	3.3%	0.065	ND			ND		

ALPQL: Average lower practical quantitation limit. ND: Not detected.

Comparison to Assessment Criteria and Water Quality Standards

Pesticide calendars in Appendix E, Tables E-25-27, provide a chronological overview of concentrations and detections during 2007-09 and compare pesticide concentrations to *assessment criteria* and *water quality standards*. Pesticide concentrations met all *criteria* and *standards*.

Pesticide Distribution

As with the majority of the Wenatchee-Entiat sites, insecticides were the most frequently detected type of pesticide in the Entiat River. Figure 20 presents the distribution of detections by pesticide type. Three types of insecticides were detected. Chlorpyrifos was most frequently found with two detections during the three-year period.

During 2007-09 no co-occurrence of insecticides with the same mode of action (acetylcholinesterase inhibition) occurred during a sampling event.

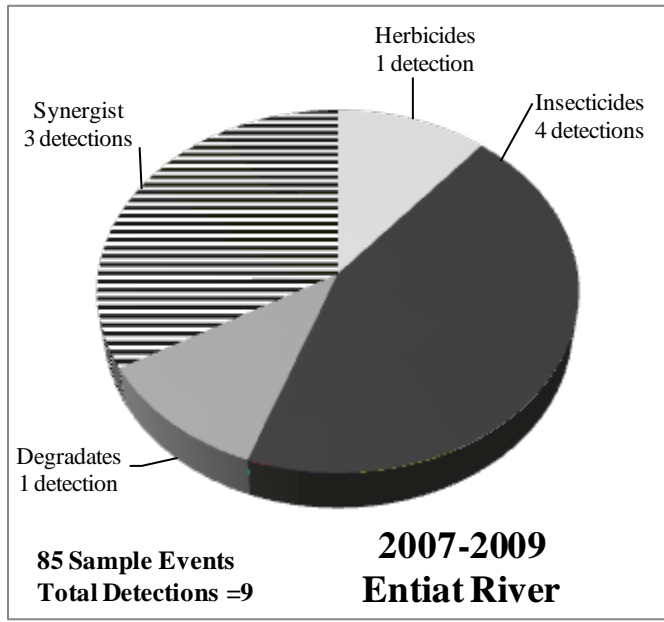


Figure 20. Pesticide distribution in the Entiat River, 2007-09.

Factors Affecting Pesticide Detections

Environmental Factors

Graphs of streamflow, precipitation, and pesticide concentrations of the most commonly detected pesticides are presented in Appendix G.

Of the Wenatchee-Entiat sites, Brender Creek was the only site to have enough pesticide detections for statistical analysis. A statistical test for correlation coefficient (Kendall's tau-b) was used to determine if there was a relationship between some of the more commonly detected pesticides and environmental factors such as flow and rainfall. Correlation analysis was also used to examine the association between commonly detected pesticides and TSS. A two-tailed, Kendall's tau-b, non-parametric correlation coefficient was used to test for correlation between parameters. NJ qualified data were used in this test. Previous rainfall was calculated based on sample time.

The results of the statistical test showed that on Brender Creek, there was:

- A positive correlation between total DDT and TSS (Figure 21) (Kendall's tau = 0.43, $p < 0.01$).
- A weak positive correlation between total DDT and flow (Kendall's tau = 0.38, $p < 0.01$).
- A very weak positive correlation between endosulfan sulfate and flow (Figure 22) (Kendall's tau = 0.38, $p < 0.01$).

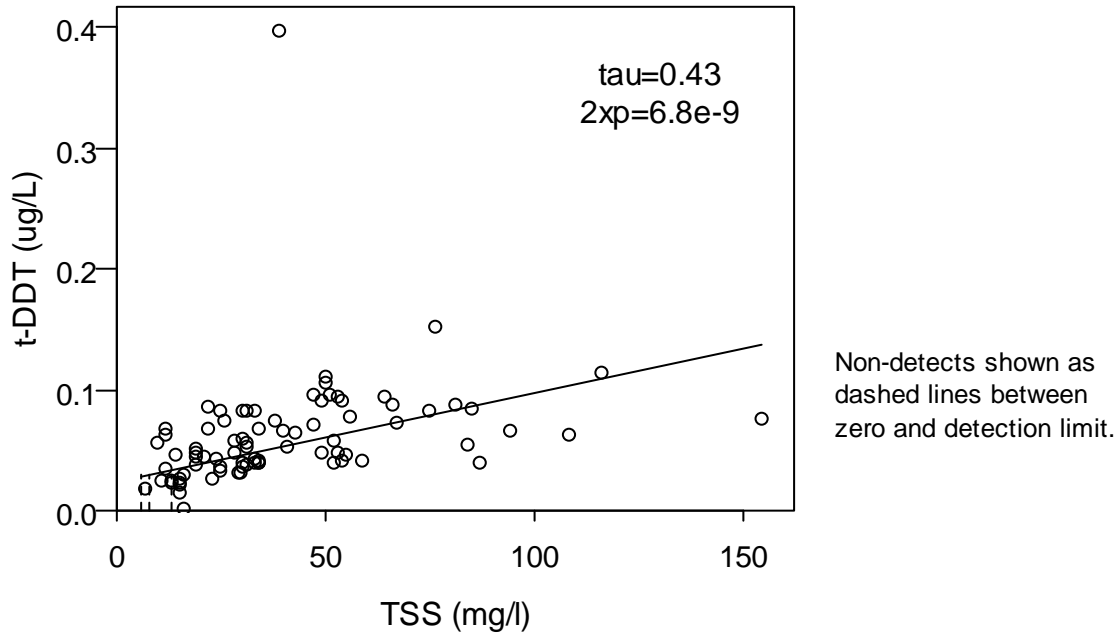


Figure 21. Brender Creek positive correlation between total DDT and total suspended solids, 2007-09.

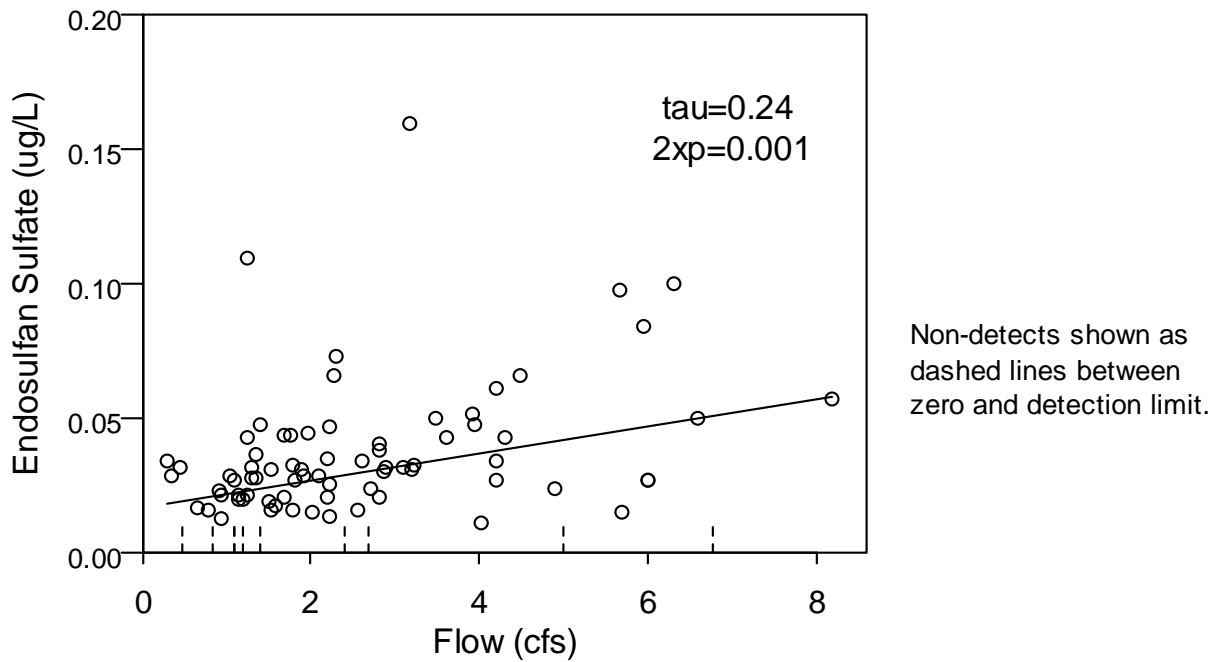


Figure 22. Brender Creek weak positive correlation between endosulfan sulfate and flow, 2007-09.

Temporal Factors

There was a seasonal pattern in pesticide detections. Figure 23 presents the type of pesticide detections by month for all the Wenatchee-Entiat sites combined. In looking at the cumulative total for all sites, the greatest number of insecticide detections was observed in April (Figure 23). For Brender Creek, the greatest number of insecticide detections occurred in April and May.

For the cumulative total, degradate detections peaked in May and then decreased each month thereafter. This pattern was similar for Brender Creek but degradate detections peaked in May and June. In Brender Creek the greatest number of herbicide detections occurred in May and June, while for the rest of the sites, a greater number of detections occurred in August.

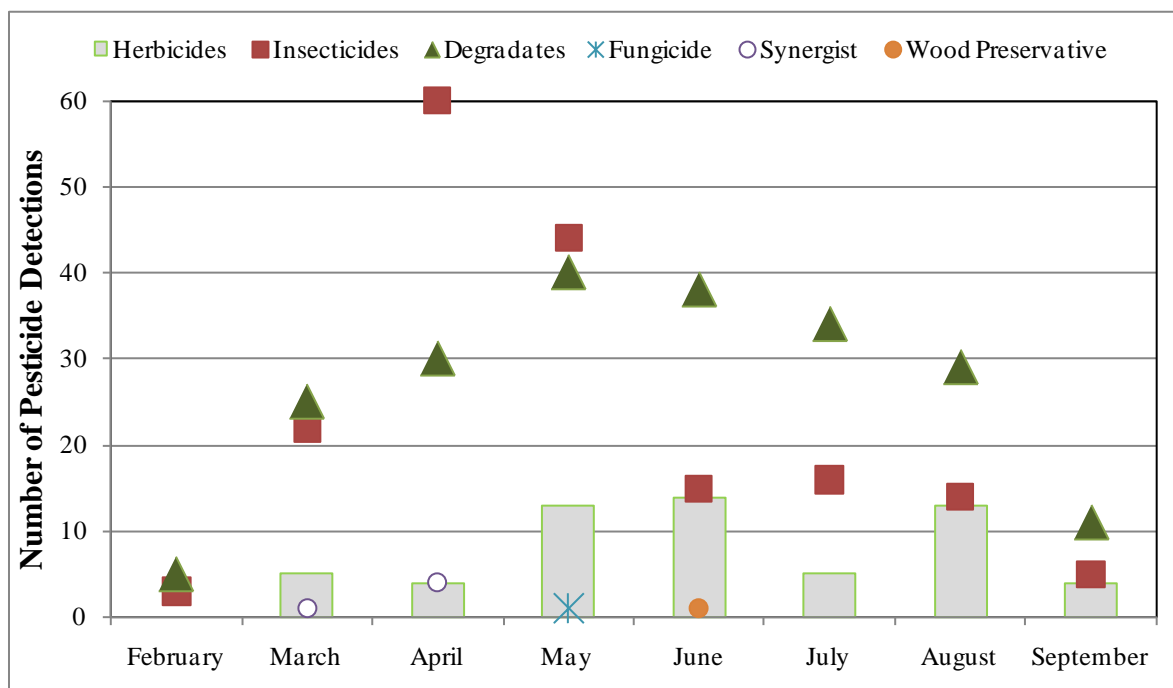


Figure 23. Number of detections by pesticide type for the Wenatchee-Entiat sites, 2007-09.

Conventional Parameters

Conventional water quality parameters were measured at all Wenatchee-Entiat sites. In 2008 and 2009, Winkler dissolved oxygen measurements were also obtained. Continuous, 30-minute interval temperature data were collected; temperature profiles are presented in Appendix F. Table 28 summarizes results for TSS, flow, pH, conductivity, and dissolved oxygen for all sites. All sites must meet freshwater quality standards.

Dissolved oxygen grab samples were collected in 2008 and 2009. All sites met the dissolved oxygen standard of a minimum of 8.0 mg/L per day.

Table 28. Arithmetic mean and range for conventional parameters (grabs) for the Wenatchee-Entiat sites, 2007-09.

	Total Suspended Solids (mg/L)			Flow (cubic feet per second)			pH (standard units)			Conductivity (µmhos/cm)			Dissolved Oxygen (mg/L)	
	2007	2008	2009	2007	2008	2009	2007	2008	2009	2007	2008	2009	2008	2009
Peshastin Creek														
Mean ₁	13	5	9	272	222	214	8.1	7.9	8.1	91	90	108	11.3	11.1
Minimum	<1	<1	<1	12	10	13	7.7	7.6	7.5	45	53	56	8.6	8.2
Maximum	218	44	67	1340	929	606	8.5	8.3	8.6	133	180	158	13.8	13.5
Number	31	27	27	31	22	27	31	27	27	31	27	27	26	27
Brender Creek														
Mean ₁	45	36	37	3.2	2.2	2.2	8.2	8.1	8.2	218	210	236	10.6	10.5
Minimum	14	6	7	1.1	0.7	0.3	7.8	7.9	7.8	123	125	151	9.0	9.2
Maximum	155	94	116	8.2	4.5	6.8	9.4	8.3	8.5	411	333	354	12.5	12.2
Number	31	27	27	31	27	27	31	27	27	31	27	27	27	27
Mission Creek														
Mean ₁	37	8	20	36	19	29	8.3	8.3	8.3	196	186	194	11.2	11.2
Minimum	<1	1	<1	0.2	0.1	0.1	7.3	7.3	7.9	120	107	110	8.9	9.2
Maximum	685	42	85	223	60	101	9.2	8.6	8.7	294	328	324	14.0	13.2
Number	31	27	27	31	27	27	31	27	27	31	27	27	26	27
Wenatchee River														
Mean ₁	10	7	8	4793	4465	3775	8.2	8.2	8.2	46	45	51	11.7	11.6
Minimum	1	<1	<1	467	669	493	7.4	7.2	7.0	23	20	22	9.2	9.3
Maximum	102	46	46	12900	19100	13400	9.1	9.2	9.1	83	76	87	15.1	14.8
Number	31	27	27	31	27	27	31	27	27	31	27	27	26.0	27.0
Entiat River														
Mean ₁	9	5	6	833	681	607	8.2	8.3	8.1	57	69	61	11.1	10.9
Minimum	1	1	1	123	107	96	7.3	7.5	7.0	24	23	23	9.0	9.1
Maximum	64	24	46	2490	2780	2330	9.7	9.2	9.0	103	409	99	13.1	13.8
Number	31	27	27	31	27	27	31	27	27	31	27	27	27	27

Mean¹: Arithmetic mean

Sites which did not meet (exceeded) the pH standard all three years are: Mission Creek, Wenatchee River, and Entiat River. Of the other two sites, Brender Creek exceeded the standard only in 2007 and Peshastin Creek slightly exceeded the standard in 2009.

Temperature

In addition to the discrete temperature measurements, continuous (30-minute interval) measurements were collected year-round. Temperature profiles based on continuous temperature measurements are presented in Appendix F, Figures F-12 to F-26. There are gaps in temperature data due to thermistors being out of the water during low water levels and datalogger battery malfunction. The temperature standard for the freshwater sites is that the 7-DADMax should not exceed 17.5°C. Mission Creek met temperature standards during 2009 but not during 2007-08 (Table 29). The rest of the sites did not meet temperature standards during 2007-09. Table 29 describes periods when temperature standards were not met.

Table 29. Periods of water temperature exceedance for the Wenatchee and Entiat sites, 2007-09.

Site	2007	2008	2009
Peshastin Creek >17.5°C	July 3-Sept 14	July 11-Aug 26, Sept 4-11	July 3-Sept 4, Sept 11-16
Mission Creek >17.5°C	July 7-17, July 24-Aug 18, Aug 31-Sept 4	July 18-25, Aug 2-19	no exceedances
Brender Creek >17.5°C	July 11-14, July 25-26	Aug 14-18	July 24-Aug 4
Wenatchee River >17.5°C	July 11-Sept 17	July 16-Aug 30, Sept 1, Sept 4-18	July 9-Sept 20
Wenatchee River >13.0°C	no exceedances	Oct 1-5	Oct 2
Entiat River >17.5°C	July 21-Sept 14	July 15-Aug 29, Sept 5-10	July 11-Sept 4, Sept 10-26

Total Suspended Solids

Boxplots of TSS concentrations and loading for the Wenatchee-Entiat sites are presented in Figures 24 and 25 respectively (both figures are in log scale). While Brender Creek had the highest average TSS concentrations, Mission Creek had the greatest range in values and the highest maximum TSS concentrations. The Wenatchee then the Entiat had the highest TSS loading averages for 2007-09. Both of these rivers have much higher flows than the other sites.

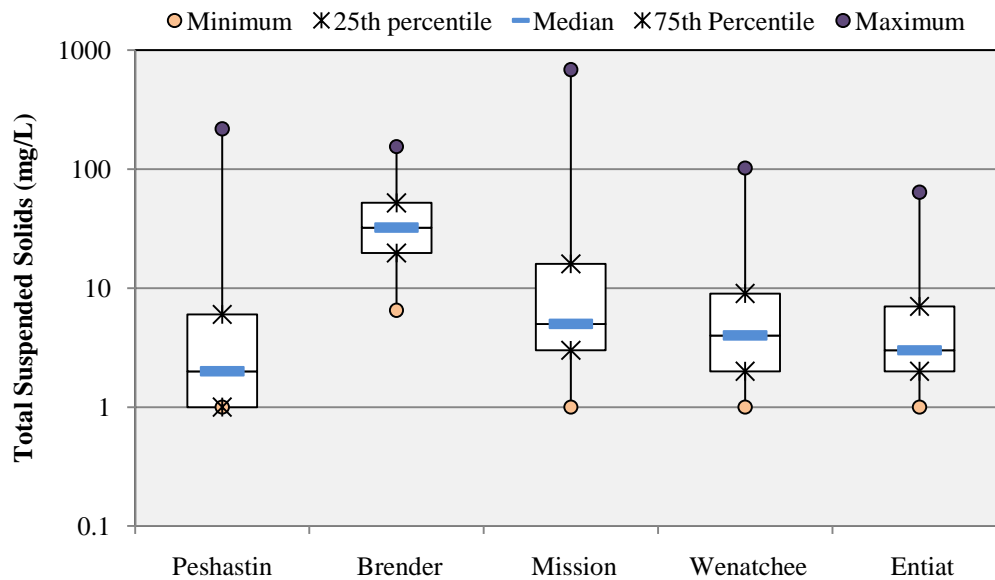


Figure 24. Summary statistics for total suspended solids concentrations for the Wenatchee-Entiat sites, 2007-09.

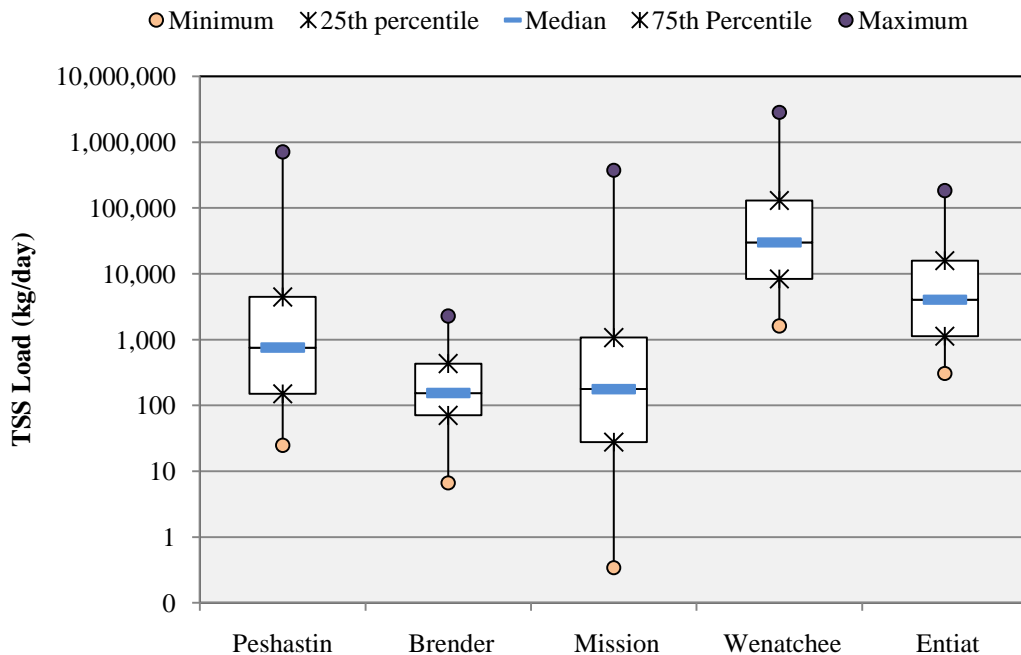


Figure 25. Summary statistics for total suspended solids loading for the Wenatchee-Entiat sites, 2007-09.

Statistical trends in TSS concentrations and loading were examined for all sites using a Seasonal-Kendall trend test (p value ≤ 0.05 , two-tailed). No trends were found, but this is likely due to the small data set.

Comparison to the Lower Mission Creek Basin TMDL

In 2004, a Total Maximum Daily Load (TMDL) was established for DDT in the lower Mission Creek basin (Serdar and Era-Miller, 2004). Target total-DDT loads were recommended for Mission, Brender, and Yaksum Creeks based on waters meeting 1 ng/L total-DDT, the chronic *water quality standard* for protection of aquatic life. Recommendations also included reductions in TSS to < 1 mg/L in order to meet target DDT loads. Phase one of the TMDL compliance schedule included interim monitoring of TSS and DDT at select locations in Yaksum and Brender Creeks. The TMDL recommended the reporting limit for DDT and its degradates, DDD and DDE, in water samples be no higher than 0.5 ng/L.

The reporting limits for this 2007-09 study are insufficient to adequately evaluate DDT levels recommended in the 2004 TMDL. Due to the cost of analyzing for a broad suite of pesticides, the reporting limit for the 2007-09 study is higher than the recommended 0.5 ng/L. The 2007-09 average reporting limit for DDT and its degradates was 0.033 ug/L (33 ng/L). Although the laboratory will report positively identified detections below this limit, the detections are qualified as estimates.

During 2007-09 there were no DDT or degradate detections reported at the Mission Creek site; however, the detection limit is higher than the recommended 0.5 ng/L. At the Brender Creek site, there were DDT or degradate compound detections for all but one sampling event in 2008 and for all but two sampling events in 2009. During 2007-09 the Brender Creek site did not meet the TMDL TSS recommended concentration of < 1 mg/L during any sampling event, ranging from a minimum of 7 to a maximum of 155 mg/L.

Discussion for the Wenatchee-Entiat Basins

Pesticide Detections

Monitoring sites in the Wenatchee-Entiat basins represent surface water quality in a tree fruit agricultural area. A large portion of acreage in the uplands is in forest land, and much of the lowland area is in agricultural production. Major crops include pears, apples, and cherries. Figure 26 presents the types of pesticides detected at the Wenatchee-Entiat sites for three years of monitoring, 2007-09.

Unlike results for the other monitoring sites, the majority of pesticide detections for the Wenatchee-Entiat sites are insecticides and insecticide degradates. The higher proportion of insecticide and degradate detections observed at the Wenatchee-Entiat sites is in part driven by DDT and DDT degradate detections in Brender Creek.

During 2007-09, 214 pesticide detections in Brender Creek were DDT or DDT degradates. DDT is a legacy pesticide that is no longer registered for use in the United States. Detections of DDT and DDT degradate compounds are a result of historic use and do not reflect current pesticide-use patterns.

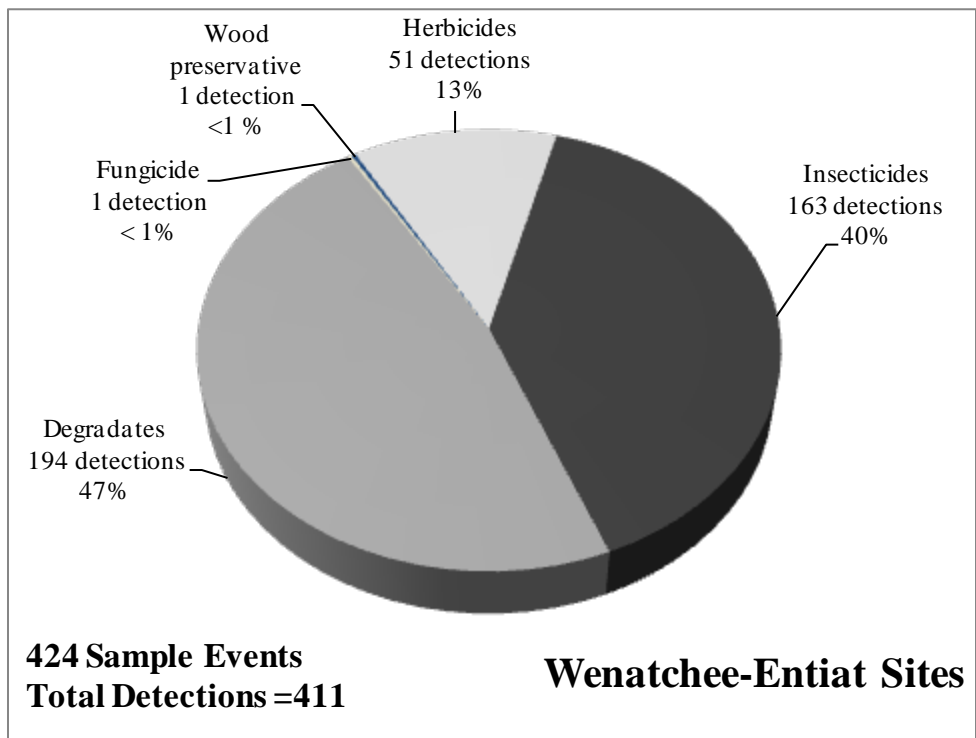


Figure 26. Distribution of pesticides detected in the Wenatchee-Entiat project area, 2007-09.

Brender Creek had the most pesticide detections and the greatest number of pesticide concentrations that exceeded an *assessment criteria* or *water quality standard*. Of the 424 pesticide detections during 2007-09, 401 were in Brender Creek. During all but three sampling events, Brender Creek did not meet (exceeded) the chronic *water quality standard* for total DDT.

In Brender Creek during each of the three years, there were four or more total endosulfan concentrations that exceeded the ESLOC for fish. The greater number of pesticide detections and concentrations found above *assessment criteria* and *water quality standards* is due in part to the low volume of water in Brender Creek as compared to the other sites.

Table 30 presents a comparison of average streamflow by year for each of the Wenatchee-Entiat sites. Flow in Mission Creek is an order of magnitude greater than in Brender Creek. Peshastin Creek and the Entiat River are two orders of magnitude greater, and the Wenatchee River flow is three orders of magnitude greater, than Brender Creek flows. Higher flows dilute pollutant concentrations.

Table 30. Average flow (cfs) during the sampling season the Wenatchee-Entiat sites, 2007-09.

Site	2007	2008	2009
Peshastin Creek	91	139	214
Mission Creek	36	19	29
Brender Creek	3.2	2.2	2.2
Wenatchee River	4790	4470	3780
Entiat River	833	681	607

Endosulfan

With the exception of the Entiat River, all sites at one time did not meet (exceeded) the total endosulfan ESLOC for fish. When pesticide concentrations are detected at the sites with greater flows, much higher pesticide loading occurs.

Figure 27 presents total endosulfan loading in grams per day for each of the Wenatchee sites. There were no endosulfan detections in the Entiat River. The Brender and Mission Creek loading scale (right side of the graph) is much less than the loading scale for the Wenatchee River and Peshastin Creek (left side of the graph).

Likely there are smaller tributaries upstream, or upstream inputs, along the Wenatchee River that are contributing endosulfan to the downstream Wenatchee River site. For example, on March 31, 2008 Wenatchee River endosulfan loading was over 400 grams per day. This would mean there would be over 400 Brender Creek loading inputs upstream to account for that amount of loading in the Wenatchee River.

In July 2010, EPA signed an agreement with the registrants of endosulfan that will result in voluntary cancellation and phase out of all existing uses in the United States. Under the agreement, all endosulfan uses will be phased out by July 2016. Endosulfan use on stonefruit

such as nectarines, peaches, cherries, and apricots will be prohibited by July 2012, on pears by July 2013, and on apples by July 2015. EPA is terminating uses of endosulfan to address its unacceptable risks to agricultural workers and wildlife (EPA, 2010).

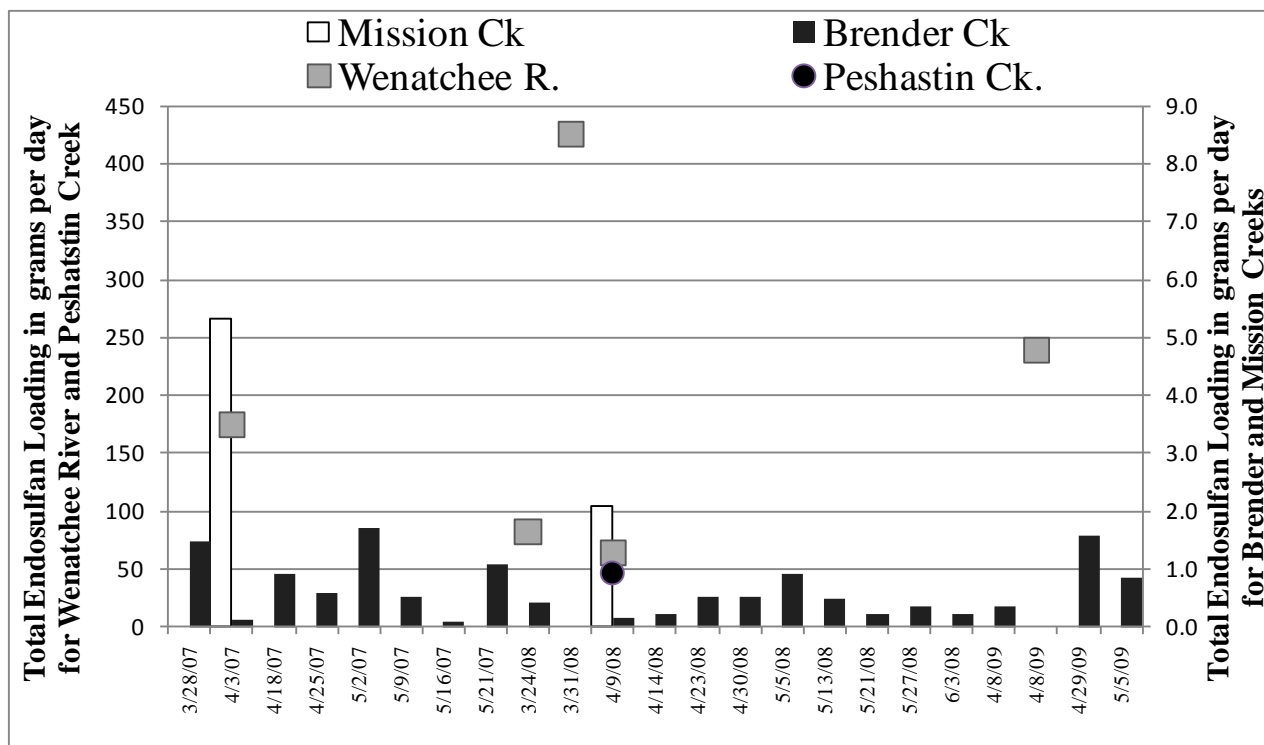


Figure 27. Total endosulfan loading (in grams per day) for the Wenatchee sites, 2007-09.

Comparing Wenatchee-Entiat Sites to Other Pesticide Monitoring Sites

Table 31 presents the ratio of pesticide detections to sampling events for each area. The Wenatchee-Entiat sites have the lowest ratio of detections per sampling event of any of the areas. This may be in part due to the higher flows at some of the Wenatchee-Entiat sites.

Table 31. Ratio of pesticide detections to the number of sampling events for each site for each project area, 2003-09.

Time Period	Thornton Creek		Samish-Skagit	Lower Yakima		Wenatchee-Entiat
	2003-05	2006-08	2006-08	2003-05	2006-08	2007-09
Number of Sampling Events	109	124	406	279	328	424
Number of Detections	317	189	1216	1078	1115	411
Mean Number of Detections per Sampling Event	2.9	1.5	3.0	3.9	3.4	1.0

Water Quality

Conventional Parameters

None of the Wenatchee-Entiat sites consistently met *water quality standards* for temperature during 2007-09. Currently 29% of Ecology's 303(d) listings of impaired waters (category 5) are for temperature. In part this is because temperature is the easiest and least costly parameter to monitor; Ecology receives more temperature data than for any other parameter.

Dissolved oxygen grab sampling was conducted in 2008 and 2009. All Wenatchee-Entiat sites met the dissolved oxygen standard of a minimum of 8.0 mg/L per day.

Sites which did not meet (exceeded) the pH standard all three years were Mission Creek, Wenatchee River, and Entiat River. Of the other two sites, Brender Creek exceeded the standard only in 2007, and Peshastin Creek slightly exceeded the standard in 2009.

Turbidity and TSS are common measures to determine the effect of suspended sediment on salmonids. There are water quality standards for turbidity but not for TSS. TSS is a direct measure of suspended sediment while turbidity is only an indicator. Thus TSS more accurately reflects possible effects on salmonids (Bolton et al., 2001). High sediment levels can have a range of effects from fatal to sub-lethal effects such as reduction of foraging capability, reduced growth, increased stress, and interference with cues necessary for orientation in homing and migration (Bolton et al., 2001). The effects of TSS on fish and aquatic invertebrates are a function of sediment concentration and duration of exposure.

Table 32 presents the range of median and maximum TSS concentrations for the three years of sampling for all sites. In 2007 maximum TSS concentration for the Wenatchee basin sites were high but this is likely due to a March 13, 2007 sampling event where TSS values were in the hundreds (mg/L) range at all sites. Higher TSS concentration on this day could have been due to the first flush of water from the irrigation system. High TSS could also be due to snow melt and runoff. The daily maximum air temperatures in Cashmere increased by 10° F on March 11, 2007. Wenatchee River daily flow went from 2600 cfs on March 11, to 11600 cfs on March 13, 2007.

Table 32. Median and maximum total suspended solids concentrations (mg/L), 2007-09.

Site	Median	Maximum
Peshastin Creek	2-4	44-218
Mission Creek	4-11	42-685
Brender Creek	27-39	94-155
Wenatchee River	4-5	46-102
Entiat River	3-6	24-64

In a review of studies on the effects of suspended solids on fish and aquatic life, Bilotta and Brazier (2008) and Newcombe and MacDonald (1991) reported sub-lethal effects such as:

- Gill hyperplasia and poor condition of fry in chinook salmon exposed to 1.5-2.0 mg/L TSS for 1440 hours (60 days).
- Reduction in growth rate of chinook salmon exposed to 6 mg/L TSS for 1440 hours (60 days).
- Reduction in growth rate of chinook salmon and steelhead exposed to 84 mg/L TSS for 336 hours (14 days).
- 100% mortality of incubating rainbow trout eggs exposed to 47 mg/L TSS for 1152 hours (48 days).
- Reduction in growth rate of rainbow trout exposed to 50 mg/L for 1848 hours (77 days).
- 40% reduction in stream invertebrate diversity after exposure to 130 mg/L TSS for 8760 hours (365 days).
- 77% reduction in population size for benthic invertebrates exposed to 62 mg/L TSS for 2400 hours (100 days).

TSS median and maximum concentrations observed at most of the Wenatchee-Entiat sites likely do not meet the time duration required for sub-lethal effects of salmon or aquatic macroinvertebrates. Brender Creek had the highest TSS concentrations, with median concentrations from 27-39 mg/L over the three-year period. If these TSS concentrations meet the time requirements above, sub-lethal effects to Chinook salmon (first two bullets above) could occur.

Recommendations in the *Lower Mission Creek Basin, Chelan County: TMDL* (Serdar and Era-Miller, 2004) include reductions in TSS to < 1 mg/L in order to meet target DDT loads. Reductions in TSS would help reduce DDT concentrations and reduce possible suspended sediment effects on fish.

Pesticides

During 2007-09 total DDT concentrations regularly exceeded the chronic *water quality standard* in Brender Creek. Brender Creek also had three exceedances for chlorpyrifos and one exceedance for azinphos-methyl. Peshastin Creek had one exceedance for azinphos-methyl as well. No detections of azinphos-methyl occurred after 2007 at any of the Wenatchee-Entiat sites. Azinphos-methyl is being phased out with uses (including on pears, cherries, and apples) prohibited after September 30, 2012.

All of the Wenatchee sites had at least one exceedance of the ESLOC for total endosulfan. Brender Creek total endosulfan concentrations exceeded the ESLOC for fish six, eight, and four times in 2007, 2008, and 2009 respectively. All exceedances occurred during late March through May.

No exceedances of any pesticide *assessment criteria* or *water quality standard* occurred for the Entiat River during 2007-09.

Sub-Lethal Effects and Co-Occurrence of Pesticides

The EPA *assessment criteria* and Washington State *water quality standards* used in this study are based on evaluating the effects of a specific chemical on an organism. The criteria and standards do not take into account (1) the additive or possible synergistic effects of pesticide mixtures or (2) the effects of pesticides when fish are stressed due to environmental factors such as high temperatures or low dissolved oxygen levels.

Recent work by Laetz et al. (2009) found additive and synergistic toxicity to juvenile coho salmon for the binary combinations of several organophosphate and carbamate insecticides. Organophosphate and carbamate insecticides inhibit the activity of acetylcholinesterase (AChE). Environmental mixtures of these insecticides have the potential to exert toxic effects on exposed organisms at concentrations lower than expected from the effects predicted from single chemical toxicity studies.

With the exception of Brender Creek, co-occurrence of the same mode-of-action insecticides did not occur at the Wenatchee-Entiat sites. During the 2007-09 Brender Creek sampling, co-occurrence of AChE-inhibiting insecticides occurred four times. The highest cumulative concentration found was 0.047 ug/L on May 21, 2007 for carbaryl (carbamate) and chlorpyrifos (organophosphate).

The Laetz et al. (2009) study found that insecticide combinations produce additive toxicity at low, environmentally-relevant concentrations (0.1 EC₅₀ for juvenile coho). For example, in the study, diazinon and chlorpyrifos were synergistic when combined at 7.3 and 0.1 ug/L, respectively.

The monitoring results from this 2007-09 study illustrate the difficulty of assessing the effects of multiple chemicals on aquatic organisms. The pesticide calendars in Appendix E demonstrate that mixtures of pesticides are common. However concentrations are typically below the effects threshold for single chemical toxicity testing, and when mixtures occur the various pesticides detected have different modes of action (e.g., not all pesticides inhibit AChE). Even when mixtures of AChE-inhibiting compounds occur, there is limited toxicity data available to assess the potential effects of the mixture. Further confounding the assessment are the effects of environmental factors such as temperature, dissolved oxygen, or pH that can further stress aquatic organisms.

Salmonid Presence during the Pesticide-Use Season (March - September)

In the Wenatchee basin, the greatest concern for salmonids is endosulfan detections not meeting (above) the ESLOC for fish:

- For Peshastin Creek, endosulfan was detected twice above the ESLOC for fish in early April, once in 2007 and again in 2008. During this period, steelhead are spawning and incubating, and spring Chinook and bull trout are incubating and rearing. Temperatures exceed criteria in late July and August with the highest 7-DADMax temperatures for 2007-09 ranging from 22.8 – 25.4°C.

- For Mission Creek, endosulfan was detected once above the ESLOC for fish in early April 2008. In April steelhead spawn and incubate, and spring and summer Chinook are rearing (Table 6). While Mission Creek has lower flows than some of the other sites, very few pesticides are detected in the creek and only the one endosulfan detection was above an *assessment criteria* or *water quality standard*. Temperatures exceed criteria in late July and August with the highest 7-DADMax temperatures for 2007-09 ranging from 18.7 – 21.7°C.
- For Brender Creek, endosulfan levels regularly exceed the ESLOC for fish during late March through mid-May. During this period, there are chronic aquatic health concerns in Brender Creek due to the frequency of endosulfan detections. As a result of these concerns, WSDA initiated outreach activities with growers in 2008. In 2009 the total number of exceedances of the *water quality standard* and the ESLOC for fish; and the number of detections for endosulfan decreased slightly in Brender Creek. In addition, total DDT is present consistently at concentrations above the chronic *water quality standard*. In April 2009, there were two consecutive weeks of chlorpyrifos detections that indicate a potential chronic aquatic health concern during this period. Temperatures rarely exceed criteria in Brender Creek. The highest 7-DADMax temperatures for 2007-09 range from 17.9 – 18.2°C.
- For the Wenatchee River, endosulfan was detected twice above the ESLOC for fish once in late March 2008, and once in early April 2009. During this period lower Wenatchee River spring and summer Chinook and bull trout are rearing; and spawning and incubating steelhead are present. Temperatures exceed criteria in late July and August with the highest 7-DADMax temperatures for 2007-09 ranging from 22.1 – 24.9°C.

In the Entiat basin, pesticides are rarely detected and pesticide levels met all pesticide *assessment criteria* and *water quality standards* for protection of aquatic life. Temperature is the biggest concern for fish during July and August when temperatures exceed 17.5° C. Highest 7-DADMax temperatures for 2007-09 ranged from 20.7 – 23.9°C.

Factors Affecting Pesticide Detections

There is a positive relationship between the legacy pesticide total DDT and TSS concentrations in Brender Creek.

During 2007-09 the greatest number of insecticide detections occurred in April, and the greatest number of pesticide degradates detections occurred in May. USGS (Embrey and Frans, 2003) found the greatest influence on pesticide concentrations and detections appeared to be the season and timing of pesticide application of specific crops or plants. The results of this 2007-09 study show similar findings.

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Conclusions for the Wenatchee-Entiat Basins

Results from the 2007-09 study of pesticide use in the Wenatchee and Entiat River basins support the following conclusions.

- The major factor in pesticide detections for Wenatchee-Entiat sites is likely the season of the year and the timing of pesticide application for specific crops.
- Brender Creek was the only site where many pesticides were detected. Only a few pesticides were found at all the other sites. This is in part due to higher streamflows at some of the other sites (e.g., Peshastin Creek and the Wenatchee and Entiat Rivers).
- Unlike in the other four basins in this study, the majority of pesticides detected in the Wenatchee-Entiat basins were insecticides and insecticide degradates. Very few herbicides were found.
- Co-occurrence of acetylcholinesterase-inhibiting insecticides rarely occurred and cumulative concentrations were low. Co-occurrence of the organochlorine compounds DDT and its degradates and endosulfan occurred on Brender Creek.
- Consistent detections of total DDT indicate chronic health concerns for aquatic life (e.g., fish) in Brender Creek. There is a positive relationship between DDT and total suspended solids (TSS); therefore, reductions in TSS would likely lead to lower DDT concentrations.
- Endosulfan levels in Brender Creek from mid-March through May indicate chronic aquatic health concerns. These levels are above the endangered species level of concern (ESLOC) for fish.
- While infrequent, the periodic detections of endosulfan in Peshastin Creek and the Wenatchee River are of concern due to the substantial endosulfan loading these detections represent (Figure 27). While these endosulfan concentrations may meet (not exceed) an EPA *assessment criteria* or Washington State *water quality standard*, the loading levels represent significant mass. Endosulfan loading sources are unknown but are likely a combination of contribution from upstream tributaries and aerial pesticide spray drift.
- There are very few pesticide detections at the Entiat River site, and all detections meet EPA *assessment criteria* and Washington State *water quality standards*.

Recommendations for the Wenatchee-Entiat Basins

Results from the 2007-09 study of pesticide use in the Wenatchee and Entiat River basins support the following recommendations.

- EPA will begin phasing out endosulfan use in 2013, with all uses prohibited by 2016. The Washington State Department of Agriculture should continue to work with agricultural stakeholders to explore mitigation measures for endosulfan concentrations found in surface water in the Wenatchee basin. Monitoring should continue to assess the effectiveness of mitigation measures.
- Total suspended solids in Brender Creek should be reduced in order to reduce DDT concentrations in the water column as recommended in the 2004 Total Maximum Daily Load (TMDL) study.
- Pesticide monitoring at the Entiat River site should be discontinued. Few pesticide detections are found at this site, and all detections meet *assessment criteria* and *water quality standards*. This station should be replaced with a lower streamflow surface water site in the Wenatchee or Entiat basin.

Summary of Project Changes for 2009

The following changes were made during the 2009 monitoring of the six basins across Washington State.

- Longfellow Creek in the Green Duwamish basin was added to represent urban land use.
- Monitoring at the upstream Thornton Creek site was discontinued.
- Sampling for the following pesticides or degradates was added for all basins:
 - 4,4'-Dichlorobenzophenone (degradate of dichlorobenzophenone)
 - Acetochlor (herbicide)
 - beta-Cypermethrin (pyrethroid insecticide)
 - Bifenthrin (pyrethroid insecticide)
 - Butachlor (herbicide)
 - Chlorpyrifos O.A. (degradate of chlorpyrifos)
 - Diazoxon (degradate of diazinon)
 - Disulfoton Sulfoxide (degradate of disulfoton)
 - Fenamiphos Sulfone (degradate of fenamiphos)
 - Fipronil (pyrethroid insecticide)
 - Fipronil Disulfanyl (degradate of fipronil)
 - Fipronil Sulfide (degradate of fipronil)
 - Fipronil Sulfone (degradate of fipronil)
 - lambda-Cyhalothrin (pyrethroid insecticide)
 - Phorate O.A. (degradates of phorate)
 - Piperonyl Butoxide (synergist)
 - Prothiofos (organophosphate insecticide)
 - Tricyclazole (fungicide)
- All 2009 results for the following carbamates were rejected due to concerns above false positives:
 - 1-naphthol
 - Aldicarb sulfone
 - Aldicarb sulfoxide
 - Oxamyl

Project Change Planned for 2010

The following change is planned for the 2010 sampling during the multi-year study.

The carbamate analysis method used for this project has been: EPA Method 8321 AM modified using electrospray ionization (ES) and atmospheric pressure chemical ionization (APCI) and selected ion monitoring mass spectrometry (SIM-MS). During 2010 the following will be used instead: EPA Method 8321 AM modified using electrospray ionization with jet stream technology and triple quadrupole mass spectrometry.

This change will increase accuracy of carbamate analyte detections by providing confirmation of detected analytes. In addition, the new instrumentation may allow for lower detection limits for carbamates.

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Appendices

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Appendix A. Glossary, Acronyms, and Abbreviations

Glossary

Additive effect: Occurs when the combined effect of two chemicals is equal to the sum of the effects of each chemical.

Assessment criteria: Assessment criteria in this report are numeric criteria included in the EPA FIFRA Pesticide Registration Toxicity Criteria and endpoints; and the EPA National Recommended Water Quality Criteria (NRWQC).

Basin: Watershed. A drainage area in which all land and water areas drain or flow toward a central collector such as a stream, river, or lake at a lower elevation.

Bioaccumulation: Progressive increase in the amount of a substance in an organism or part of an organism which occurs because the rate of intake exceeds the organism's ability to remove the substance from the body.

Boxplot: A graphical depiction of a data set showing the 25th percentile, 50th percentile or median, the 75th percentile, range of data, and outliers.

Carbamate insecticide: N-methyl carbamate insecticides are similar to organophosphate insecticides in that they are nerve agents that inhibit cholinesterase enzymes. However they differ in action from the organophosphate compounds in that the inhibitory effect on cholinesterase is brief.

Clean Water Act: A federal act passed in 1972 that contains provisions to restore and maintain the quality of the nation's waters. Section 303(d) of the Clean Water Act establishes the TMDL program.

Conductivity: A measure of water's ability to conduct an electrical current. Conductivity is related to the concentration and charge of dissolved ions in water.

Degradate: Pesticide breakdown product.

Dissolved oxygen: A measure of the amount of oxygen dissolved in water.

Endosulfan: Endosulfan is an organochlorine insecticide that is registered for use on a number of agricultural commodities. In 2010, EPA signed an agreement with the registrants of endosulfan that will result in voluntary cancellation and phase out of all existing endosulfan uses in the United States. Under this agreement, all endosulfan uses will be phased out by July 2016. EPA is terminating uses of endosulfan to address its unacceptable risks to agricultural workers and wildlife (EPA, 2010).

Exceeded criteria: Did not meet criteria.

Grab sample: A discrete sample from a single point in the water column or sediment surface.

Herbicide: A substance used to kill plants or inhibit their growth.

K_{oc} (sorption coefficient): The tendency of a pesticide to bind to soil particles. Sorption retards movement and may also increase persistence because the pesticide is protected from

degradation. The higher the K_{oc} , the greater the sorption potential. K_{oc} is derived from laboratory data. Many soil and pesticide factors may influence the actual sorption of a pesticide to soil.

Legacy pesticide: Banned pesticides no longer used but that persist in the environment.

Loading: The input of pollutants into a waterbody.

Organochlorine insecticide: Organochlorine insecticides are neurotoxins that are highly lipophilic, very hydrophobic, and chemically stable. As a result, organochlorine insecticides are persistent in the environment and have a long half-life. The lethal mechanism of action is a persistent opening of the sodium channels in neurons, resulting in repetitive firing of action potentials.

Organochlorine pesticide: Organochlorine pesticides are hydrocarbons that contain chlorine (e.g., DDT, endrin, and endosulfan).

Organophosphate pesticide: Organophosphate pesticides are derived from phosphoric acid and are highly neurotoxic, typically inhibiting cholinesterase.

Parameter: Water quality constituent being measured (analyte). A physical, chemical, or biological property whose values determine environmental characteristics or behavior.

Pesticide: A pesticide is any substance or mixture of substances intended for killing, repelling or mitigating any pest. Pests include nuisance microbes, plants, fungus, and animals.

pH: A measure of the acidity or alkalinity of water. A low pH value (0 to 7) indicates that an acidic condition is present, while a high pH (7 to 14) indicates a basic or alkaline condition. A pH of 7 is considered to be neutral. Since the pH scale is logarithmic, a water sample with a pH of 8 is ten times more basic than one with a pH of 7.

Point source: Sources of pollution that discharge at a specific location from pipes, outfalls, and conveyance channels to a surface water. Examples of point source discharges include municipal wastewater treatment plants, municipal stormwater systems, industrial waste treatment facilities, and construction sites that clear more than 5 acres of land.

Pollution: Such contamination, or other alteration of the physical, chemical, or biological properties, of any waters of the state. This includes change in temperature, taste, color, turbidity, or odor of the waters. It also includes discharge of any liquid, gaseous, solid, radioactive, or other substance into any waters of the state. This definition assumes that these changes will, or are likely to, create a nuisance or render such waters harmful, detrimental, or injurious to (1) public health, safety, or welfare, or (2) domestic, commercial, industrial, agricultural, recreational, or other legitimate beneficial uses, or (3) livestock, wild animals, birds, fish, or other aquatic life.

Reach: A specific portion or segment of a stream.

Salmonid: Any fish that belong to the family *Salmonidae*. Basically, any species of salmon, trout, or char. www.fws.gov/le/ImpExp/FactSheetSalmonids.htm

Surface waters of the state: Lakes, rivers, ponds, streams, inland waters, salt waters, wetlands and all other surface waters and water courses within the jurisdiction of Washington State.

Suspended sediment: Solid fragmented material (soil and organic matter) in the water column.

Synergistic: A synergistic effect occurs when the combined effects of two chemicals are greater than the predicted sum of each chemical's effects.

Total Maximum Daily Load (TMDL): Water cleanup plan. A distribution of a substance in a waterbody designed to protect it from exceeding water quality standards. A TMDL is equal to the sum of all of the following: (1) individual wasteload allocations for point sources, (2) the load allocations for nonpoint sources, (3) the contribution of natural sources, and (4) a Margin of Safety to allow for uncertainty in the wasteload determination. A reserve for future growth is also generally provided.

Total suspended solids (TSS): The suspended particulate matter in a water sample as retained by a filter.

Water quality standards: Washington State water quality standards.

Watershed: Basin. A drainage area in which all land and water areas drain or flow toward a central collector such as a stream, river, or lake at a lower elevation.

303(d) list: Section 303(d) of the federal Clean Water Act requires Washington State periodically to prepare a list of all surface waters in the state for which beneficial uses of the water – such as for drinking, recreation, aquatic habitat, and industrial use – are impaired by pollutants. These are water quality limited estuaries, lakes, and streams that fall short of Washington State surface water quality standards and are not expected to improve within the next two years.

7-DADMax or 7-day average of the daily maximum temperatures: The arithmetic average of seven consecutive measures of daily maximum temperatures. The 7-DADMax for any individual day is calculated by averaging that day's daily maximum temperature with the daily maximum temperatures of the three days prior and the three days after that date.

Acronyms and Abbreviations

7-DADMax	7-day Average of the Daily Maximum Temperatures
AChE	Acetylcholinesterase enzyme
ALPQL	Average practical quantitation limit
DDD	Dichloro-diphenyl-dichloroethane
DDE	Dichloro-diphenyl-dichloroethylene
DDT	Dichloro-diphenyl-trichloroethane
DPS	Distinct Population Segment
EC ₅₀	Effective concentration to cause immobility in 50% of an invertebrate species, or a reduction in growth of 50% of an aquatic plant species.
Ecology	Washington State Department of Ecology
EIM	Environmental Information Management (Ecology)
EPA	United States Environmental Protection Agency
ESLOC	Endangered Species Level of Concern (EPA)
GC	Gas chromatograph
GCMS	Gas chromatograph coupled with mass spectrometer
K _{oc}	Sorption coefficient

LC ₅₀	Lethal concentration to cause mortality in 50% of test species
LCS	Laboratory control sample
MEL	Manchester Environmental Laboratory
MS	Mass spectrometer
MS/MSD	Matrix spike/matrix spike duplicate
n	Number
NAD	North American Datum
NRWQC	National Recommended Water Quality Criteria (EPA)
NIST	National Institute of Standards and Technology
NOAA	National Oceanic and Atmospheric Administration
QA	Quality assurance
QC	Quality control
RM	River mile
RPD	Relative percent difference
RSD	Relative standard deviation
SOP	Standard operation procedures
TMDL	(See Glossary above)
TSS	(See Glossary above)
USFWS	United States Fish and Wildlife Service
USGS	United States Geological Survey
WAC	Washington Administrative Code
WRIA	Water Resource Inventory Area
WSDA	Washington State Department of Agriculture

Units of Measurement

°C	degrees centigrade
cfs	cubic feet per second
cms	cubic meters per second, a unit of flow
ft	feet
g	gram, a unit of mass
kg	kilograms, a unit of mass equal to 1,000 grams
kg/d	kilograms per day
km	kilometer, a unit of length equal to 1,000 meters
m	meter
mg	milligrams
mg/d	milligrams per day
mg/L	milligrams per liter (parts per million)
mL	milliliters
mm	millimeters
ng/g	nanograms per gram (parts per billion)
NTU	nephelometric turbidity units
psu	practical salinity units
s.u.	standard units
ug/L	micrograms per liter (parts per billion)
umhos/cm	micromhos per centimeter

Appendix B. Monitoring Sites and Duration of Sampling

Table B-1. Station locations, duration of monitoring, and site location descriptions for 2009.

Site	Duration	Latitude	Longitude	Location Description
Cedar-Sammamish Watershed				
TC-3	March - Sept	47.6958	122.2757	Downstream of pedestrian footbridge near Mathews Beach Park.
Green-Duwamish Watershed				
LC-1	March - Sept	47.5625	122.367	Upstream of the culvert under the 12th Fairway on the West Seattle Golf Course.
Skagit-Samish Watershed				
BD-1	March - Sept	48.3086	122.3473	Upstream side of bridge at Milltown Road.
BD-2	March - Sept	48.3887	122.3329	Upstream side of bridge at Lenor Lane.
BS-1	March - Sept	48.3406	122.4140	Downstream of tidegate on Fir Island Road.
IS-1	March - Sept	48.4506	122.4651	Inside upstream side of tidegate at Bayview-Edison Road.
SR-1	March - Sept	48.5209	122.4113	Upstream side of bridge at Thomas Road.
Lower Yakima Watershed				
MA-2	March - October	46.3306	120.1989	Approximately 15 meters upstream of bridge at Indian Church Road.
SP-2	March - Sept	46.2583	119.7101	Downstream side of culvert on McCready Road
SP-3	March - Sept	46.2344	119.6845	Approximately 3 meters downstream of Chandler Canal overpass.
SU-1	March - Sept	46.2509	120.0202	Downstream side of bridge at Holaday Road.
Wenatchee Watershed				
WE-1	March - Sept	47.4721	120.3710	Upstream side of Sleepy Hollow bridge.
MI-1	March - Sept	47.4893	120.4815	Above Woodring Canyon Road and Mission Creek Road.
PE-1	March - Sept	47.5570	120.5825	Approximately 30 meters downstream of bridge at Saunders Road.
BR-1	March - Sept	47.5211	120.4862	Upstream side of culvert at Evergreen Drive.
Entiat Watershed				
EN-1	March - Sept	47.6633	120.2506	Upstream side of bridge at Keystone Road.

Datum in NAD 83.

Appendices C - G

Surface Water Monitoring Program for Pesticides in Salmonid-Bearing Streams, 2009

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Appendix C. Quality Assurance

Laboratory

Data may be qualified if one or more analytical factors affect confidence in the prescribed data value. Manchester Environmental Laboratory qualifies data according to the National Functional Guidelines for Organic Data Review (EPA, 1999, 2007). Definitions of data qualifiers are presented in Table C-1.

Table C-1. Data qualification.

Qualifier	Definition
(No qualifier)	The analyte was detected at the reported concentration. Data are not qualified.
E	Reported result is an estimate because it exceeds the calibration range.
J	The analyte was positively identified; the associated numerical value is the approximate concentration of the analyte in the sample.
NJ	The analysis indicates the presence of an analyte that has been “tentatively identified,” and the associated numerical value represents its approximate concentration.
NAF	Not analyzed for.
NC	Not calculated.
REJ	The sample results are rejected due to serious deficiencies in the ability to analyze the sample and meet quality control criteria. The presence or absence of the analyte cannot be verified.
U	The analyte was not detected at or above the reported sample quantitation limit.
UJ	The analyte was not detected at or above the reported sample quantitation limit. However, the reported quantitation limit is approximate and may or may not represent the actual limit of quantitation necessary to accurately measure the analyte in the sample.

MEL, 2000, 2008; EPA, 1999, 2007.

Performance measures for quality assurance (QA) and quality control (QC) are presented in Table C-2. Lowest concentrations of interest for surface water grab samples are below reporting limits. Detections quantified below reporting limits are qualified as estimates.

Table C-2. Performance measures for quality assurance and quality control.

Analysis Method ¹	Analysis ²	Field/Lab Replicates, MS/MSD ³ , and Lab Control Samples	MS/MSD ³ , Surrogates, and Lab Control Samples
		RPD ⁴	% Recovery
GCMS	Pesticide-Cl	±40	30-130
	Pesticide-N	±40	30-130
	Pesticide-OP	±40	30-130
	Pesticide-Py	±40	30-130
GCMS-H	Herbicides	±50	40-130
LCMS	Pesticide-C	±40	50-150
TSS	TSS	±20	80-120
TOC	TOC	±20	80-120
DOC	DOC	±20	80-120

¹ GCMS = Gas chromatography/mass spectroscopy, EPA method (modified) SW 846 3535M/8270M.

GCMS-H = Derivatizable acid herbicides by GCMS, EPA method (modified) SW 846 3535M/8270M.

LCMS = Liquid chromatography/mass spectroscopy, EPA method (modified) SW 846 3535M/8321AM.

TSS = Total suspended solids, EPA method 2540D.

TOC, DOC = Total and Dissolved Organic Carbon, EPA method 415.1.

² C l = chlorinated, N=nitrogen containing, OP=organophosphorus, Py=pyrethroid, C=carbamate.

³ MS/MSD = Matrix spike and matrix spike duplicate.

⁴ RPD = Relative percent difference.

Lower Practical Quantitation Limits

Lower practical quantitation limits (LPQLs) are the limits at which laboratories may report data without classifying the concentration as an estimate below the lowest calibration standard. The LPQL is determined by averaging the lower reporting values, per analyte, for all batches over each study period. LPQL data are presented in Table C-3.

Table C-3. Mean performance Lower Practical Quantitation Limits (ug/L).

Chemical	¹ Use	Parent	² Analysis Method	LPQL ³		
				2007	2008	2009
1-Naphthol	D-C		LCMS	0.051	0.053	0.050
2,3,4,5-Tetrachlorophenol	D-WP		GCMS-H	0.062	0.063	0.063
2,3,4,6-Tetrachlorophenol	D-WP		GCMS-H	0.062	0.063	0.063
2,4,5-T	H		GCMS-H	0.062	0.063	0.063
2,4,5-TP	H		GCMS-H	0.062	0.063	0.063
2,4,5-Trichlorophenol	F		GCMS-H	0.062	0.063	0.063
2,4,6-Trichlorophenol	F		GCMS-H	0.062	0.063	0.063
2,4-D	H		GCMS-H	0.062	0.063	0.063
2,4-DB	H		GCMS-H	0.062	0.063	0.063
2,4'-DDD	D-OC	DDT	GCMS	0.033	0.033	0.033
2,4'-DDE	D-OC	DDT	GCMS	0.033	0.033	0.033
2,4'-DDT	D-OC	DDT	GCMS	0.033	0.033	0.033
3,5-Dichlorobenzoic Acid	H		GCMS-H	0.062	0.063	0.063
3-Hydroxycarbofuran	D-C	Carbofuran	LCMS	0.040	0.050	0.050
4,4'-DDD	D-OC	DDT	GCMS	0.033	0.033	0.034
4,4'-DDE	D-OC	DDT	GCMS	0.033	0.033	0.034
4,4'-DDT	I-OC		GCMS	0.033	0.033	0.034
4,4'-Dichlorobenzophenone	D		GCMS			0.101
4-Nitrophenol	D-H		GCMS-H	0.062	0.063	0.063
Acetochlor	H		GCMS			0.101
Acifluorfen	H		GCMS-H	0.062	0.063	0.063
Alachlor	H		GCMS	0.033	0.033	0.033
Aldicarb	I-C		LCMS	0.074	0.100	0.100
Aldicarb Sulfone	D-C	Aldicarb	LCMS	0.060	0.050	0.053
Aldicarb Sulfoxide	D-C	Aldicarb	LCMS	0.017	0.020	0.054
Aldrin	I-OC		GCMS	0.033	0.033	0.033
Alpha-BHC	I-OC		GCMS	0.033	0.033	0.033
Atrazine	H		GCMS	0.033	0.033	0.034
Azinphos Ethyl	I-OP		GCMS	0.033	0.033	0.033
Azinphos Methyl	I-OP		GCMS	0.033	0.033	0.050
Benefin	H		GCMS	0.033	0.033	0.033
Bensulide	H		GCMS	0.033	0.033	
Bentazon	H		GCMS-H	0.062	0.063	0.063
Benthiocarb	H-C		GCMS	0.099	0.100	0.101
Beta-BHC	I-OC		GCMS	0.033	0.033	0.033
beta-Cypermethrin 65731-84-2 ([(1R)-1a(S*), 3a] isomer)	I-Py		GCMS			0.101
Bifenthrin	I-Py		GCMS			0.101
Bromacil	H		GCMS	0.034	0.033	0.033
Bromoxynil	H		GCMS-H	0.062	0.063	0.063
Butachlor	H		GCMS			0.304
Butylate	H		GCMS	0.033	0.033	0.033
Captan	F		GCMS	0.033	0.033	0.033
Carbaryl	I-C		LCMS	0.017	0.020	0.020

Chemical	¹ Use	Parent	² Analysis Method	LPQL ³		
				2007	2008	2009
Carbofuran	I-C		LCMS	0.017	0.020	0.020
Carboxin	F		GCMS	0.033	0.034	0.044
Chlorothalonil	F		GCMS	0.033	0.033	0.033
Chlorpropham	H		GCMS	0.033	0.033	0.033
Chlorpyrifos	I-OP		GCMS	0.033	0.033	0.034
Chlorpyrifos O.A.	D-OP		GCMS			0.101
Cis-Chlordane	I-OC		GCMS	0.033	0.033	0.033
Cis-Nonachlor	I-OC		GCMS	0.033	0.033	0.051
Cis-Permethrin	I-Py		GCMS	0.050	0.050	0.051
Clopyralid	H		GCMS-H	0.062	0.063	0.063
Coumaphos	I-OP		GCMS	0.033	0.033	0.051
Cyanazine	H		GCMS	0.033	0.033	0.033
Cycloate	H		GCMS	0.033	0.033	0.033
DCPA	H		GCMS-H	0.062	0.063	0.063
DDVP	I-OP		GCMS	0.059	0.050	0.051
Delta-BHC	I-OC		GCMS	0.033	0.033	0.033
Deltamethrin	I-Py		GCMS	0.099	0.100	0.101
Diallate	H		GCMS	0.033	0.033	0.033
Diazinon	I-OP		GCMS	0.033	0.033	0.033
Diazoxon	D-OP	Diazinon	GCMS			0.101
Dicamba I	H		GCMS-H	0.062	0.063	0.063
Dichlobenil	H		GCMS	0.033	0.033	0.033
Dichlorprop	H		GCMS-H	0.062	0.063	0.063
Diclofop-Methyl	H		GCMS-H	0.062	0.063	0.063
Dieldrin	I-OC		GCMS	0.050	0.050	0.051
Dimethoate	I-OP		GCMS	0.033	0.033	0.033
Dinoseb	H		GCMS-H	0.062	0.063	0.063
Dioxocarb	I-C		LCMS	0.050		
Diphenamid	H		GCMS	0.033	0.033	0.033
Disulfoton	I-OP		GCMS	0.033	0.052	0.112
Disulfoton sulfone	I-OP		GCMS	0.099	0.100	0.101
Disulfoton sulfoxide	D-OP		GCMS			0.135
Diuron	H		GCMS	0.060	0.050	0.058
Endosulfan I	I-OC		GCMS	0.050	0.050	0.051
Endosulfan II	I-OC		GCMS	0.050	0.050	0.051
Endosulfan Sulfate	D-OC	Endosulfan	GCMS	0.033	0.033	0.034
Endrin	I-OC		GCMS	0.050	0.050	0.051
Endrin Aldehyde	D-OC	Endrin	GCMS	0.050	0.050	0.051
Endrin Ketone	D-OC	Endrin	GCMS	0.033	0.033	0.033
EPN	I-OP		GCMS	0.033	0.033	0.033
Eptam	H		GCMS	0.033	0.033	0.033
Ethalfuralin	H		GCMS	0.033	0.033	0.033
Ethion	I-OP		GCMS	0.033	0.033	0.033
Ethoprop	I-OP		GCMS	0.033	0.033	0.033
Fenamiphos	I-OP		GCMS	0.033	0.033	0.038
Fenamiphos Sulfone	D-OP		GCMS			0.101

Chemical	¹ Use	Parent	² Analysis Method	LPQL ³		
				2007	2008	2009
Fenarimol	F		GCMS	0.033	0.033	0.033
Fensulfothion	I-OP		GCMS		0.033	0.033
Fenthion	I-OP		GCMS		0.048	0.033
Fenvalerate (2 isomers)	I-Py		GCMS	0.033	0.033	0.033
Fipronil	I-Pyra		GCMS			0.101
Fipronil Disulfinyl	D-Pyra		GCMS			0.101
Fipronil Sulfide	D-Pyra		GCMS			0.101
Fipronil Sulfone	D-Pyra		GCMS			0.101
Fluridone	H		GCMS	0.099	0.100	0.101
Fonofos	I-OP		GCMS	0.033	0.033	0.033
Heptachlor	I-OC		GCMS	0.033	0.033	0.033
Heptachlor Epoxide	D-OC	Heptachlor	GCMS	0.033	0.033	0.033
Hexachlorobenzene	F		GCMS	0.033	0.034	0.033
Hexazinone	H		GCMS	0.050	0.050	0.051
Imidacloprid	I-N		LCMS		0.020	0.020
Imidan	I-OP		GCMS	0.033	0.033	0.068
Ioxynil	H		GCMS-H	0.062	0.063	0.063
Kelthane	I-OC		GCMS	0.295	0.314	0.304
lambda-Cyhalothrin	I-Py		GCMS			0.101
Lindane	I-OC		GCMS	0.033	0.033	0.033
Linuron	H		GCMS	0.059	0.050	0.051
Malathion	I-OP		GCMS	0.033	0.033	0.033
MCPA	H		GCMS-H	0.062	0.063	0.063
MCPP	H		GCMS-H	0.062	0.063	0.063
Metalaxyl	F		GCMS	0.033	0.033	0.033
Methidathion	I-OP		GCMS	0.295	0.293	0.304
Methiocarb	I-C		LCMS	0.017	0.020	0.021
Methomyl	I-C		LCMS	0.037	0.050	0.050
Methomyl oxime	D-C	Thiodicarb	LCMS	0.017	0.020	0.020
Methoxychlor	I-OC		GCMS	0.033	0.033	0.051
Methyl Chlorpyrifos	I-OP		GCMS	0.033	0.033	0.033
Methyl Paraoxon	D-OP	Methyl parathion	GCMS	0.099	0.100	0.101
Methyl Parathion	I-OP		GCMS	0.033	0.033	0.033
Metolachlor	H		GCMS	0.033	0.033	0.033
Metribuzin	H		GCMS	0.033	0.033	0.033
Mevinphos	I-OP		GCMS	0.050	0.050	0.051
MGK-264	Sy		GCMS	0.033	0.033	0.051
Mirex	I-OC		GCMS	0.033	0.033	0.035
Monocrotophos	I-OP		GCMS	0.050	0.050	0.051
Naled	I-OP		GCMS	0.042	0.059	0.035
Napropamide	H		GCMS	0.050	0.050	0.051
Norflurazon	H		GCMS	0.033	0.033	0.034
Oryzalin	H		GCMS	0.099	0.100	0.114
Oxamyl	I-C		LCMS	0.042	0.050	0.052
Oxamyl oxime	D-C	Oxamyl	LCMS	0.017	0.020	0.020
Oxychlorane	D-OC	Chlordane	GCMS	0.033	0.033	0.033

Chemical	¹ Use	Parent	² Analysis Method	LPQL ³		
				2007	2008	2009
Oxyfluorfen	H		GCMS	0.033	0.033	0.101
Parathion	I-OP		GCMS	0.033	0.033	0.033
Pebulate	H		GCMS	0.033	0.033	0.033
Pendimethalin	H		GCMS	0.033	0.033	0.034
Pentachlorophenol	WP		GCMS-H	0.062	0.063	0.063
Phenothrin	I-Py		GCMS	0.033	0.033	0.033
Phorate	I-OP		GCMS	0.296	0.299	0.291
Phorate O.A.	I-OP		GCMS			0.193
Picloram	H		GCMS-H	0.062	0.063	0.063
Piperonyl Butoxide	Sy		GCMS			0.101
Promecarb	I-C		LCMS	0.031	0.020	0.020
Prometon	H		GCMS	0.033	0.033	0.033
Prometryn	H		GCMS	0.033	0.033	0.033
Pronamide	H		GCMS	0.033	0.033	0.033
Propachlor	H		GCMS	0.033	0.033	0.033
Propargite	I-SE		GCMS	0.033	0.033	0.051
Propazine	H		GCMS	0.033	0.033	0.033
Propoxur	I-C		LCMS	0.040	0.050	0.050
Prothiofos	I-OP		GCMS			0.101
Resmethrin	I-Py		GCMS	0.050	0.050	0.036
Simazine	H		GCMS	0.033	0.033	0.033
Simetryn	H		GCMS	0.099	0.100	0.101
Sulfotepp	I-OP		GCMS	0.033	0.033	0.033
Sulprofos	I-OP		GCMS		0.033	
Tebuthiuron	H		GCMS	0.033	0.033	0.033
Terbacil	H		GCMS	0.033	0.033	0.034
Tetrachlorvinphos	I-OP		GCMS	0.050	0.050	0.051
Thiodicarb	I-C		LCMS		0.020	
Tokuthion	I-OP		GCMS	0.050	0.050	
Total Suspended Solids			TSS			1.059
Tralomethrin	I-Py		GCMS	0.099	0.100	0.101
Trans-Chlordane	I-OP		GCMS	0.033	0.033	0.033
Trans-Nonachlor	I-OC		GCMS	0.033	0.033	0.051
trans-Permethrin	I-Py		GCMS			0.101
Triadimefon	F		GCMS	0.033	0.033	0.033
Triallate	H		GCMS	0.033	0.033	0.033
Trichloronat	I-OP		GCMS	0.050	0.050	0.051
Triclopyr	H		GCMS-H	0.062	0.063	0.063
Tricyclazole	F		GCMS			0.101
Trifluralin	H		GCMS	0.033	0.033	0.034

¹ C = Carbamate, D = Degradate, F = Fungicide, I = Insecticide, H = Herbicide, OC = Organochlorine, OP = Organophosphorus, Py = Pyrethroid, SE = Sulfite Ester, Sy = Synergist, WP = Wood Preservative.

² GCMS = Gas chromatography/mass spectroscopy, EPA method (modified) SW 846 3535M/8270M.

GCMS-H = Derivizable acid herbicides by GCMS, EPA method (modified) SW 846 3535M/8270M.

LCMS = Liquid chromatography/mass spectroscopy, EPA method (modified) SW 846 3535M/8321AM.

³ Blank cells indicate no analysis for the compound in that year.

Quality Assurance Samples

Quality Assurance (QA) samples were collected each year to assure consistency and accuracy of sample analysis.

For this project, QA samples included field replicates, field blanks, and matrix spike and matrix spike duplicates (MS/MSD). QA samples for the laboratory included split sample duplicates, laboratory control samples, surrogate spikes, and method blanks.

Field QA samples as a percentage of total sample budget increased yearly from 2007 to 2009. Each year, more than 10% of field samples had an associated QA sample (Table C-4).

The total count of field QA samples is in Table C-4. The total count of laboratory QA samples is in Table C-5.

Table C-4. Total field QA samples per analysis type, 2007-09.

QA type	Field Replicates				Field Blanks				MS/MSD ²			Field QA % of sample budget
	Analysis ¹	GCMS	GCMS-H	LCMS	TSS	GCMS	GCMS-H	LCMS	TSS	GCMS	GCMS-H	
2007	28	26	24	25	12	12	11	13	25	23	24	11%
2008	33	30	32	32	17	17	16	16	17	16	16	16.3%
2009	40	36	36	37	23	18	19	20	17	16	16	16.5%
Total	101	92	92	94	52	47	46	49	59	55	56	14.6%

¹ GCMS = Gas chromatography/mass spectroscopy, EPA method (modified) SW 846 3535M/8270M.

GCMS-H = Derivatizable acid herbicides by GCMS, EPA method (modified) SW 846 3535M/8270M.

LCMS = Liquid chromatography/mass spectroscopy, EPA method (modified) SW 846 3535M/8321AM.

TSS = Total suspended solids, EPA method 2540D.

²MS/MSD = Matrix spike and matrix spike duplicates.

Table C-5. Total laboratory QA samples per analysis type, 2007-09.

QA	LDP ²	Lab Blanks				Surrogates			Laboratory Control Samples			
		Analysis ¹	TSS	GCMS	GCMS-H	LCMS	TSS	GCMS	GCMS-H	LCMS	GCMS	GCMS-H
2007	76	92	71	74	89	679	659	663	46	67	43	89
2008	76	35	31	28	66	557	529	526	59	47	44	66
2009	115	40	52	32	61	532	498	504	71	93	88	23
Total	193	199	163	130	207	1633	1562	1530	142	146	119	208

¹ GCMS = Gas chromatography/mass spectroscopy, EPA method (modified) SW 846 3535M/8270M.

GCMS-H = Derivatizable acid herbicides by GCMS, EPA method (modified) SW 846 3535M/8270M.

LCMS = Liquid chromatography/mass spectroscopy, EPA method (modified) SW 846 3535M/8321AM.

TSS = Total suspended solids, EPA method 2540D.

²LDP = Laboratory duplicates.

Results for each QA sample method are outlined in the sections below.

Field Replicates

Results for pesticide field replicates are presented in Tables C-6 and C-7. Table C-6 presents the data value, data qualification (if assigned), and relative percent difference (RPD) between the results for compounds which were consistently identified in both the grab sample and replicate.

Consistent identification refers to compounds which were identified in both the original sample and field replicate. Inconsistently identified replicate pairs are those in which the compound was identified in one sample but not the other. Inconsistently identified grab sample replicates are presented in Table C-7.

Field replicates were used with 5.2%, 7.8%, and 8.1% of all field samples in 2007, 2008, and 2009, respectively. 2.5% of the analysis pairs had a detection in at least one replicate.

Excluding total suspended solids (TSS), 39 parameters were detected in 113 replicate pairs. Of these, 81% were consistently identified in both samples. 79% of consistent pairs were within the 40% RPD criterion.

TSS was detected in 32 replicate pairs. All TSS replicate pairs were consistently identified. 75% were within the 20% RPD criterion.

The 2009 ratio of consistent to inconsistent replicate sets is similar to results from this program's 2003-05 surveys (71%; Burke et al., 2006) and 2006-08 surveys (75%; Sargeant et al., 2010), and the USGS-NAWQA replicate analysis (1992-1997 samples) when the median detected pesticide concentration was less than 0.1 ug/L (84%; calculated from Table 2 in Martin, 2002). In both the USGS and our studies, the associated error of inconsistent replicate sets precludes use in variability analysis.

The average RPD of consistent field replicate pairs was very low, 9.6% (Table C-6). Similarly, the median pooled relative standard deviation (RSD) of all replicates was 6.3%. This variation is lower than our 2006-2008 results (8.1%, Sargeant et al., 2010) and the NAWQA median pooled RSD of 15% at concentrations <0.01 ug/L and 12% at concentrations near 0.1 ug/L (Table 8 in Martin, 2002).

Among consistent replicates, 4 parameters had a maximum RPD over criteria (Table C-6):

- 4,4'-DDE
- DCPA
- Endosulfan I
- Total Suspended Solids

Overall RPD for the parameters not meeting (exceeding) RPD criteria ranged from 0% to 67%. RPDs for other analyte pairs ranged from 0% to 40%. The failure of these samples to fall within the acceptable range is most likely due to the high amount of variability in detections near the minimum reporting limit (Martin, 2002; Mathieu, 2006).

Table C-6. Detected pairs within field replicate results, 2009 (µg/L).

Parameter	Sample	Replicate	RPD
2,4-D	0.079	0.078	1
	0.99	0.91	8
	0.15	0.1	40
	0.02 J	0.02 J	0
	0.098	0.096	2
	0.11	0.09	20
	0.051 J	0.053 J	4
	0.079	0.078	1
	0.019 J	0.022 J	15
	0.036 J	0.034 J	6
	Mean =		10
4,4'-DDD	0.019 J	0.019 J	0
	0.015 J	0.013 J	14
	Mean =		7
4,4'-DDE	0.022 J	0.02 J	10
	0.016 J	0.016 J	0
	0.026 J	0.014 J	60
	0.044	0.042	5
	Mean =		19
4,4'-DDT	0.022 J	0.023 J	4
	0.027 J	0.022 J	20
	0.036	0.035	3
	Mean =		9
Bentazon	0.025 J	0.024 J	4
	0.13	0.15	14
	Mean =		9
Bromacil	0.019 J	0.027 J	35
	0.074	0.068	8
	0.046	0.042	9
	0.07	0.069	1
	0.045	0.047	4
	0.14	0.15	7
	0.058	0.059	2
	Mean =		10
Bromoxynil	0.072	0.073	1
Carbaryl	0.021	0.022	5
Carbofuran	0.099	0.105	6

Parameter	Sample	Replicate	RPD
Chlorpyrifos	0.041	0.048	16
	0.03 J	0.028 J	7
	0.053	0.056	6
	0.037	0.039	5
	0.08	0.086	7
	0.023 J	0.021 J	9
	Mean =		8
	DCPA	0.12	0.064 J
0.017 J		0.012 J	34
Mean =		48	
Diazinon	0.027 J	0.026 J	4
Dicamba I	0.032 J	0.028 J	13
	0.13	0.12	8
	0.011 J	0.01 J	10
	0.008 J	0.009 J	12
	0.021 J	0.021 J	0
	0.01 J	0.01 J	0
Mean =		7	
Dichlobenil	0.022 J	0.023 J	4
	0.004 J	0.004 J	0
	0.009 J	0.009 J	0
	0.008 J	0.008 J	0
	0.014 J	0.013 J	7
	0.013 J	0.012 J	8
	0.01 J	0.01 J	0
	0.019 J	0.021 J	10
	0.064	0.069	8
	0.026 J	0.024 J	8
	0.011 J	0.01 J	10
	0.011 J	0.011 J	0
	0.008 J	0.009 J	12
Mean =		5	
Diphenamid	0.031 J	0.032 J	3
Endosulfan I	0.024 J	0.03 J	22
	0.018 J	0.017 J	6
	0.044 J	0.028 J	44
	Mean =		24
Endosulfan II	0.063 J	0.052 J	19

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Table C-6. (continued).

Parameter	Sample	Replicate	RPD
Endosulfan Sulfate	0.041 J	0.046 J	11
	0.043	0.044	2
	0.092 J	0.076 J	19
	0.03 J	0.032 J	6
	Mean =		10
Hexazinone	0.056	0.057	2
Imidacloprid	0.038	0.043	12
	0.092	0.09	2
	Mean =		7
MCPA	0.16	0.15	6
	0.09	0.079	13
	0.091	0.086	6
	Mean =		8
MCPP	0.089	0.077	14
Metalaxyl	0.05	0.051	2
Metolachlor	0.086	0.083	4
	0.061	0.056	9
	0.029 J	0.028 J	4
	Mean =		5
Metribuzin	0.045	0.053	16
Pendimethalin	0.06	0.063	5
Pentachlorophenol	0.053 J	0.051 J	4
Prometon	0.072	0.077	7
Tebuthiuron	0.027 J	0.03 J	11
	0.047	0.041	14
	Mean =		12
Terbacil	0.11	0.12	9
	0.11	0.13	17
	Mean =		13
Triclopyr	0.5	0.46	8
	0.06 J	0.057 J	5
	0.076	0.071	7
	Mean =		7
Trifluralin	0.025 J	0.026 J	4
	0.017 J	0.017 J	0
	Mean =		2

Parameter	Sample	Replicate	RPD
Total Suspended Solids	6	6	0
	19	17	11
	30	29	3
	13	12	8
	9	9	0
	8	8	0
	8	8	0
	3	3	0
	32	29	10
	9	9	0
	5	4	22
	5	5	0
	4	4	0
	8	7	13
	13	13	0
	21 J	23	9
	3	6	67
	16	13	21
	49	49	0
	62	56	10
	1	2	67
	8	8	0
	16	16	0
	3	4	29
	10	10	0
	11	10	10
	5	4	22
3	3	0	
2	1	67	
3	3	0	
2	3	40	
9	10	11	
Mean =		13	

Inconsistent replicate detections are an indicator of sampling uncertainty. Table C-7 compares inconsistent replicate detections to the Lower Practical Quantitation Limit (LPQL) for non-detections in the paired replicate. Most inconsistent detections were found at concentrations near or below the LPQL.

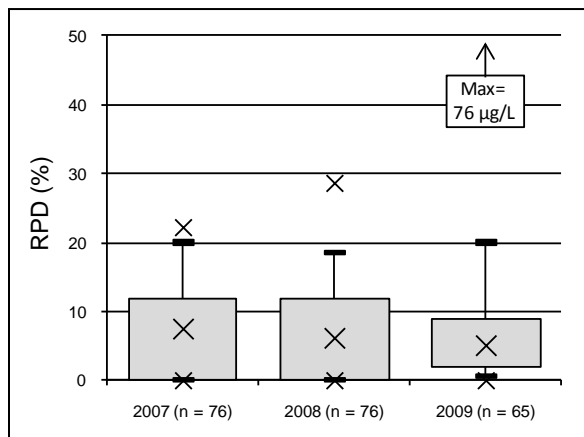
Table C-7. Inconsistent field replicate detections compared to the LPQL, 2009 ($\mu\text{g/L}$).

Parameter	Sample	Replicate	RPD
2,4-D	0.023 J	0.025 NJ	8
	0.024 J	0.028 NJ	15
	Mean =		12
2,4'-DDE	0.009 J	<0.033 U	114
4,4'-DDT	0.020 J	<0.033 U	49
Atrazine	0.016 NJ	0.015 J	6
	0.049	0.055 NJ	12
	<0.032 U	0.022 J	37
	Mean =		18
Bromacil	<0.033 U	0.021 J	44
Dicamba I	0.016 NJ	0.016 J	0
Dichlobenil	0.009 J	0.010 NJ	11
Endosulfan II	0.023 J	<0.051 UJ	76
Imidacloprid	<0.020 U	0.023	14
MCPA	0.022 NJ	0.026 J	17
Methiocarb	<0.020 U	0.033	49
Norflurazon	0.030 J	0.027 NJ	11
Pendimethalin	0.029 NJ	0.028 J	4
Pentachlorophenol	0.018 NJ	0.020 J	11
Picloram	<0.064 U	0.180	95
Simazine	<0.034 U	0.015 J	78
Tebuthiuron	0.034 NJ	0.037	8

¹ Non-detections are listed as less than the Lower Practical Quantitation Limit (<LPQL).

Laboratory Duplicates

Manchester Laboratory used laboratory split sample duplicates to ensure consistency of TSS analyses. Boxplots of RPD for TSS lab duplicates are presented in Figure C-1.



From 2007-09, 95% of all TSS lab duplicate RPDs were less than or equal to the 20% RPD criteria. Some outlier pairs exceeded 20% but did not represent overall recovery.

Figure C-1. TSS laboratory duplicate relative percent difference (%).¹

¹Boxes show 25th and 75th percentiles, whiskers show 5th and 95th percentiles, and 'X' indicates the minimum, median, and maximum values.

Field Blanks

Field blank detections indicate the potential for sample contamination in the field and laboratory and the potential for false detections due to analytical error.

Field blank detections for 2009 are listed in Table C-8.

Table C-8. Grab sample field blank detections for 2009 (µg/L).

Analysis ¹	Chemical	Field Date	Site	Sample	Blank
GCMS	Dichlobenil	3/11/09	LC-1	0.033 U	0.016 J
GCMS	Tricyclazole	4/8/09	BR-1	0.099 U	0.030 J

¹GCMS = Gas chromatography/mass spectroscopy, EPA method (modified) SW 846 3535M/8270M.

On March 11, 2009, dichlobenil was found in a field blank for Longfellow Creek (LC-1). Dichlobenil was not found in the associated sample for LC-1, but it was detected at other western Washington sites on the same day. None of these detections were greater than 5 times the blank concentration, so dichlobenil was qualified as tentatively undetected (UJ) for these samples.

On April 8, 2009, tricyclazole was detected in a field blank for Brender Creek (BR-1) but was not detected in any associated samples (Table C-8).

Laboratory Blanks

Manchester Laboratory uses laboratory blanks to assess the precision of equipment and the potential for internal laboratory contamination. If lab blank detections occur, the sample LPQL may be increased, and detections may be qualified as estimates.

Laboratory blank detections for 2009 are presented in Table C-9.

Table C-9. Laboratory blank detections, 2009 ($\mu\text{g/L}$).

Analysis	Chemical	Analysis Date	Value
LCMS	Aldicarb Sulfone	4/16/09	0.010 J
		4/22/09	0.009 J
	Aldicarb Sulfoxide	4/16/09	0.015 J
	Methiocarb	4/20/09	0.016 J
	Methomyl	4/16/09	0.013 J
	Oxamyl	4/16/09	0.006 J
		4/22/09	0.008 J
	Oxamyl oxime	6/22/09	0.022 J

All lab blank detections were carbamate compounds analyzed by LCMS (Table C-9). Problems with LCMS lab blanks were due to an unidentified low-level interference in the LCMS equipment that resembled the compounds in question (D. Huntamer, 2009, personal communication).

For all lab blank detections, any analytes found in associated samples below 5 times the lab blank detection were reported at the level detected, but qualified as not detected at an estimated detection limit (UJ).

A March 11, 2009 field sample from Sulphur Creek Wasteway had a methiocarb detection of 0.269 $\mu\text{g/L}$ associated with a lab blank detection of 0.016 $\mu\text{g/L}$. The sample concentration was more than 5 times the lab blank concentration and was not qualified. No other sample detections were associated with lab blank detections.

Surrogates

Surrogates are compounds that are spiked into field samples at the laboratory. They are used to check recovery for a group of compounds. For instance, triphenyl phosphate is a surrogate for organophosphorus insecticides (Table C-10).

High pesticide surrogate recovery requires related detections to be qualified as estimates. Low pesticide surrogate recovery requires all related data to be qualified as estimates.

Table C-10. Pesticide surrogates.

Surrogate compound	Surrogate for...
2,4,6-tribromophenol	Acid-derivitizable herbicides
2,4-dichlorophenylacetic acid	
Chloramben	
Carbaryl C13	Carbamate pesticides
4,4'-DDE-13C12	Chlorinated pesticides
4,4-DDE-D8	
Decachlorobiphenyl (DCB)	
Gamma-BHC-D6	
1,3-dimethyl-2-nitrobenzene	Nitrogen pesticides
Chlorpyrifos-D10	Organophosphorus pesticides
Triphenyl phosphate	

Grab sample surrogate recoveries are presented in Figure C-2.

The majority of surrogate recoveries fell within the control limits established by Manchester Laboratory for all compounds except chlorpyrifos-D10 (Figure C-2). Chlorpyrifos-D10 was used as a surrogate for organophosphorus pesticides in GCMS analysis in late 2009. No sample results were qualified because all other GCMS surrogates, including the other organophosphorus surrogate, triphenyl phosphate, were acceptably recovered in all samples.

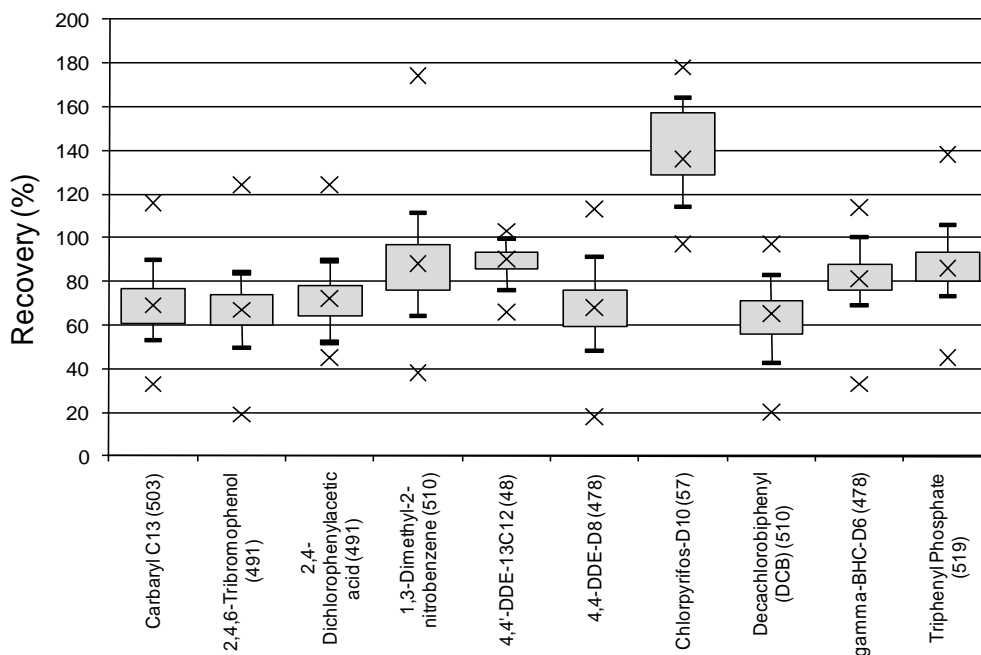


Figure C-2. Grab sample surrogate recoveries (%).¹

¹Boxes show 25th and 75th percentiles, whiskers show 5th and 95th percentiles, and 'X' indicates the minimum, median, and maximum values.

Outlier recoveries were outside of control limits for all surrogates. However, outliers represented a small part of overall surrogate recovery and did not qualify the majority of data.

Matrix Spike/Matrix Spike Duplicates (MS/MSD)

MS/MSD results reflect the process of sample duplication (field), analyte degradation, matrix interaction (sample/standard), extraction efficiency, and analyte recovery. This measure is the best overall indicator of accuracy and reproducibility of the entire sampling process.

Figure C-3 shows percent matrix spike recovery for selected pesticides. Figure C-4 shows the RPD between the matrix spike and the matrix spike duplicate for the same set.

In 2009, the average recovery of matrix-spiked compounds was 88.7%, and the average RPD between MS/MSD pairs was 12.2%. For most compounds, the RPD and recovery of MS/MSD pairs showed acceptable performance and were within defined limits for the project. Due to high variability, Aldicarb sulfoxide and Picloram had an average RPD outside the $\pm 40\%$ criteria. Any unqualified detections of these chemicals were qualified as estimates.

In 2009, diuron recovered very high in some matrix spikes (maximum recovery of 482%). In these cases, Diuron was reanalyzed using derivitization confirmation and passed quality control (J. Westerlund, 2009, personal communication). No Diuron detections were associated with these high matrix spike recoveries.

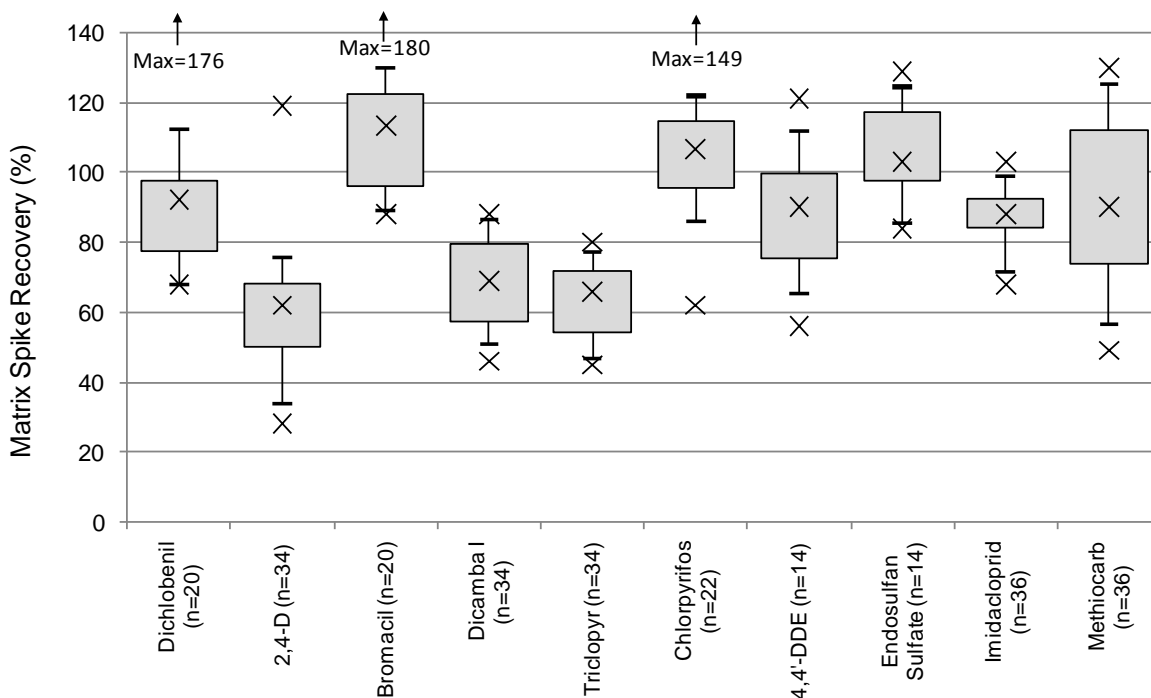


Figure C-3. Matrix spike recovery for selected pesticides. ¹

¹Boxes show 25th and 75th percentiles, whiskers show 5th and 95th percentiles, and 'X' indicates the minimum, median, and maximum values.

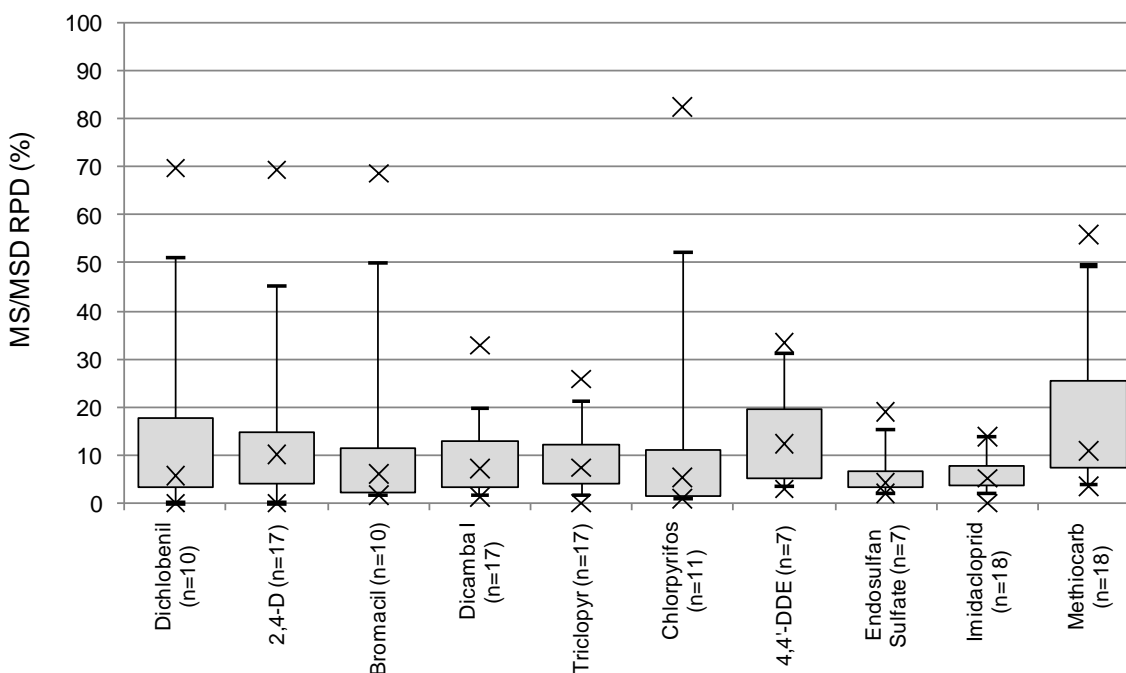


Figure C-4. Paired matrix spike relative percent differences (RPDs) for selected pesticides.¹
¹Boxes show 25th and 75th percentiles, whiskers show 5th and 95th percentiles, and 'X' indicates the minimum, median, and maximum values.

Laboratory Control Samples

Laboratory control samples (LCS) are analyte compounds spiked into deionized water at known concentrations and subjected to analysis. They are used to evaluate accuracy of pesticide residue recovery for a specific analyte. Detections may be qualified based on low LCS recovery and/or high RPD between paired LCS.

Figures C-5 through C-8 show LCS recovery results. LCS tests were conducted with each grab sample analysis. Specific analytes were tested on a rotating basis.

Most grab sample LCS recoveries for pesticide analyses fell within the acceptance criteria established by Manchester Laboratory (Table C-2). Results associated with high or low LCS recoveries were qualified as estimates.

All conventional parameter LCS recoveries fell within the criteria of 80 to 120% recovery (Table C-7).

Figures C-9 through C-11 show paired LCS RPDs. Paired LCS tests were conducted for a subset of LCS to understand recovery consistency. If paired LCS show inconsistent recoveries, additional pairs may be tested. If paired LCS recoveries are still inconsistent, associated sample detections may be qualified as tentative or not detected.

The majority of LCS pairs showed acceptable recovery for all analytes. Sample detections associated with high RPD between LCS pairs were qualified as estimates.

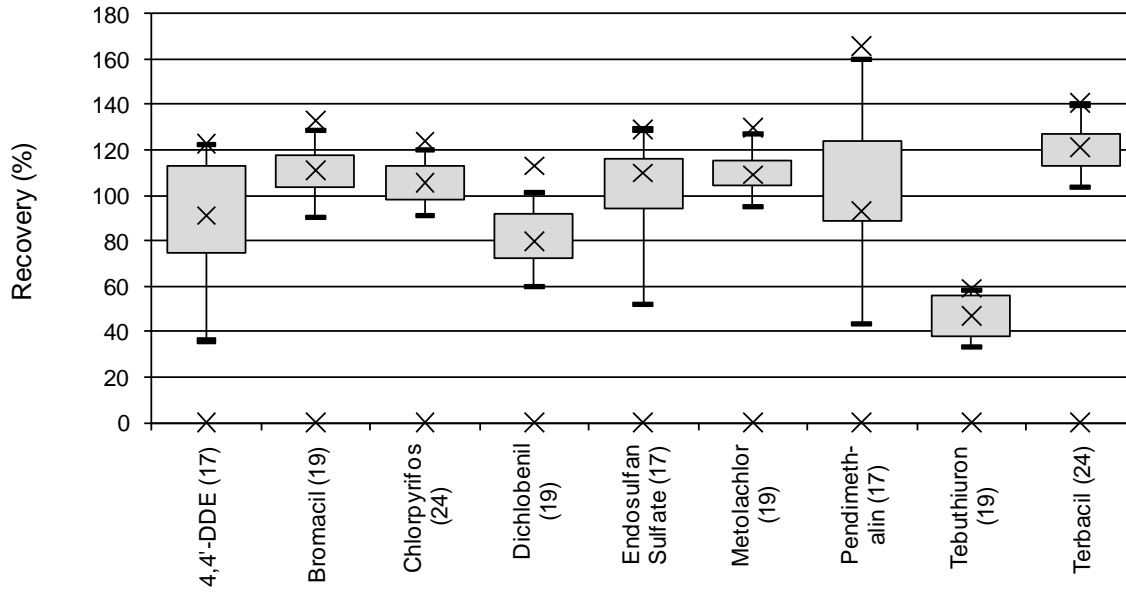


Figure C-5. Laboratory control sample recoveries (%) for selected pesticides by GCMS.^{1,2}

¹Boxes show 25th and 75th percentiles, whiskers show 5th and 95th percentiles, and 'X' indicates the minimum, median, and maximum values.

²GCMS = Gas chromatography/mass spectroscopy, EPA method (modified) SW 846 3535M/8270M.

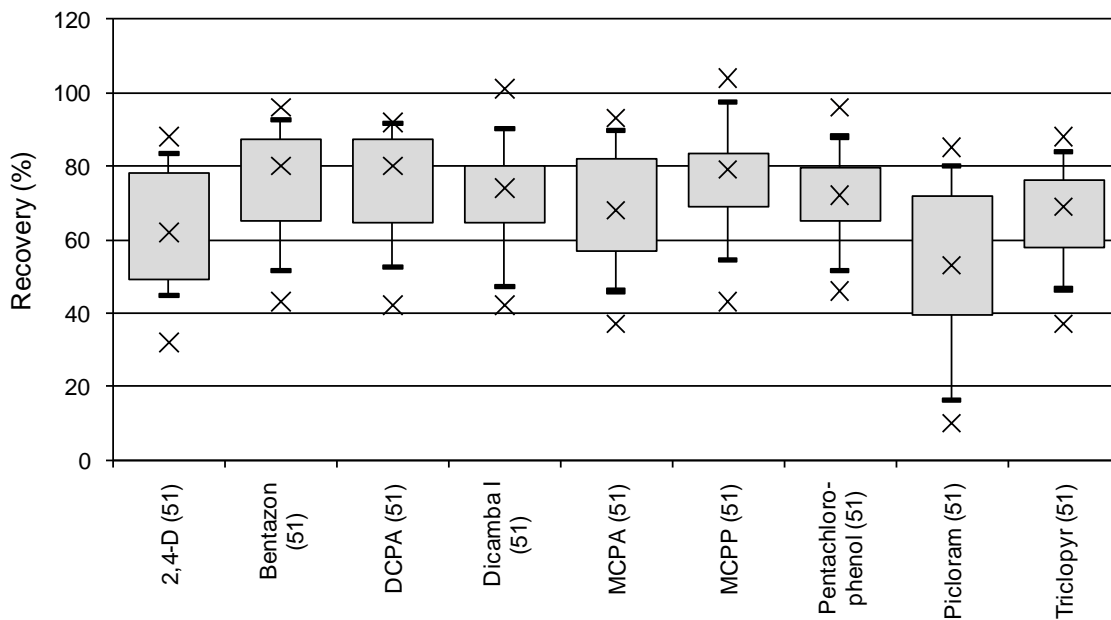


Figure C-6. Laboratory control sample recoveries (%) for selected herbicides by GCMS-H.^{1,2}

¹Boxes show 25th and 75th percentiles, whiskers show 5th and 95th percentiles, and 'X' indicates the minimum, median, and maximum values.

²GCMS-H = Derivatizable acid herbicides by GCMS, EPA method (modified) SW 846 3535M/8270M.

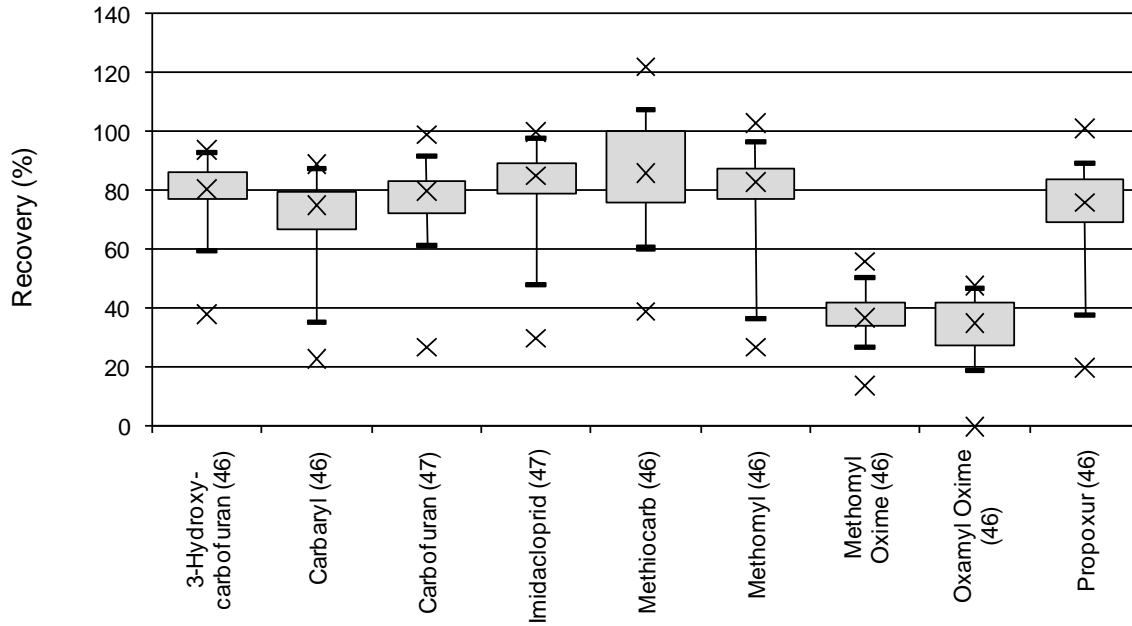


Figure C-7. Laboratory control sample recoveries (%) for selected pesticides by LCMS. ^{1,2}

¹Boxes show 25th and 75th percentiles, whiskers show 5th and 95th percentiles, and 'X' indicates the minimum, median, and maximum values.

²LCMS = Liquid chromatography/mass spectroscopy, EPA method (modified) SW 846 3535M/8321AM.

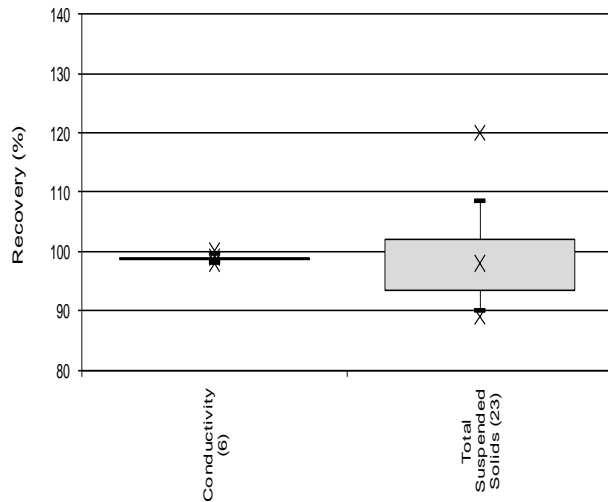


Figure C-8. Laboratory control sample recoveries (%) for conventional parameters. ¹

¹Boxes show 25th and 75th percentiles, whiskers show 5th and 95th percentiles, and 'X' indicates the minimum, median, and maximum values.

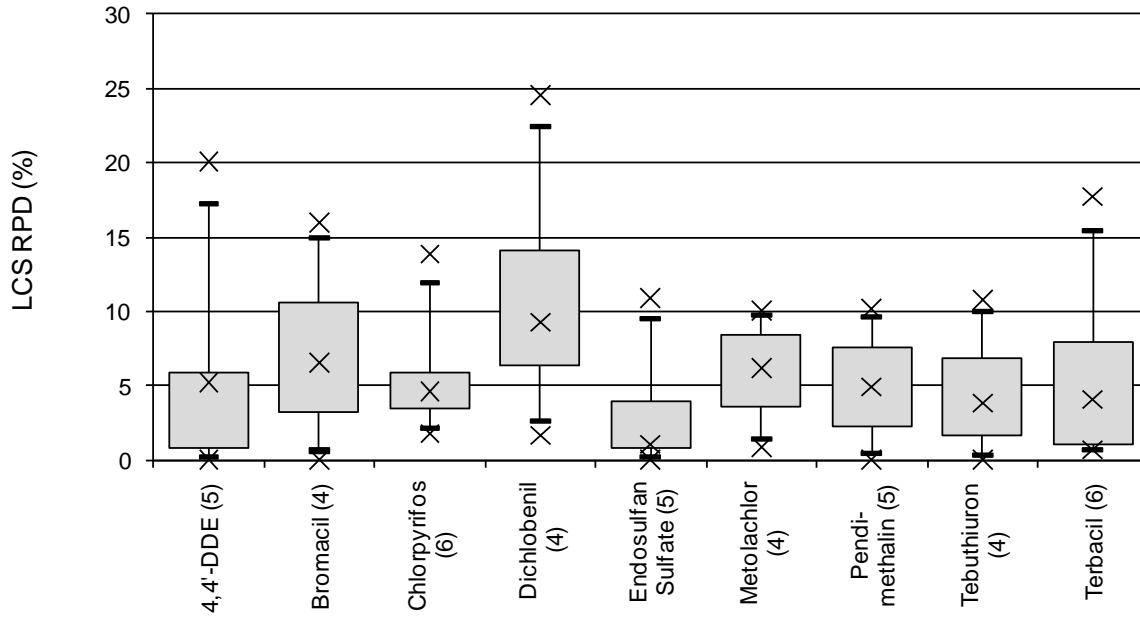


Figure C-9. Paired LCS relative percent differences (%) for pesticides by GCMS. ^{1,2}

¹Boxes show 25th and 75th percentiles, whiskers show 5th and 95th percentiles, and 'X' indicates the minimum, median, and maximum values.

²GCMS = Gas chromatography/mass spectroscopy, EPA method (modified) SW 846 3535M/8270M.

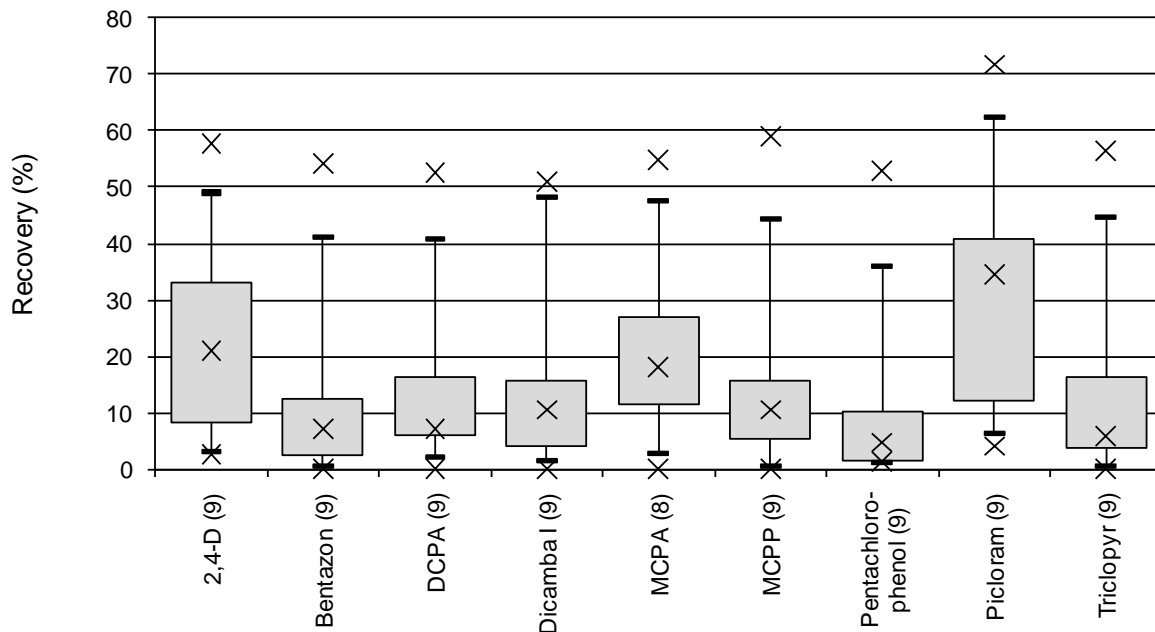


Figure C-10. Paired LCS relative percent differences (%) for pesticides by GCMS-H. ^{1,2}

¹Boxes show 25th and 75th percentiles, whiskers show 5th and 95th percentiles, and 'X' indicates the minimum, median, and maximum values.

²GCMS-H = Derivatizable acid herbicides by GCMS, EPA method (modified) SW 846 3535M/8270M.

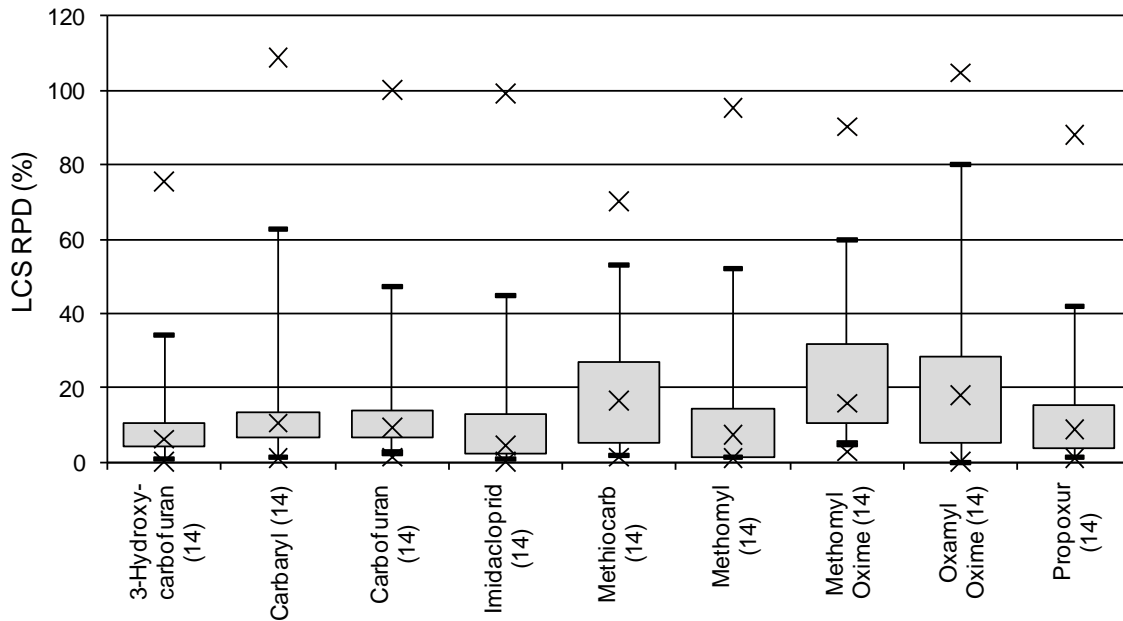


Figure C-11. Paired LCS relative percent differences (%) for pesticides by LCMS. ^{1,2}

¹Boxes show 25th and 75th percentiles, whiskers show 5th and 95th percentiles, and 'X' indicates the minimum, median, and maximum values.

²LCMS = Liquid chromatography/mass spectroscopy, EPA method (modified) SW 846 3535M/8321AM.

Field Data Quality

Quality Control Procedures

Prior to the start of the 2009 sampling season, it was identified that numerical field meter quality objectives for conventional parameters were not included in the original QA Project Plan or subsequent addendums. These quality objectives were never identified because when the project started in 2003 conventional parameters were considered ancillary. As the monitoring program evolved over the last six years, conventional parameters became a larger part of understanding the effects of pesticides on salmonids and understanding the fate and transport of pesticides. To ensure the continued use of quality data in future study reports, field meter quality objectives and other QA/QC procedures were documented in Addendum 3 (Sargeant and Anderson, 2009). These are summarized below.

Field meters were calibrated at the beginning of the field day according to manufacturers' specifications, using Ecology standard operating procedures (Swanson, 2007). Meters were post-checked at the end of the field day using known standards. Conventional parameters measured in the field were replicated once per sample day. Dissolved oxygen meter results were compared to Winkler laboratory titration results from grab samples. Two to three Winkler grab samples were obtained during each sample day.

Results for 2009

In 2009 the field meter for the lower Yakima sites and Wenatchee-Entiat sites (eastside sites) met QC objectives including post-checks and Winkler comparisons (Table C-11) except on March 18, 2009. Dissolved oxygen meter readings were biased high that day; meter and Winkler dissolved oxygen %RSD ranged from 11.1% - 13.6% RSD. Only Winkler dissolved oxygen results will be reported for this day.

Table C-11. Quality control results (%RSD) for field meter and Winkler replicates.

Replicate Meter Parameter	Westside		Eastside	
	Average	Maximum	Average	Maximum
Winkler and meter DO	1.5%	7.7%	2.3%	13.7%
Replicate Winkler's for DO	0.6%	2.2%	0.3%	1.3%
Meter DO	1.3%	7.1%	0.8%	5.8%
Meter conductivity	1.3%	17.5%	1.9%	15.6%
Meter pH	0.7%	3.1%	0.7%	2.8%
Meter flow	4.5%	21.5%	4.8%	23.7%

The field meter for the urban sites and the lower Skagit-Samish sites (westside sites) did not meet post-check QC objectives for conductivity for the following dates: March 16 and 25, April 22 and 27, and May 6, 20, and 26, 2009. Conductivity results for these days are rejected and not reported.

Pooled replicate measurements or Winkler results met data quality objectives; all pooled results were less than 10% RSD (C-11). The %RSD for eastside DO meter results were biased high on March 18, 2009; these results were rejected and Winkler results were used for this day.

On July 17, 2009 an Indian Slough (westside meter) conductivity and flow result exceeded data QC objectives. Due to the tidal influence at this site (and Brown's Slough), conductivity and flow results may vary more due to environmental conditions; results are acceptable.

Three replicate flow results and a conductivity result exceeded data QC objectives for the eastside sites. Flow replicates were during low-flow conditions when the RSD statistic produces higher variability. Flow results for these days are acceptable. The June 24, 2009 Mission Creek conductivity result was qualified as an estimate.

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Appendix D. Assessment Criteria and Water Quality Standards

EPA pesticide assessment documents were reviewed to determine the most comparable and up-to-date toxicity guidelines for freshwater (Table D-1) and marine species (Table D-2).

EPA Toxicity Criteria

Rainbow trout are a surrogate for freshwater endangered and threatened species. *Daphnia magna* (invertebrate) and *Selenastrum capricornutum* (green algae also called *pseudokirchneria subcapitata*) represent components of the aquatic food web that may be affected by pesticide use. Alternative species are used only if no data are available for rainbow trout, *Daphnia magna*, or *Selenastrum capricornutum*.

Marine toxicity criteria were evaluated for detections at sites with estuarine influence. These sites were all in the Skagit-Samish watersheds and included lower Big Ditch, Browns Slough, and Indian Slough. Criteria were generated for marine species including (1) sheepshead minnow and tidewater silverside for fish; (2) pink shrimp, Eastern Oyster, Grass Shrimp, *Acartia tonsa* (copepod), and Mysid shrimp for invertebrates; and (3) *Isochrysis galbana*, and *Skeletonema costatum* for aquatic plants.

EPA classifies a laboratory study as ‘core’ if it meets guidelines appropriate for inclusion in pesticide registration. Usually a core designation may be made if the study is appropriately designed, monitored, and conditions controlled, and duration of exposure is consistent with other studies. Core study criteria are used in the assessment table. Keeping with pesticide review precedent, the most toxic, acceptable criteria from core studies are used.

Water Quality Standards and Assessment Criteria

The most recent versions of Washington State water quality standards and EPA National Recommended Water Quality Criteria (NRWQC) were applied for this report. The NRWQC remained largely unchanged from the 2003 update through 2008.

The toxic standards for Washington State waters also used. These remain essentially unchanged following the 1997 rule and 2003 updates (Washington Administrative Code (WAC), Chapter 173-201A).

Table D-1. Freshwater toxicity and regulatory guideline values. All values reported in ug/L.

Chemical	¹ Freshwater Toxicological and Reregistration Criteria													Freshwater Standards and Criterion			
	Fisheries					Invertebrate				Plant				² WAC		³ NRWQC	
	Acute	Chronic	ESLOC	Spp.	Ref	Acute	Chronic	Spp.	Ref	Acute	Chronic	Spp.	Ref	Acute	Chronic	CMC	CCC
1-Naphthol	1400		70	RT	10	700		DM	10	1100		SC	10				
2,4-D (Acids, Salts, Amines) ^m	101000	14200	5050	RT; FM	1	25000	16050	DM	1	3880	1440	ND	1				
2,4-D (BEE Ester) ^m	428		21.4	BS	1	4970	200	DM	1	1020	538	ND	1				
2,4'-DDD																	
2,4'-DDT																	
3-Hydroxycarbofuran	362	5.7	18.1	RT	54; 60	2.23	0.75	CD	54								
	88		4.4	BG	54	29	9.8/27	DM	60								
4,4'-DDD																	
4,4'-DDE														1.1 ^{a,b}	0.001 ^{a,c}	1.1 ^a	0.001 ^a
4,4'-DDT														1.1 ^{a,b}	0.001 ^{a,c}	1.1 ^a	0.001 ^a
4-Nitrophenol																	
Alachlor	2100	187	105	RT	2	1550	110	DM	2	1.64	0.35	SC	2				
Aldicarb	560	78	28	RT;FM	3	410	20	DM	3								
Aldicarb Sulfone	42000	78	2100	RT;FM	3	280	20	DM	3								
Aldicarb Sulfoxide	7140	78	357	RT-A; FM-C	3	696	20	DM	3								
Atrazine	5300	65	265	RT; BT	4	6900	140	DM	4	49		SC	4				
Azinphos Methyl	2.9	0.23	0.145	RT	5	1.1	0.25	DM	5								0.01
	3.2		0.16	Coho	5												
Bentazon	>100000		>5000	RT	6	>100000		DM	6	4500		SC	6				
Bromacil	36000		1800	RT	7	121000		DM	7	6.8		SC	7				
Bromoxynil	50	18/ 39	2.5	RT-A; FM-C	8	11	2.5/5.9	DM	8	80		SC	8				
Carbaryl	1200		60	RT	9	5.6	1.5	DM	10	1100	370	SC	10				
	2400		120	Chinook	10												
	2400		120	Coho	10												
Carbofuran	362	5.7	18.1	RT	54; 60	2.23	0.75	CD	54								
	88		4.4	BG	54	29	9.8/27	DM	60								
Chlorothalonil	42.3	3	2.12	RT; FM	46	68	39	DM	46	190		SC	46				
Chlorpropham	5700		285	RT	47	3700		DM	47								
Chlorpyrifos	3	0.57	0.15	RT; FM	11; 12	0.1	0.04	DM	11					0.083d	0.041e	0.083	0.041

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Table D-1 (continued). Freshwater toxicity and regulatory guideline values.

Chemical	¹ Freshwater Toxicological and Reregistration Criteria													Freshwater Standards and Criterion			
	Fisheries					Invertebrate				Plant				² WAC		³ NRWQC	
	Acute	Chronic	ESLOC	Spp.	Ref	Acute	Chronic	Spp.	Ref	Acute	Chronic	Spp.	Ref	Acute	Chronic	CMC	CCC
cis-Permethrin ⁿ	2.9;17	0.30/ 0.41	0.145	RT;CS-A FM-C	58	0.039	0.039/ 0.084	DM	58								
	0.79		0.0395	BG	58												
Clopyralid	1968000	N/A	98400	RT	59, 64	113000	N/A	DM	59, 64	6900		SC	59				
Cycloate	4500		225	RT	48	24000		DM	48								
DCPA	6600	N/A	330	RT	56	27000	N/A	DM	56	>12380		SC	56				
Diazinon	90	0.8	4.5	RT; BT	13; 14	0.8	0.17	DM	13	3700		SC	13			0.17	0.17
Dicamba I	28000		1400	RT	15	34600	16400	DM	15	3700	5	SC; AFA	15				
Dichlobenil	4930	330	246.5	RT	16; 17	6200	560	DM	17	1500	160	SC	17				
Dimethoate	6200	430	310	RT	18	3320	40	DM	18								
Diphenamid	97000		4850	RT	59	58000		DM	59								
Disulfoton Sulfone	9200		460	RT	20, 66	35.2	0.14/ 0.27	DM	20, 66								
Diuron	1950	26.4	97.5	RT; FM	21; 22	1400	200	DM	22	2.4		SC	22				
Endosulfan I	0.8	0.1	0.04	RT	23	166	2	DM	23					0.22 ^{b,f}	0.056 ^{c,f}	0.22 ⁱ	0.056 ⁱ
Endosulfan II	0.8	0.1	0.04	RT	23	166	2	DM	23					0.22 ^{b,f}	0.056 ^{c,f}	0.22 ⁱ	0.056 ⁱ
Endosulfan Sulfate	2.2		0.11	ND	23	580		DM	23								
Endrin Aldehyde																	
Eptam	14000		700	ND	24	6500		ND	24	1360		SC	24				
Ethoprop	1020	180	51	RT; FM	25	44	0.8	DM	25								
Fenarimol	2100	430	105	RT	67	6800	113	DM	67		100	SC	67				
Hexachlorobenzene	1000	3.68	50	CH-A; RT-C	59, 26	30	16	DM	26	30		SC	26				
Hexazinone	180000	17000	9000	RT; FM	27; 28	151600	20000	DM	27	7	4	SC	27				
	317000		15850	Chinook	27												
	246000		12300	Coho	27												
	317000		15850	Sockeye	27												
Imidacloprid	>83000	1200/ 2500	4150	RT	61	69	1800/ 3600	CT-A; DM-C	61								
						85200		DM	59								
Linuron	3000	<42	150	RT	49	120		DM	50	67		SC	49				

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Table D-1 (continued). Freshwater toxicity and regulatory guideline values.

Chemical	¹ Freshwater Toxicological and Reregistration Criteria													Freshwater Standards and Criterion			
	Fisheries					Invertebrate				Plant				² WAC		³ NRWQC	
	Acute	Chronic	ESLOC	Spp.	Ref	Acute	Chronic	Spp.	Ref	Acute	Chronic	Spp.	Ref	Acute	Chronic	CMC	CCC
Malathion	4.1	21	0.205	RT	30	1	0.06	DM	30								0.1
	170		8.5	Coho	31												
MCPA	1150	916	57.5	RT	32	280	77	DM	32	250	32	SC	32				
MCPP	93000	N/A	4650	RT	65	91000	50800/ 102700	DM	65	14	9	SC	65				
Metalaxyl	132000	9100	6600	RT; FM	51	29000	1270	DM	51	140000		SC	51				
Methiocarb	436		21.8	RT	C	19		DM	C								
Methomyl	860	57/ 117	43	RT-A; FM-C	57	5	>0.4	DM	57								
Methomyl Oxime																	
Metolachlor	3900	780	195	ND	33	25100		DM	33								
Metribuzin	77000		3850	RT	52	4200	1290	DM	52	11.9	8.9	NP	51				
Napropamide	6400	1100	320	RT	53	14300	1100	DM	53	3400		SC					
Norflurazon	8100	770/ 1500	405	RT	34	15000	1000/ 2600	DM	34	9.7	3.2	SC	34-A 59-C				
Oryzalin	3260		163	RT	D	1400		DM	D								
Oxamyl	4200	770/ 1500	210	RT	62	180	1000/ 4200	CP-A; DM-C	62	120	4.6	SC	62				
Oxamyl Oxime																	
Oxyfluorfen	250	38/74	12.5	RT-A; FM-C	35, 36	80	13/28	DM	35, 36	0.29	0.1	SC	35, 36				
Pendimethalin	138	6.3	6.9	RT; FM	37	280	14.5	DM	37	5.4	3	SC	37				
Pentachlorophenol	15	11	0.75	RT	38	450	240	DM	38	50		SC	38	8.2 to 41.0 ^{d,g}	5.2-25.9 ^{e,h}	7.9-107.6 ^j	6.1-82.6 ^k
Picloram	5500	N/A	275	RT	53	34400	N/A	DM	53								
Promecarb																	
Prometon	12000	9500	600	RT-A; FM-C	68	25700	3500/ 6800	DM	68	98	32	SC	68				
Propargite	118	16	5.9	RT; FM	40	74	9	DM	40	66.2	5	SC	40				
Propoxur	3700		185	RT	63	11		DM	63								
Simazine	70500	1200	3525	RT; FM	41	1100		DM	41	100		SC	41				

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Table D-1 (continued). Freshwater toxicity and regulatory guideline values.

Chemical	¹ Freshwater Toxicological and Reregistration Criteria													Freshwater Standards and Criterion			
	Fisheries					Invertebrate				Plant				² WAC		³ NRWQC	
	Acute	Chronic	ESLOC	Spp.	Ref	Acute	Chronic	Spp.	Ref	Acute	Chronic	Spp.	Ref	Acute	Chronic	CMC	CCC
Tebuthiuron	143000	9300	7150	RT; FM	42	297000	21800	DM	42	50	13	SC	42				
Terbacil	46200		2310	RT	43	65000		DM	43	18	4	SC	43				
Triadimefon	4100	41/ 116	205	RT	55	1600	52/119	DM	55	100/1710		SC	55				
Triclopyr	650		32.5	RT	44	12000		DM	44	2300	2	SC; NP	44				
Trifluralin	41	1.14	2.05	RT	45	560	2.4	DM	45	7.52	5.37	SC	45				

*Values are not analytically qualified. Non-asterisk values have been J-qualified as estimates, normally below the practical quantitation limit.

¹Criteria identified in EPA reregistration and review documents or peer reviewed literature. References listed separately.

Time component of standards are explained in body of report.

ESLOC refers to Endangered Species Level of Concern.

Species abbreviated in table: RT-Rainbow Trout, CS-Coho Salmon, CH-Chinook salmon, FM- Fathead Minnow, BT-Brook Trout, BS-Bluegill Sunfish, ND-Not Described,

DM-Daphnia magna, CD-Ceriodaphnia dubia, SC-Selenastrum capricornutum (aka; Pseudokirchneria subcapitata), Anabaena flos-aquae, and Navicula pelliculosa,

SM-sheepshead Minnow, CT-Chironomus tentans (midge).

²WAC: Promulgated standards according to Chapter 173-201A WAC.

³EPA National Recommended Water Quality Criteria (EPA-822-R-02-047).

CMC: Criteria Maximum Concentration; estimate of the highest concentration of a material in surface water to which an aquatic community can be exposed briefly without resulting in an unacceptable effect.

CCC: Criteria Continuous Concentration; estimate of the highest concentration of a material in surface water to which an aquatic community can be exposed indefinitely without resulting in an unacceptable effect.

a -Criteria applies to DDT and its metabolites (ΣDDT).

b-An instantaneous concentration not to be exceeded at any time.

c-A 24-hour average not to be exceeded.

d-A 1-hour average concentration not to be exceeded more than once every three years on average.

e-A 4-day average concentration not to be exceeded more than once every three years on average.

f-Chemical form of endosulfan is not defined in WAC 173-201A. Endosulfan sulfate may be applied in this instance.

g≤ e[1.005(pH)-4.830], pH range of 6.9 to 9.5 shown.

h≤ e[1.005(pH)-5.29], pH range of 6.9 to 9.5 shown.

i-Value refers to Σα and β-endosulfan.

j≤ e[1.005(pH)-4.869], pH range of 6.9 to 9.5 shown.

k≤ e[1.005(pH)-5.134], pH range of 6.9 to 9.5 shown.

m-There are many forms of 2,4-D that include acids, salts, amines, and esters all of which have unique toxicity values. The criteria presented are in acid equivalents and are intended to provide a range of possible effects. Toxicity values for each form of 2,4-D are available in the referenced document.

n-Assessment criteria for permethrin are based on a formulation of cis and trans-permethrin isomers. Manchester Laboratory analysis includes only the cis-permethrin isomer, the more toxic of the two; and cis-permethrin concentrations are compared to the assessment criteria for permethrin.

Table D-2. Marine toxicity and regulatory guideline values for three estuarine sites. All values are reported in ug/L

Chemical	EPA Marine Toxicological and Registration Criteria													Marine Standards and Criterion			
	Fisheries					Invertebrate				Plant				² WAC		³ NRWQC	
	Acute	Chronic	ESLOC	Spp.	Ref	Acute	Chronic	Spp.	Ref	Acute	Chronic	Spp.	Ref	Acute	Chronic	CMC	CCC
1-Naphthol	1200		60	SM	10	2100		EO	10								
2,4-D (Acids, Salts, Amines) ^m	>80,000 (175,000 definitive)	no data	4000	TS	1	57000	no data	EO	1								
2,4-D (BEE Ester) ^m	no data	555		SM	1	1800	no data	EO	1								
2,4'-DDD																	
2,4'-DDT																	
3-Hydroxycarbofuran	33	2.6	1.65	AS; SM	60	4.6	0.4	PS; MS	60								
4,4'-DDD																	
4,4'-DDE														0.13 ^a	0.001 ^b		
4,4'-DDT														0.13 ^a	0.001 ^b		
4-Nitrophenol																	
Alachlor																	
Aldicarb																	
Aldicarb Sulfone																	
Aldicarb Sulfoxide																	
Atrazine	2000	2542	100	SM	4	94	80	AT; M	4	22		IG	4				
Azinphos Methyl																	
Bentazon	136		6.8	SM	6	>132.5; >109		PS; EO	6								
Bromacil	162		8.1	SM		12.9; 130		M; EO	7								
Bromoxynil																	
Carbaryl	2600		130	SM	10	32; >2		PS; EO	10								
Carbofuran	33	2.6	1.65	AS; SM	60	4.6	0.4	PS; MS	60								
Chlorothalonil	32		1.6	SM	46	154; 3.6	1.2	PS; EO; M	46								
Chlorpropham																	
Chlorpyrifos	270	0.38	13.5	SM; TS	11	2.4	<0.0046	PS; M	11					0.011 ^c	0.0056 ^d	0.011 ^G	0.0056 ^G
cis-Permethrin ⁿ	2.2	0.83	0.11	AS; SM	58	0.019	0.011	M	58								
Clopyralid																	
Cycloate																	

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Table D-2 (continued). Marine toxicity and regulatory guideline values for three estuarine sites.

Chemical	EPA Marine Toxicological and Registration Criteria													Marine Standards and Criterion			
	Fisheries					Invertebrate				Plant				² WAC		³ NRWQC	
	Acute	Chronic	ESLOC	Spp.	Ref	Acute	Chronic	Spp.	Ref	Acute	Chronic	Spp.	Ref	Acute	Chronic	CMC	CCC
DCPA	>1000		50	SM	56	620		EO	56	>11000		SkC	56				
Diazinon						4.2	0.23	M	13							0.82	0.82
Dicamba I	>180000		>9000	SM	15												
Dichlobenil	14000		700	SM	16	>1000; 2500		PS; EO	16								
Dimethoate																	
Diphenamid																	
Disulfoton Sulfone																	
Diuron	6700		335	SM	22		270	M	22								
Endosulfan I														0.034 ^a	0.0087 ^b	0.034 ^a	0.0087 ^b
Endosulfan II														0.034 ^a	0.0087 ^b	0.034 ^a	0.0087 ^b
Endosulfan Sulfate																	
Endrin Aldehyde																	
Eptam																	
Ethoprop																	
Fenarimol																	
Hexachlorobenzene																	
Hexazinone																	
Imidacloprid	163000		8150	SM	61	37	>0.6/1.3	MS	61								
Linuron	890		44.5	SM	49	4500; 890		M; EO									
Malathion																	
MCPA	>4100	4100	>205	SM	32	150000	115000	EO	32	300	15	SkC	32				
MCPP																	
Metalaxyl						25700; 4600		M; EO	51								
Methiocarb																	
Methomyl	1160		58	SM	57	>140000; 230		EO; M	57								
Methomyl Oxime																	
Metolachlor	7900	1000	395	ND	33												

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Table D-2 (continued). Marine toxicity and regulatory guideline values for three estuarine sites.

Chemical	EPA Marine Toxicological and Registration Criteria													Marine Standards and Criterion			
	Fisheries					Invertebrate				Plant				² WAC		³ NRWQC	
	Acute	Chronic	ESLOC	Spp.	Ref	Acute	Chronic	Spp.	Ref	Acute	Chronic	Spp.	Ref	Acute	Chronic	CMC	CCC
Metribuzin	85000		4250	SM	52	48300; 49800		M; EO	52	8.7	5.8	SkC	52				
Napropamide	14000		700	SM	53	4200; 1400		M; EO									
Norflurazon																	
Oryzalin																	
Oxamyl	2600		130	SM	62	0.4		EO	62								
Oxamyl Oxime																	
Oxyfluorfen																	
Pendimethalin																	
Pentachlorophenol	240		12	SM	38	48		PO	38	27		SkC	38	13.0 ^c	7.9 ^d		
Picloram																	
Promecarb																	
Prometon	47300		2365	SM	68	18000		MS	68								
Propargite																	
Propoxur																	
Simazine	>4300		215	SM	41	113000; >3700		PS; EO	41	600		SkC	41				
Tebuthiuron						62000		PS	42	31		SkC	42				
Terbacil																	
Triadimefon																	
Triclopyr	450		22.5	TS	44	2470		GS	44	1170	209	SkC	44				
Trifluralin	190		9.5	SM	45	638.5		GS	45	28		SkC	45				

*Values are not analytically qualified. Non-asterisk values have been J-qualified as estimates, normally below the practical quantitation limit.

¹Criteria identified in EPA registration and review documents or peer reviewed literature. References listed separately.

Time component of standards are explained in body of report.

ESLOC refers to Endangered Species Level of Concern

Species abbreviated in table: ND-Not determined, AS-Atlantic silverside, IS-Inland silverside, TS-Tidewater silverside, PS-Pink Shrimp, EO-Eastern Oyster, AT-Acartia tonsa (copepod), M-Mysid, IG-Isochrysis galbana, LG-Lemna gibba, CT-Chironomus tentans (midge), GS - Grass Shrimp, SkC - Skeletonema costatum, PO-Pacific Oyster

²WAC: Promulgated standards according to Chapter 173-201A WAC.

³EPA National Recommended Water Quality Criteria (EPA-822-R-02-047).

CMC: Criteria Maximum Concentration; estimate of the highest concentration of a material in surface water to which an aquatic community can be exposed briefly without resulting in an unacceptable effect.

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CCC: Criteria Continuous Concentration; estimate of the highest concentration of a material in surface water to which an aquatic community can be exposed indefinitely without resulting in an unacceptable effect.

a-Criteria applies to DDT and its metabolites (Σ DDT).

b-An instantaneous concentration not to be exceeded at any time.

c-A 24-hour average not to be exceeded.

d-A 1-hour average concentration not to be exceeded more than once every three years on average.

e-A 4-day average concentration not to be exceeded more than once every three years on average.

f-Chemical form of endosulfan is not defined in WAC 173-201A. Endosulfan sulfate may be applied in this instance.

g $\leq e[1.005(\text{pH})-4.830]$, pH range of 6.9 to 9.5 shown.

h $\leq e[1.005(\text{pH})-5.29]$, pH range of 6.9 to 9.5 shown.

i-Value refers to $\Sigma\alpha$ and β -endosulfan.

j $\leq e[1.005(\text{pH})-4.869]$, pH range of 6.9 to 9.5 shown.

k $\leq e[1.005(\text{pH})-5.134]$, pH range of 6.9 to 9.5 shown.

m-There are many forms of 2,4-D that include acids, salts, amines, and esters all of which have unique toxicity values. The criteria presented are in acid equivalents and are intended to provide a range of possible effects. Toxicity values for each form of 2,4-D are available in the referenced document.

n-Assessment criteria for permethrin are based on a formulation of cis- and trans-permethrin isomers. Manchester Laboratory analysis includes only the cis-permethrin isomer, the more toxic of the two; and cis-permethrin concentrations are compared to the assessment criteria for permethrin.

References for Appendix D

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Appendix E. Pesticide Calendars

To determine if water quality concentrations were healthy for aquatic life, monitoring data were compared to EPA pesticide registration toxicity criteria and EPA National Recommended Water Quality Criteria (NRWQC), referred to as *assessment criteria* in this report. Data were also compared to numeric Washington State water quality standards, referred to as *water quality standards*. Refer to Appendix D, *Assessment Criteria and Water Quality Standards*, in this report for information on assessment criteria development.

Table E-1 presents the color codes used to compare detected pesticide concentrations to assessment criteria.

Table E-1. Color codes for comparison to assessment criteria in the pesticide calendars.

	Each square represents the period when a sample was taken. If blank, then no pesticide residue detected.
	Analysis not completed.
	Pesticide residue detected. Assessment criteria not available.
	Detection of pesticide residue, concentration below regulatory or toxicological endpoint.
	Magnitude of detection above an EPA ¹ acute or chronic invertebrate registration endpoint.
	Magnitude of detection above a WAC ² or NRWQC ³ acute or chronic regulatory endpoint.
	Magnitude of detection above the ESLOC ⁴ for fish, which is 1/20th of the acute toxicity endpoint.

¹ EPA = United States Environmental Protection Agency

² WAC = Washington Administrative Code

³ NRWQC = EPA's National Recommended Water Quality Criteria

⁴ ESLOC = Endangered Species Level of Concern

Detection of a pesticide concentration above an assessment criteria does not indicate exceedance of (not meeting) the regulatory criteria. The temporal component of the criteria must also be exceeded. The Washington State Department of Agriculture (WSDA) advises pesticide user groups and other stakeholders on the results of this study and determines if assessment criteria are exceeded. If an exceedance is determined, WSDA advises stakeholders of appropriate measures to reduce pesticide concentrations.

For additional information on pesticide assessment criteria, contact the Washington State Department of Agriculture, Natural Resources Assessment Section, toll free at (877) 301-4555, #6 or (360) 902-2067, or e-mail: nras@agr.wa.gov. Their web site is <http://agr.wa.gov/PestFert/natresources/SWM/>.

Cedar-Sammamish Basin

Thornton Creek

A total of 19 pesticides and degradates were detected in Thornton Creek in 2009 (Table E-2).

No detections were above assessment criteria or water quality standards.

Table E-2. Thornton Creek 2009.

Month		March				April				May				June				July				August				September		
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
2,4,6-Trichlorophenol	D-M								0.510																			
2,4-D	H									0.110	0.037	0.130	0.019									0.020						0.040
3-Hydroxycarbofuran	D-C									0.054		0.076																
4-Nitrophenol	D-M			0.120																								
Carbaryl	I-C					0.025																						
Carbofuran	I-C					0.031																						
Chlorothalonil	F					0.028																						
Dicamba I	H											0.010																
Dichlobenil	H		0.023	0.046	0.017	0.010	0.025	0.014	0.012	0.053	0.017	0.049	0.015	0.020	0.020	0.011	0.012	0.014	0.037	0.018	0.014	0.027	0.024	0.030	0.028	0.024	0.026	0.051
Diuron	H				0.057																							
MCPP	H			0.041						0.042		0.086																
Methiocarb	I-C	0.099	0.215																									
Methomyl	I-C	0.065																										
Methomyl oxime	D-C											0.079																
Oxamyl oxime	D-C					0.028																						
Pentachlorophenol	WP												0.007														0.015	0.024
Prometon	H							0.075	0.039																			
Propoxur	I-C																	0.053										
Triclopyr	H									0.080		0.040																0.044
Total Suspended Solids		3	7	17	5	7	6	6	6	25	4	11	7	10	5	11	4	3	4	4	4	3	3	3	2	3	3	4

D = Degradate, F = Fungicide, H = Herbicide, I = Insecticide, WP = Wood Preservative, C = Carbamate, M = Multiple sources

Green-Duwamish Basin

Longfellow Creek

A total of nine pesticides and degradates were detected in Longfellow Creek in 2009 (Table E-3).

No detections were above assessment criteria or water quality standards.

Table E-3. Longfellow Creek 2009.

Month		March				April				May				June				July				August				September		
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
2,4,6-Trichlorophenol	D-M			0.510																								
2,4-D	H									0.110	0.038	0.085				0.058	0.110							0.042	0.022		0.027	0.035
3,5-Dichlorobenzoic Acid	D-M			0.520																								
Dichlobenil	H		0.046	0.010	0.016	0.013	0.047	0.014	0.014	0.130	0.019	0.025	0.013	0.012	0.011	0.008	0.010	0.009	0.022	0.011	0.011	0.023	0.021	0.021	0.030	0.025	0.030	0.033
MCPA	H											0.025																
MCPP	H									0.051	0.009																	
Methiocarb	I-C	0.117	0.200																									
Pentachlorophenol	WP		0.028							0.037	0.009																	0.020
Triclopyr	H		0.095							0.110	0.024	0.071				0.014	0.098					0.015		0.047	0.048	0.034	0.052	0.074
Total Suspended Solids		13	20	3	3	2	7	3	3	38	2	5	4	5	16	4	3	6	3	18	2	2	2	1		3	2	1

D = Degradate, H = Herbicide, I = Insecticide, WP = Wood Preservative, C = Carbamate, M = Multiple sources

Skagit-Samish Basins

Big Ditch

A total of 27 pesticides and degradates were detected in Big Ditch in 2009. Of these, 18 were found at the upper Big Ditch site (Table E-4). A total of 23 pesticides and degradates were found at the lower Big Ditch site (Table E-5).

At the upper Big Ditch site, one detection of malathion did not meet (exceeded) the Endangered Species Level of Concern (ESLOC) for freshwater fish and the EPA chronic invertebrate assessment criteria in May 2009. No detections were above assessment criteria or water quality standards at the lower Big Ditch site.

The malathion ESLOC is 1/20th of the 50% mortality concentration with a 96-hour exposure time (LC50) for rainbow trout in the lab. A single detection in a weekly sample does not prove or disprove that the 96-hour time component of the ESLOC was exceeded.

The EPA chronic invertebrate criteria for malathion is based on a 21-day No Observable Effects Concentration (NOEC) exposure for *Daphnia magna* (water flea). The single detection of malathion did not exceed the time component of this standard.

Comparison of Upper Big Ditch to Lower Big Ditch

In 2009, the upper and lower sites on Big Ditch were sampled weekly on the same day. During the year, 14 pesticides were detected in common between the two sites: 2,4-D; 3-hydroxycarbofuran; 4-nitrophenol; bromacil; chlorothalonil; dicamba I; dichlobenil; MCPA; MCPP; metalaxyl; methiocarb; metolachlor; pentachlorophenol; and triclopyr.

Four compounds were detected only at the upper site: imidacloprid, malathion, picloram, and tebuthiuron. Nine compounds were detected only at the lower site: atrazine, bentazon, carbaryl, carbofuran, diuron, eptam, ethoprop, metribuzin, and trifluralin.

Table E-4. Lower Big Ditch 2009.

Month		March				April				May				June				July					August					September			
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37			
2,4-D	H						0.210				0.950	1.100	0.370						0.024		0.021						0.045				0.037
3-Hydroxycarbofuran	D-C										0.074																				
4-Nitrophenol	D-M											0.110																			
Atrazine	H							0.076			0.860	0.150																			
Bentazon	H							0.086					0.040																		
Bromacil	H		0.047		0.045			0.069	0.071			0.062	0.046	0.025													0.043		0.026		
Carbaryl	I-C						0.024																								
Carbofuran	I-C											0.102																			
Chlorothalonil	F																0.014	0.072													
Dicamba I	H										0.125	0.250	0.089	0.012																	
Dichlobenil	H		0.018	0.009	0.012	0.010	0.110	0.016	0.013	0.110	0.073	0.032	0.011													0.019	0.022				
Diuron	H				0.140																										
Eptam	H					0.360	0.130	0.200																							
Ethoprop	I-OP										0.160	0.740	0.310																		
MCPA	H					0.093	0.190	1.100			0.155		0.060																		
MCPP	H	0.029									0.083	0.260	0.052																		
Metalaxyl	F																													0.160	
Methiocarb	I-C	0.075	0.085																												
Metolachlor	H	0.035	0.054	0.084	0.059	0.160	0.500	0.058	0.085	1.200	0.400	1.900	0.059	0.023	0.018																
Metribuzin	H												0.200																		
Pentachlorophenol	WP										0.052	0.036	0.015																		
Triclopyr	H						0.097				0.480	0.220	0.140													0.046		0.040			
Trifluralin	H			0.019																											
Total Suspended Solids		30	24	35	38	22	19	8	12	11	31	8	5	12	13	4	4	5	10	3	3	2					3	4	2		

D = Degradate, F = Fungicide, H = Herbicide, I = Insecticide, WP = Wood Preservative, C = Carbamate, M = Multiple sources, OP = Organophosphate

Table E-5. Upper Big Ditch 2009.

Month		March				April				May				June				July					August				September	
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
2,4-D	H			0.270			0.088			0.220	1.200	0.840				0.510			0.023		0.480							
3-Hydroxycarbofuran	D-C																			0.054								
4-Nitrophenol	D-M			0.150																								
Bromacil	H	0.140	0.120		0.100	0.120	0.074	0.120	0.140		0.070	0.170	0.100	0.190	0.180	0.170	0.145	0.220	0.170	0.210	0.190	0.110	0.130	0.088	0.150	0.120	0.120	0.120
Chlorothalonil	F											0.017																
Dicamba I	H		0.022				0.028			0.035	0.380	0.042																
Dichlobenil	H		0.019	0.050	0.013	0.014	0.095	0.025	0.021	0.071	0.067	0.055	0.017	0.016	0.013	0.010	0.011	0.010		0.020	0.015	0.027	0.037	0.028	0.026	0.022	0.028	
Imidacloprid	I-N								0.107	0.082	0.026		0.029	1.740	0.091	0.025		0.026	0.071		0.025			0.057				
Malathion	I-OP											0.940																
MCPA	H									0.077	0.092																	
MCPP	H	0.110	0.150	0.210						0.051	0.200	0.120																
Metalaxyl	F								0.330		0.051			0.075					1.300				0.075	0.096				
Methiocarb	I-C	0.095	0.110																									
Metolachlor	H											0.021																
Pentachlorophenol	WP											0.018									0.009	0.027						0.021
Picloram	H	0.120				0.057			0.220				0.087	0.060	0.130	0.180	0.210	0.210	0.087	0.067	0.063		0.150	0.065	0.040		0.035	
Tebuthiuron	H		0.031			0.023		0.032	0.044					0.030			0.029	0.036	0.044	0.039	0.035			0.037	0.032			
Triclopyr	H						0.043			0.160	0.210	0.260					0.350			0.021		0.360						0.051
Total Suspended Solids		10	6	118	7	4	14	5	3	17	15	59	6	9	11	9	7	13	8	14	19	11	10	37	3	8	6	28

D = Degradate, F = Fungicide, H = Herbicide, I = Insecticide, WP = Wood Preservative, C = Carbamate, M = Multiple sources, N = Neonicotinoid, OP = Organophosphate

Indian Slough

A total of 21 pesticides and degradates were detected in Indian Slough in 2009 (Table E-6).

One detection of malathion at Indian Slough exceeded the ESLOC for freshwater fish and the EPA chronic invertebrate assessment criteria in March 2009. The malathion ESLOC is 1/20th of the LC50 for rainbow trout in the lab. A single detection in a weekly sample does not prove or disprove the 96-hour time component of the ESLOC was exceeded.

The EPA chronic invertebrate assessment criteria for malathion is based on a 21-day exposure for *Daphnia magna* (water flea). The single detection of malathion did not exceed the time component of this criterion.

Table E-6. Indian Slough 2009.

Month		March				April				May				June				July				August			September				
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37	
2,4-D	H						0.092		0.065	0.130	1.100	0.240						0.210		0.050			0.056		0.085				
4-Nitrophenol	D-M																			0.026									
Atrazine	H							0.200	0.080	0.049	0.058	0.039																	
Bentazon	H													0.023					0.033	0.025	0.023	0.021			0.017				
Bromacil	H	0.055	0.060	0.059	0.055	0.044	0.086	0.048	0.067	0.044	0.097	0.110	0.041	0.033		0.037		0.084		0.052		0.028			0.060		0.022		
Carbofuran	I-C						0.021																						
Diazinon	I-OP										0.019				0.017											0.034			
Dicamba I	H																			0.010									
Dichlobenil	H		0.027	0.037	0.015	0.013	0.490	0.019	0.013	0.020	0.085	0.110	0.012	0.009	0.011		0.006			0.013	0.009					0.031			
Diphenamid	H	0.020						0.021						0.017	0.018	0.018	0.017	0.030	0.012	0.034	0.032			0.020	0.016	0.013	0.018	0.015	0.020
Hexazinone	H						0.500				0.210	0.240	0.071	0.070	0.064	0.068	0.051	0.065		0.063	0.057				0.065				
Imidacloprid	I-N		0.024								0.023																		
Malathion	I-OP			0.900																									
MCPA	H								0.093	0.091		0.035																	
MCPP	H						0.031																						
Metalaxyl	F																							0.036					
Methomyl	I-C	0.074																											
Metolachlor	H											0.170	0.022						0.037	0.051	0.029				0.037				
Pentachlorophenol	WP			0.018							0.018																		
Tebuthiuron	H			0.040	0.036	0.046		0.059	0.071	0.037			0.039	0.039	0.037	0.033	0.035	0.051	0.052	0.044	0.044	0.038	0.036		0.049		0.058		
Triclopyr	H								0.059	0.710	0.230							0.120		0.014			0.028	0.020	0.160				
Total Suspended Solids		23	15	13	12	9	15	9	12	9	16	9	9	6	6	2	4	5	4	5	5	5	3		2	4	11	3	3

D = Degradate, F = Fungicide, H = Herbicide, I = Insecticide, WP = Wood Preservative, C = Carbamate, M = Multiple sources, N = Neonicotinoid, OP = Organophosphate

Browns Slough

A total of 12 pesticides and degradates were detected in Browns Slough in 2009 (Table E-7). No detections were above assessment criteria or water quality standards.

Table E-7. Browns Slough 2009 – Freshwater and Marine Criteria.

Month		March				April				May				June				July				August				September		
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
2,4-D	H							0.061		0.140	0.056																	
Bentazon	H																											
Carbofuran	I-C										0.026																	
DCPA	H	0.520	0.420	0.049	0.900	0.910	0.150	0.360	0.080	0.072		0.120	0.025				0.015			0.025								
Dicamba I	H									0.040																		
Dichlobenil	H					0.005	0.010			0.011			0.007		0.007													
Eptam	H								0.840	0.086																		
Metolachlor	H						0.400	0.075	0.130	0.090	0.048	0.036																
Metribuzin	H													0.030	0.049													
Pentachlorophenol	WP										0.130																	
Simazine	H	0.046	0.085		0.043	0.022	0.034				0.025	0.026																
Triclopyr	H										0.038																	
Total Suspended Solids		9	14	9	9	9	15	13	11	7	7	5	8	4	7	4	8	9	9	8	18	8	8	13	5	5	7	7

H = Herbicide, I = Insecticide, WP = Wood Preservative, C = Carbamate

Samish River

A total of eight pesticides were detected in the Samish River in 2009 (Table E-8). No detections were above assessment criteria or water quality standards.

Table E-8. Samish River 2009.

Month		March				April				May				June				July				August				September		
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
2,4-D	H									0.068	0.125										0.021							
Dicamba I	H									0.014	0.016																	
Dichlobenil	H		0.010	0.005			0.010			0.010	0.013		0.007															
Hexazinone	H										0.071																	
MCPA	H										0.085					0.019												
Metolachlor	H			0.015							0.020					0.012												
Pentachlorophenol	WP																											0.015
Triclopyr	H									0.038	0.059																	
Total Suspended Solids		8	13	18	8	20	60	14	9	20	89	16	12	6	8	7	3	3	2	3	4	4	5	2	4	4	8	4

H = Herbicide, WP = Wood Preservative

Lower Yakima Basin

Spring Creek

A total of 19 pesticides and degradates were detected in Spring Creek in 2009. Of these, 15 were detected at the upper Spring Creek site (Table E-9) and 18 were detected in the lower Spring Creek site (Table E-10).

At the upper Spring Creek site, one April sample of 4,4'-DDE was above the chronic water quality standard and NRWQC for fish. A single detection does not prove or disprove the 24-hour time component of this chronic criterion was exceeded.

At the lower Spring Creek site three consecutive detections of chlorpyrifos from March to April were above the chronic water quality standard and NRWQC for fish, and EPA chronic invertebrate criteria. Consecutive detections in three weeks show the 4-day time component of the chlorpyrifos water quality standard and NRWQC were exceeded. In addition, the chronic invertebrate 21-day exposure criterion was also likely exceeded.

Comparison of Upper Spring Creek to Lower Spring Creek

In 2009, the upper and Spring Creek site was sampled biweekly and the lower site was sampled weekly. During the year, 14 chemicals were detected in common between the two sites: 2,4-D; atrazine; bentazon; carbaryl; chlorpyrifos; diazinon; dicamba I; dichlobenil; MCPA; norflurazon; oryzalin; pendimethalin; propoxur; and simazine.

One DDT degradate was detected only at the upper site: 4,4'-DDE. Four compounds were detected only at the lower site: bromacil, endosulfan sulfate, imidan, and pentachlorophenol.

Table E-9. Upper Spring Creek 2009.

Month		March		April		May		June		July			August		Sep
Chemical	Type	11	13	15	17	19	21	23	25	27	29	31	33	35	37
2,4-D	H					0.084	0.020				0.079	0.046			0.028
4,4'-DDE	D-OC				0.011										
Atrazine	H	0.023		0.015						0.025		0.024			0.020
Bentazon	H	0.035	0.040				0.025	0.016							0.029
Carbaryl	I-C			0.031											
Chlorpyrifos	I-OP			0.029	0.033										
Diazinon	I-OP								0.069		0.077	0.027			
Dicamba I	H					0.046					0.010				0.017
Dichlobenil	H			0.004	0.013	0.009					0.009				
MCPA	H					0.027									
Norflurazon	H			0.030					0.025		0.066				
Oryzalin	H				0.300	0.150			0.086		0.310				
Pendimethalin	H					0.027	0.022		0.021						
Propoxur	I-C												0.064		
Simazine	H			0.015											
Total Suspended Solids		7	4	8	27	68	19	27	59	25	29	18	12	7	4

D = Degradate, F = Fungicide, H = Herbicide, I = Insecticide, C = Carbamate, OC = Organochlorine, OP = Organophosphate

Table E-10. Lower Spring Creek 2009.

Month		March				April				May				June				July					August				September	
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
2,4-D	H									0.110	0.038	0.057	0.021	0.026	0.038	0.038	0.072	0.100	0.055	0.120	0.033	0.061	0.021		0.024	0.024	0.020	0.038
Atrazine	H	0.025	0.027																									
Bentazon	H			0.028																			0.012					
Bromacil	H	0.036	0.041	0.040	0.023	0.019	0.034	0.046	0.043	0.020		0.022								0.055	0.059		0.035		0.039		0.024	
Carbaryl	I-C					0.046																						
Chlorpyrifos	I-OP				0.045	0.076	0.046	0.028	0.024	0.020																		
Diazinon	I-OP															0.013			0.060		0.024							
Dicamba I	H									0.051		0.011							0.009	0.007		0.007		0.011		0.012	0.017	
Dichlobenil	H					0.005	0.009	0.012	0.009	0.009	0.009	0.006							0.009									
Endosulfan Sulfate	D-OC		0.022																									
Imidan	I-OP																											0.059
MCPA	H									0.024																	0.030	
Norflurazon	H	0.033	0.060	0.034												0.023			0.062									
Oryzalin	H						0.540			0.120																		
Pendimethalin	H		0.024				0.044	0.046		0.030	0.032	0.032	0.028	0.032		0.021												
Pentachlorophenol	WP										0.008																	
Propoxur	I-C																					0.099						
Simazine	H	0.024	0.045	0.020																								
Total Suspended Solids		5	3	1	14	12	8	9	25	30	50	20	28	19	49	11	5	2	3	17	8	14	5	7	7	8	8	7

D = Degradate, H = Herbicide, I = Insecticide, WP = Wood Preservative, C = Carbamate, OC = Organochlorine, OP = Organophosphate

Marion Drain

A total of 20 pesticides and degradates were detected in Marion Drain in 2009 (Table E-11).

Chlorpyrifos was found above the EPA chronic invertebrate assessment criteria once in 2009. This single event did not exceed the 21-day time component of the chronic invertebrate criteria. In 2009 no other detections were above assessment criteria or water quality standards.

Table E-11. Marion Drain 2009.

Month		March				April				May				June				July				August				September				October								
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37	38	39	40	41	42	43	44			
2,4-D	H								0.032	0.079	0.041	0.032	0.020	0.021	0.078	0.029	0.023	0.055	0.048	0.043	0.033		0.032	0.092	0.031	0.028	0.030	0.034										
Atrazine	H																	0.022																				
Bentazon	H													0.062	0.064	0.130	0.140	0.110	0.130	0.140	0.260	0.260	0.280	0.180	0.140	0.078	0.069	0.075										
Bromacil	H						0.026	0.028	0.042		0.020																											
Bromoxynil	H								0.065	0.073	0.039	0.030	0.017	0.010	0.008																							
Chlorpropham	H																									0.049												
Chlorpyrifos	I-OP				0.026	0.029	0.040		0.038		0.018			0.020	0.008													0.016	0.015				0.016					
Dicamba I	H								0.020	0.030	0.015	0.014			0.010	0.011	0.009	0.009	0.010	0.010	0.010	0.008	0.011	0.021	0.014	0.012	0.013	0.016										
Disulfoton sulfone	I-OP																									0.034	0.046	0.044		0.031								
Disulfoton sulfoxide	D-OP																									0.035			0.038	0.160		0.032	0.018					
Eptam	H										0.067																											
Ethoprop	I-OP										0.033																0.480	0.610	0.380	0.130	0.088	0.082	0.070					
Imidacloprid	I-N													0.041																								
Malathion	I-OP																		0.029	0.045																		
MCPA	H									0.026		0.020	0.009	0.013																								
Metolachlor	H																									0.091	0.120		0.037									
Pendimethalin	H								0.080	0.065	0.053	0.074	0.062	0.061	0.040	0.034	0.023	0.028																				
Simazine	H		0.023																																			
Terbacil	H							0.051	0.070	0.590	0.140	0.200	0.115	0.100	0.089	0.110	0.120	0.120	0.069	0.067	0.680	0.290	0.063	0.045	0.033	0.045	0.066	0.260	0.220	0.250	0.360	0.230	0.053					
Trifluralin	H								0.016	0.019	0.024	0.026	0.024	0.022	0.015	0.015	0.017	0.009																				
Total Suspended Solids		15	12	12	22	24	40	12	31	19	19	20	25	23	7	7	5	4	3	2	2	3	4	7	24	26	10	8	8	10	14	6	4	13	12			

D = Degradate, H = Herbicide, I = Insecticide, C = Carbamate, N = Neonicotinoid, OP = Organophosphate

Sulphur Creek Wasteway

A total of 21 pesticides and degradates were detected in Sulphur Creek Wasteway in 2009 (Table E-12).

The DDT degradate, 4,4'-DDE, was found above the chronic water quality standard and NRWQC in March and April. The chronic water quality standard for DDT and its metabolites are not met (exceeded) when the 24-hour average concentration exceeds the numerical criteria. Because 4,4'-DDE was detected in two consecutive weeks, the 24-hour time component of this criterion was exceeded.

Chlorpyrifos had three consecutive detections above the chronic water quality standard, NRWQC, and EPA chronic invertebrate assessment criteria in March and April. One of these detections was also above the ESLOC for fish. The chlorpyrifos chronic water quality standard and NRWQC are exceeded when the 4-day average concentration exceeds the numerical criteria. Because chlorpyrifos was detected in three consecutive weeks, the time component of the chronic water quality standard and NRWQC was exceeded.

The chlorpyrifos ESLOC is 1/20th of the LC50 for rainbow trout. A single detection in a weekly sample does not prove or disprove the 96-hour time component of the ESLOC was exceeded.

Table E-12. Sulphur Creek Wasteway 2009

Month		March				April				May				June				July					August					September	
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37	
2,4-D	H							0.032	0.074	0.170	0.041	0.051	0.077	0.028	0.055	0.097	0.230	0.110	0.052	0.062	0.049	0.074	0.050	0.055	0.061	0.071	0.040	0.041	
4,4'-DDE	D-OC			0.022	0.021				0.005																				
Atrazine	H																	0.046											
Bentazon	H	0.028	0.037																			0.015				0.012			
Bromacil	H	0.047	0.054	0.021	0.017	0.019	0.038	0.044	0.045		0.027	0.025					0.023	0.067	0.036	0.043				0.024		0.039	0.027	0.021	
Carbaryl	I-C	0.030	0.024					0.026			0.039							0.022											
Chlorpyrifos	I-OP				0.050	0.280	0.046	0.030			0.020																		
DCPA	H	0.019				0.032				0.018			0.005				0.013							0.023	0.030	0.033			
Diazinon	I-OP																		0.031			0.027		0.025			0.087		
Dicamba I	H							0.013	0.048	0.072	0.014	0.022	0.009		0.008	0.011	0.008		0.021	0.015	0.011	0.023	0.017	0.019	0.016	0.012		0.021	
Dichlobenil	H						0.012	0.009		0.011	0.010		0.007		0.007		0.007	0.007			0.009								
Dimethoate	I-OP											0.120																	
Hexazinone	H							0.110	0.099		0.047																		
MCPA	H									0.032					0.014				0.089			0.012	0.012						
Methiocarb	I-C	0.269																											
Metribuzin	H																0.420												
Norflurazon	H								0.044																				
Pendimethalin	H	0.043					0.039					0.024																	
Simazine	H																0.690												
Terbacil	H							0.120				0.039	0.024	0.033					0.039	0.045	0.039								
Trifluralin	H											0.018	0.021	0.022			0.015				0.032								
Total Suspended Solids		18	7	94	83	23	32	41	67	98	36	38	44	66	64	31	47	13	25	16	10	28	22	20	27	11	44	81	

D = Degradate, H = Herbicide, I = Insecticide, C = Carbamate, OC = Organochlorine, OP = Organophosphate

Wenatchee and Entiat Basins

Peshastin Creek

A total of 11 pesticides and degradates were detected in Peshastin Creek from 2007-09 (Tables E-13 to E-15).

Endosulfan I was detected above the ESLOC for rainbow trout once in April 2008 and once in April 2009. A single detection of azinphos methyl exceeded the chronic NRWQC in May 2007. No other detected compounds exceeded assessment criteria or water quality standards.

Table E-13. Peshastin Creek 2007.

Month		February			March				April				May				June				July				August			September				
Chemical	Type	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
Azinphos Methyl	I-OP															0.024																
Carbaryl	I-C									0.019																						
Methomyl	I-C																												0.023			
Oxamyl	I-C																											0.026				
Oxamyl oxime	D-C																										0.012					
Total Suspended Solids		2	6	2	3	218	25	12	4	13	3	3	5	31	11	12	7	12	3	3	2	2	2	2	2	2	1	2	1	15		

Table E-14. Peshastin Creek 2008.

Month		March				April				May				June				July				August			September				
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37	
Endosulfan I	I-OC					0.130																							
Endosulfan II	I-OC					0.046																							
Total Endosulfan	I-OC					0.176																							
Oxamyl	I-C						0.010																						
Total Suspended Solids		3	1	1	1	1	11	3	4	8	4	44	16	6	3	2	1	5	2	1	2	1	1	1	1				

Table E-15. Peshastin Creek 2009.

Month		March				April				May				June				July				August			Sep			
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
Endosulfan I	I-OC					0.040	0.013																					
Fipronil Sulfide	D-Pyra									0.015																		
Fipronil Sulfone	D-Pyra									0.016																		
Simazine	H				0.014																							
Simetryn	H																											0.055
Total Suspended Solids		2		3	3	25	14	52	4	5	6	67	13	11	7	4	2	1	2	1								1

D = Degradate, H = Herbicide, I = Insecticide, OC = Organochlorine, Pyra = Pyrazole

Mission Creek

A total of 11 pesticides, a degradate, and a pesticide synergist were detected in Mission Creek from 2007-09 (Tables E-16 to E-18).

One detection of endosulfan I was above the ESLOC for fish in April 2008. No other detections were above assessment criteria or water quality standards.

Table E-16. Mission Creek 2007.

Month		February			March				April					May				June				July					August			September		
Chemical	Type	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
Chlorpyrifos	I-OP						0.024																									
Endosulfan I	I-OC							0.017																								
Endosulfan II	I-OC							0.022																								
Total Endosulfan	I-OC							0.039																								
Methiocarb	I-C									0.034					0.015																	
Methomyl	I-C																			0.019												
Norflurazon	H																											0.027	0.041			
Oxamyl oxime	D-C																								0.017						0.018	
Total Suspended Solids		9	42	17	9	685	82	42	16	33	5	4	31	8	7	30	4	4	3	4	4	4	4	9	3	2	1	4	2			3

Table E-17. Mission Creek 2008.

Month		March				April				May				June				July					August			September						
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37				
Carbaryl	I-C												0.014																			
Endosulfan I	I-OC					0.047																										
Norflurazon	H																								0.034		0.018	0.018				
Simazine	H																							0.019								
Total Suspended Solids		8	3	2	2	2	42	3	8	17	5	25	32	10	8	5	4	4	5	3	2	2	3	2	2	4	1	1				

Table E-18. Mission Creek 2009.

Month		March				April				May				June					July					August			Sep					
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37				
3-Hydroxycarbofuran	D-C																					0.051										
Endosulfan I	I-OC					0.024																										
Piperonyl Butoxide	Sy					0.095																										
Total Suspended Solids		6	3	11	10	73	42	85	13	23	13	71	14	11	16	17	8	6	8	8	5	20	5	5	3	2				41		

D = Degradate, I = Insecticide, Sy = Pesticide synergist, C = Carbamate, OC = Organochlorine, OP = Organophosphate

Brender Creek

A total of 28 pesticides and degradates were detected in Brender Creek from 2007-09 (Tables E-19 to E-21).

Endosulfan was detected above the ESLOC for rainbow trout between March and May in six samples in 2007, eight samples in 2008, and four samples in 2009. Each year showed consecutive detections in two or more weeks. The endosulfan ESLOC is 1/20th of the LC50 for rainbow trout. Consecutive detections in two or more weeks show the 96-hour time component of the ESLOC criteria for endosulfan was not met (exceeded).

Three detections of chlorpyrifos in March and April of 2007 and 2009 were above the chronic water quality standard, NRWQC; and the EPA acute and chronic assessment criteria for invertebrates. The two 2009 detections were in consecutive weeks. Consecutive weekly chlorpyrifos detections over numerical criteria indicate the 4-day exposure criteria for the chronic water quality standard and NRWQC were exceeded. The 21-day exposure criterion for the EPA chronic criteria was probably not exceeded with only two consecutive weeks of detections. The single detection over the EPA acute invertebrate criteria in week 17 is not enough data to determine if the 48-hour time component of this criterion was exceeded.

DDT was found consistently throughout all three years, except for one week in April 2008 and two weeks in May and June 2009. The weeks where no detections of DDT and DDT breakdown products occurred coincided with some of the lower TSS detections for all years. All DDT and DDT metabolite concentrations were above the chronic water quality standard and NRWQC. The chronic water quality standard is based on a 24-hour average concentration.

Table E-19. Brender Creek 2007.

Month		February			March				April				May				June				July					August					September			
Chemical	Type	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37		
2,4'-DDD	D-OC								0.018									0.008																
2,4'-DDT	D-OC												0.017		0.011	0.009	0.011																	
4,4'-DDD	D-OC		0.025	0.024	0.023	0.020	0.018		0.020		0.016			0.013		0.012		0.009	0.009	0.010										0.004				
4,4'-DDE	D-OC		0.046	0.032	0.034	0.036	0.036	0.019	0.034	0.022	0.014	0.024	0.071	0.026	0.027	0.042	0.030	0.019	0.039	0.032	0.029	0.015	0.017	0.026	0.021	0.011	0.003	0.012	0.030		0.017	0.021		
4,4'-DDT	I-OC	0.016	0.036	0.027	0.026	0.021	0.019	0.023	0.023		0.024		0.050	0.021	0.019	0.025	0.027	0.017	0.025	0.033	0.027	0.017	0.013	0.020	0.018	0.013		0.025	0.029		0.017	0.018		
<i>DDT and metabolites</i>	OC	0.016	0.107	0.083	0.083	0.077	0.073	0.042	0.095	0.022	0.053	0.024	0.138	0.060	0.057	0.088	0.068	0.045	0.096	0.091	0.065	0.032	0.041	0.068	0.039	0.024	0.003	0.037	0.063		0.034	0.051		
Azinphos Methyl	I-OP																	0.033																
Carbaryl	I-C													0.010	0.023	0.040						0.012												
Chlorpyrifos	I-OP							0.110	0.038	0.027	0.030	0.027	0.019	0.015	0.007	0.007																		
Diazinon	I-OP									0.021																								
Diuron	H																																	
Endosulfan I	I-OC							0.096	0.020		0.096	0.026	0.050	0.019		0.014																		
Endosulfan II	I-OC							0.071			0.071	0.030	0.060	0.031	0.015	0.040																		
<i>Total Endosulfan</i>	I-OC							0.167	0.020		0.167	0.056	0.110	0.050	0.015	0.054																		
Endosulfan Sulfate	D-OC		0.034				0.015	0.043	0.032	0.041	0.073	0.034	0.100	0.043	0.038	0.057	0.032	0.021	0.027	0.024	0.024		0.020											
MCPA	H															0.072																		
Methomyl	I-C					0.017																												
Norflurazon	H													0.029	0.027	0.055		0.035	0.031	0.160	0.023			0.140		0.027		0.027						
Oxamyl	I-C																														0.027			
Prometon	H															0.009																		
Simazine	H															0.022								0.028										
Triadimefon	F															0.015																		
Total Suspended Solids		34	50	30	31	155	67	34	53	15	41	21	76	30	31	81	34	19	51	49	43	84	30	22	59	13	16	25	108		14	19		

D = Degradate, F = Fungicide, H = Herbicide, I = Insecticide, C = Carbamate, OC = Organochlorine, OP = Organophosphate

Table E-20. Brender Creek 2008.

Month		March				April				May				June				July					August				September	
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
2,4'-DDD	D-OC															0.015												
2,4'-DDT	D-OC									0.019	0.053																	
4,4'-DDD	D-OC			0.007					0.007		0.015	0.017	0.013	0.011	0.004	0.025	0.020	0.015	0.019	0.006	0.002	0.003	0.005	0.005	0.001	0.001	0.008	0.008
4,4'-DDE	D-OC	0.023					0.019	0.019	0.014	0.023		0.018	0.040	0.030	0.024	0.045	0.030	0.027	0.034	0.010	0.019	0.025	0.018	0.019	0.021	0.036	0.009	0.018
4,4'-DDT	I-OC	0.019	0.021	0.020	0.018		0.021	0.015	0.013	0.025	0.300	0.023	0.026	0.020	0.010	0.027	0.020	0.025	0.022	0.010	0.010	0.012	0.010	0.009	0.008	0.009	0.014	0.016
<i>DDT and metabolites</i>	OC	0.042	0.021	0.027	0.018		0.040	0.034	0.034	0.067	0.368	0.058	0.079	0.061	0.038	0.112	0.070	0.067	0.075	0.026	0.031	0.040	0.032	0.033	0.030	0.046	0.031	0.042
Carbaryl	I-C											0.024																
Chlorpyrifos	I-OP						0.028	0.015	0.009	0.025	0.019																	
Dichlobenil	H												0.008															
Diuron	H														0.220		0.036											
Endosulfan I	I-OC			0.060		0.049	0.046	0.048	0.089																			
Endosulfan II	I-OC			0.058				0.049	0.084	0.120	0.045	0.040	0.036	0.026														
<i>Total Endosulfan</i>	I-OC			0.118		0.049	0.046	0.097	0.173	0.120	0.045	0.040	0.036	0.026														
Endosulfan Sulfate	D-OC		0.016	0.016	0.018	0.032	0.045	0.047	0.110	0.160	0.066	0.066	0.061	0.050	0.026		0.033	0.048	0.037	0.022	0.017		0.029	0.023	0.013	0.011	0.016	0.014
Imidacloprid	I-N						0.060																	0.012				
Norflurazon	H						0.110			0.032		0.047			0.250		0.110						0.042	0.029	0.028		0.032	0.023
Oxamyl oxime	D-C						0.140																					
Simazine	H													0.012														
Total Suspended Solids		54	34	23	15	6	49	87	25	40	39	28	47	38	30	50	25	94	26	13	33	33	29	30	31	55	12	28

D = Degradate, H = Herbicide, I = Insecticide, C = Carbamate, N = Neonicotinoid, OC = Organochlorine, OP = Organophosphate

Table E-21. Brender Creek 2009.

Month		March				April				May				June				July				August				Sep		
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
2,4'-DDE	D-OC							0.009															0.007					
2,4'-DDT	D-OC																0.012		0.019									
4,4'-DDD	D-OC		0.023	0.023	0.023			0.019		0.014	0.013	0.013				0.012		0.025			0.029	0.029	0.030		0.030			
4,4'-DDE	D-OC	0.024	0.021	0.033	0.029	0.026	0.021	0.016	0.047	0.020	0.006	0.006			0.007	0.013	0.046	0.024	0.043	0.043	0.016	0.004	0.028	0.026	0.019	0.030	0.037	0.026
4,4'-DDT	I-OC	0.030	0.025	0.027	0.035		0.023	0.020	0.037	0.025	0.021	0.019			0.024	0.014	0.024	0.025	0.028	0.036		0.023	0.027	0.029			0.022	
<i>DDT and metabolites</i>	OC	0.054	0.069	0.083	0.087	0.026	0.044	0.064	0.084	0.059	0.040	0.038			0.031	0.027	0.094	0.049	0.115	0.079	0.016	0.056	0.091	0.085	0.019	0.060	0.037	0.048
3-Hydroxycarbofuran	D-C																								0.106			
Chlorpyrifos	I-OP					0.034	0.055	0.083	0.034	0.022	0.020																	
Dicamba I	H																									0.012		
Dichlobenil	H	0.010	0.010	0.005			0.009	0.008	0.010	0.010			0.007						0.030		0.009							
Endosulfan I	I-OC					0.100	0.027	0.018	0.058	0.036																		
Endosulfan II	I-OC					0.049	0.023		0.055	0.058	0.030	0.021																
<i>Total Endosulfan</i>	I-OC					0.149	0.050	0.018	0.113	0.094	0.030	0.021																
Endosulfan Sulfate	D-OC		0.022	0.022	0.021	0.048	0.044	0.044	0.098	0.084	0.050	0.043	0.032	0.029	0.028	0.027	0.030	0.029	0.035	0.031		0.034	0.031	0.031				
Imidacloprid	I-N																				0.022							
Methiocarb	I-C															0.033												
Norflurazon	H		0.031										0.028	0.048	0.039	0.032	0.028					0.045						
Piperonyl Butoxide	Sy					0.070																						
Simazine	H												0.096															
Total Suspended Solids		31	12	33	22	11	24	12	75	52	52	19	13	8	16	15	64	19	116	56	15	10	54	85	7	66	47	53

D = Degradate, H = Herbicide, I = Insecticide, Sy = Pesticide synergist, C = Carbamate, N = Neonicotinoid, OC = Organochlorine, OP = Organophosphate

Wenatchee River

A total of eight pesticides were detected in the Wenatchee River in 2009 (Tables E-22 to E-24).

Endosulfan I was detected above the ESLOC for rainbow trout, the chronic water quality standard, and the NRWQC once in March 2008 and once April 2009.

The single weekly detections of endosulfan are not enough data to determine if the 96-hour time component of the ESLOC for fish or the 24-hour time component of the chronic water quality standard and NRWQC were exceeded.

Table E-22. Wenatchee River 2007.

Date	2/14	2/21	2/27	3/6	3/13	3/20	3/28	4/3	4/9	4/18	4/25	5/2	5/9	5/16	5/21	5/30	6/6	6/13	6/18	6/27	7/2	7/10	7/17	7/23	7/31	8/7	8/14	8/22	8/28	9/5	9/10	
Month	February			March				April				May				June				July				August		September						
Chemical	Type	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
Chlorpyrifos	I-OP								0.035																							
Endosulfan I	I-OC								0.014																							
Methomyl	I-C										0.016																					
Oxamyl	I-C																												0.016			
Total Suspended Solids			4	2	2	102	16	10	7	8	3	3	5	18	13	17	12	25	4	4	3	4	4	3	2	3	4	4	1		2	2

Table E-23. Wenatchee River 2008.

Month	March				April				May				June				July				August		September								
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37			
Endosulfan I	I-OC				0.079	0.024																									
Endosulfan II	I-OC			0.025	0.076																										
Total Endosulfan	I-OC			0.025	0.155	0.024																									
Imidacloprid	I-N						0.028																								
Total Suspended Solids		3	2	2	1	2	9	3	6	10	4	46	14	12	9	13	6	17	8	3	3	2	2	3	4	3	1				

Table E-24. Wenatchee River 2009.

Month	March				April				May				June				July				August		Sep								
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37			
2,4-D	H																									0.018					
Chlorpyrifos	I-OP						0.038																								
Endosulfan I	I-OC					0.061																									
Pentachlorophenol	WP															0.014															
Total Suspended Solids		2	1	2	2	6	7	46	4	7	4	37	12	13	15	6	5	4	5	3	4	3	2	8	9	2	3	3			

H = Herbicide, I = Insecticide, WP = Wood Preservative, C = Carbamate, N = Neonicotinoid, OC = Organochlorine, OP = Organophosphate

Entiat River

A total of six compounds were detected in the Entiat River from 2007 to 2009 (Tables E-25 to E-27). No pesticide detections exceeded assessment criteria or water quality standards.

Table E-25. Entiat River 2007.

Month		February			March				April				May				June				July				August			September				
Chemical	Type	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
Carbaryl	I-C													0.016																		
Chlorpyrifos	I-OP								0.034																							
Dichlobenil	H																									0.065						
Total Suspended Solids		4	4	2	2	64	13	7	3	8	3	5	10	41	20	9	11	18	6	5	3	3	4	7	2	14	3	3	3	3	2	

Table E-26. Entiat River 2008.

Month		March				April				May				June				July				August			September					
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37		
3-Hydroxycarbofuran	D-C			0.014																										
Total Suspended Solids		3	2	2	1	2	7	3	6	16	5	24	13	8	5	7	4	9	5	3	3	3	3	2	3	2	1	2		

Table E-27. Entiat River 2009.

Month		March				April				May				June				July				August			Sep			
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
Chlorpyrifos	I-OP					0.023																						
Endosulfan I	I-OC					0.024																						
Piperonyl Butoxide	Sy			0.068		0.083	0.100																					
Total Suspended Solids				2		2	3	12	3	4	5	46	19	13	11	5	4	4	3	2	2	3	2	2	2	2	2	2

D = Degradate, H = Herbicide, I = Insecticide, Sy = Pesticide synergist, C = Carbamate, OC = Organochlorine, OP = Organophosphate

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Appendix F. Continuous Temperature Profiles

Temperature measurements were made at 30-minute intervals for the duration of the 2009 calendar year.

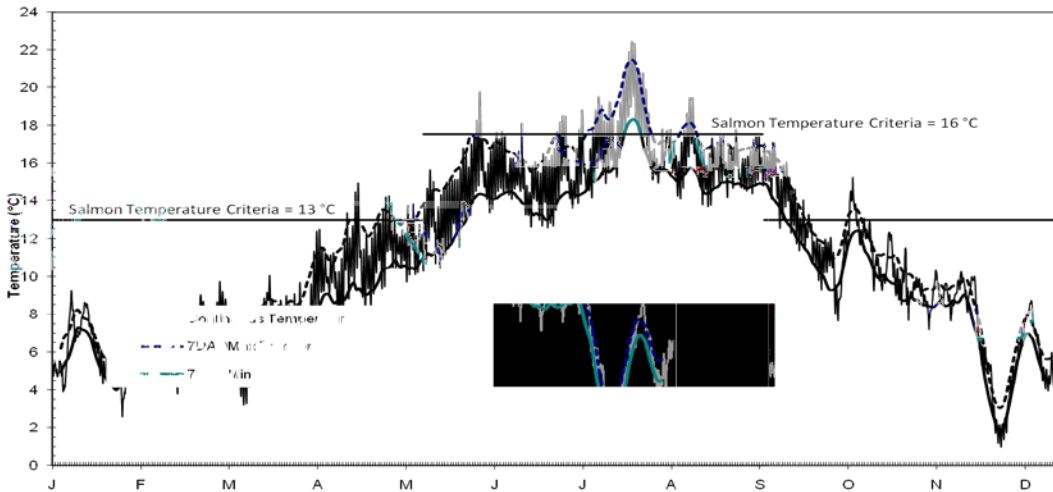


Figure F-1. 2009 continuous temperature profile for the mainstem of Thornton Creek.

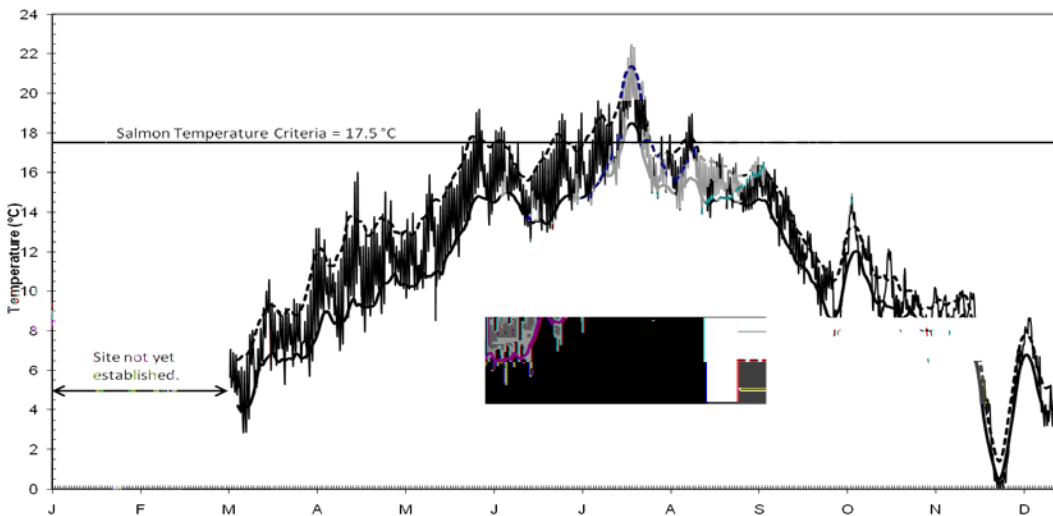


Figure F-2. 2009 continuous temperature profile for Longfellow Creek.

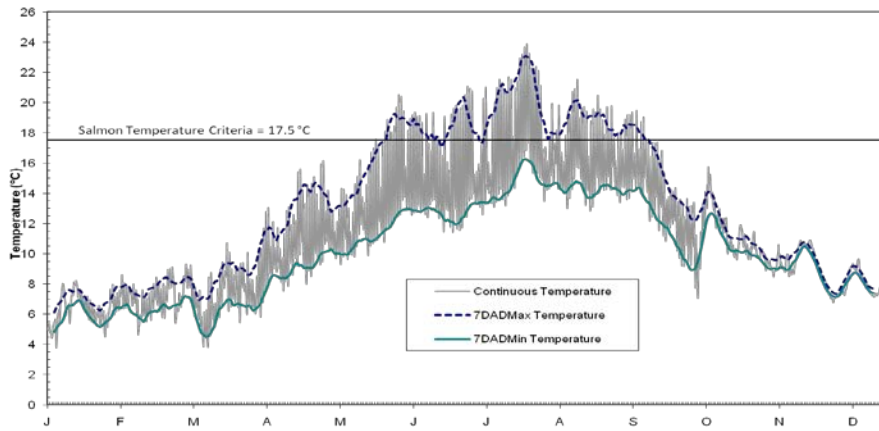


Figure F-3. 2009 continuous temperature profile for upper Big Ditch.

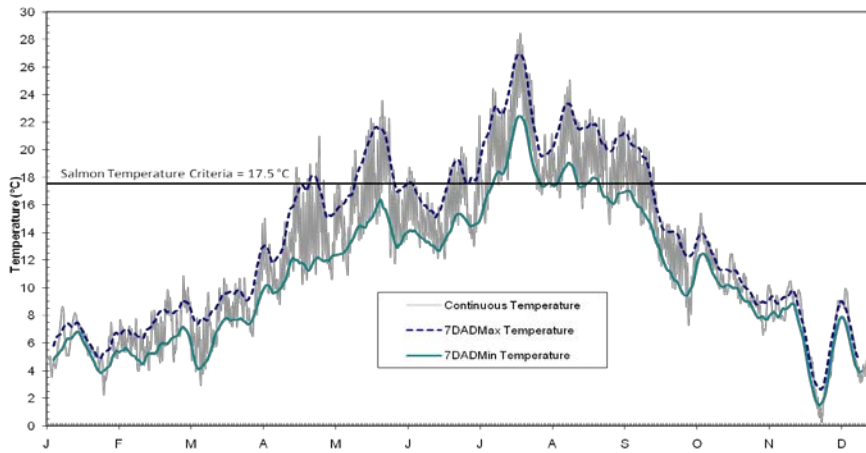


Figure F-4. 2009 continuous temperature profile for lower Big Ditch.

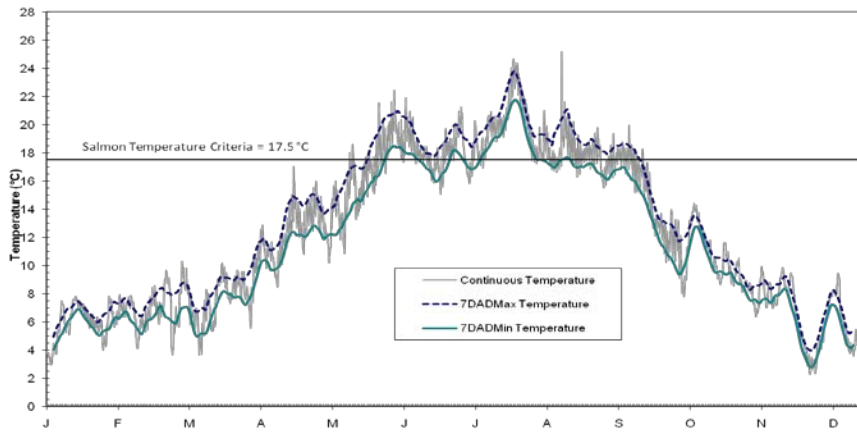


Figure F-5. 2009 continuous temperature profile for Indian Slough.

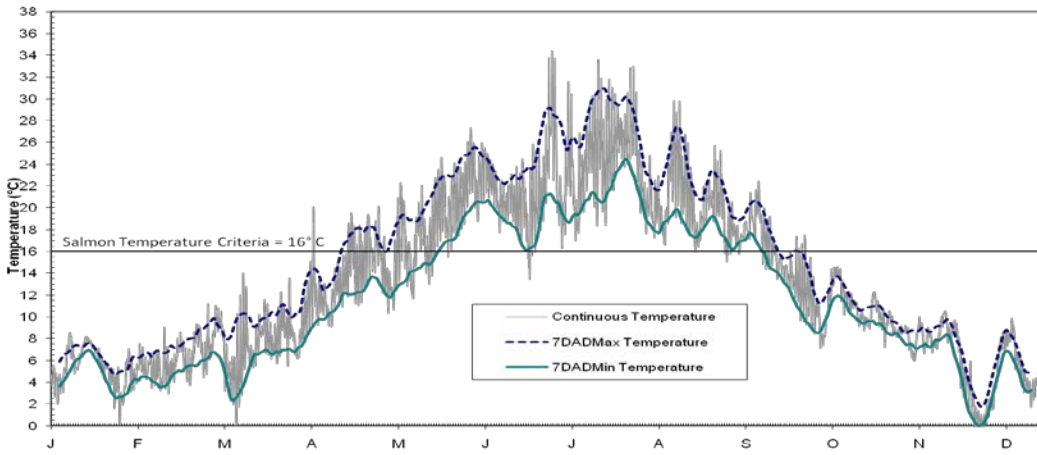


Figure F-6. 2009 continuous temperature profile for Brown Slough.

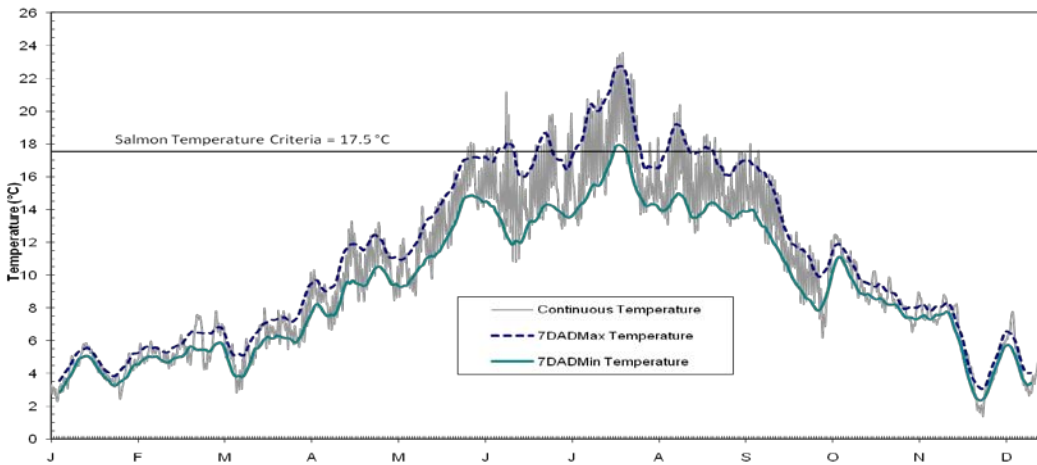


Figure F-7. 2009 continuous temperature profile for the Samish River.

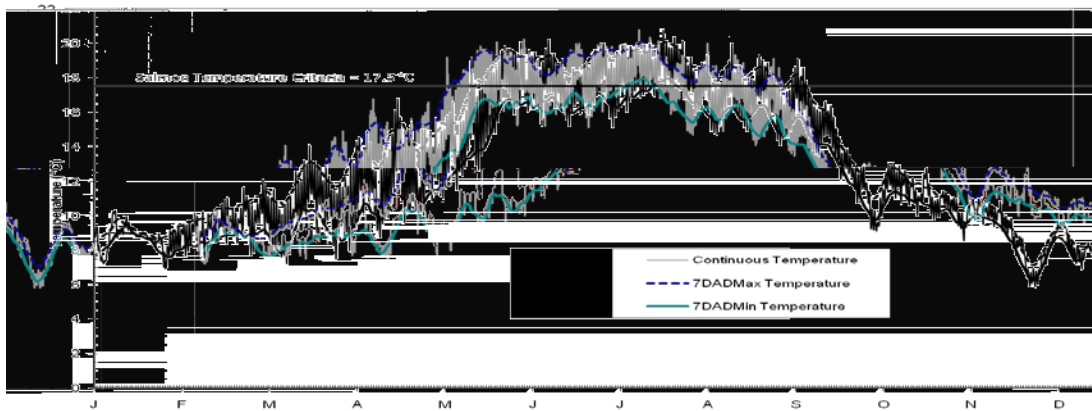


Figure F-8. 2009 continuous temperature profile for upper Spring Creek.

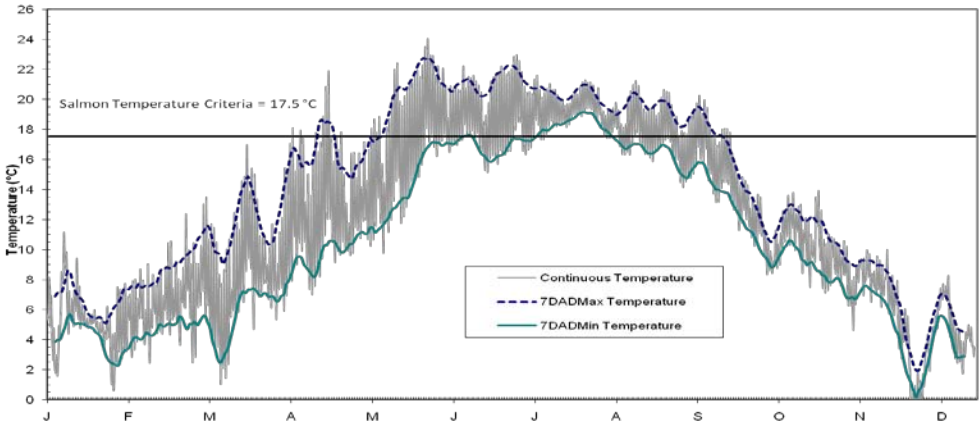


Figure F-9. 2009 continuous temperature profile for lower Spring Creek.

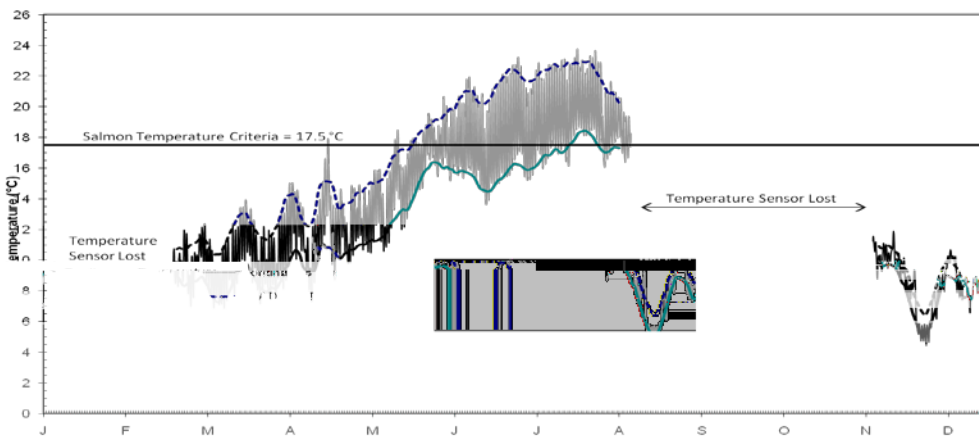


Figure F-10. 2009 continuous temperature profile for Marion Drain.

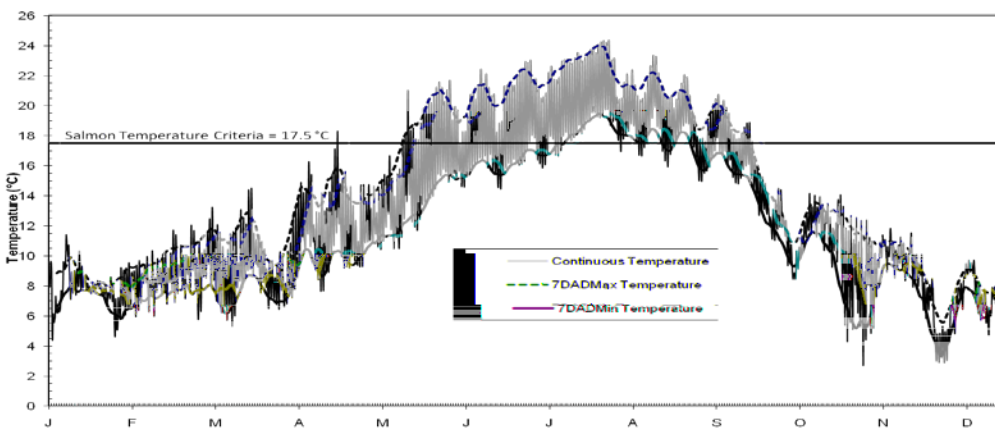


Figure F-11. 2009 continuous temperature profile for Sulphur Creek Wasteway.

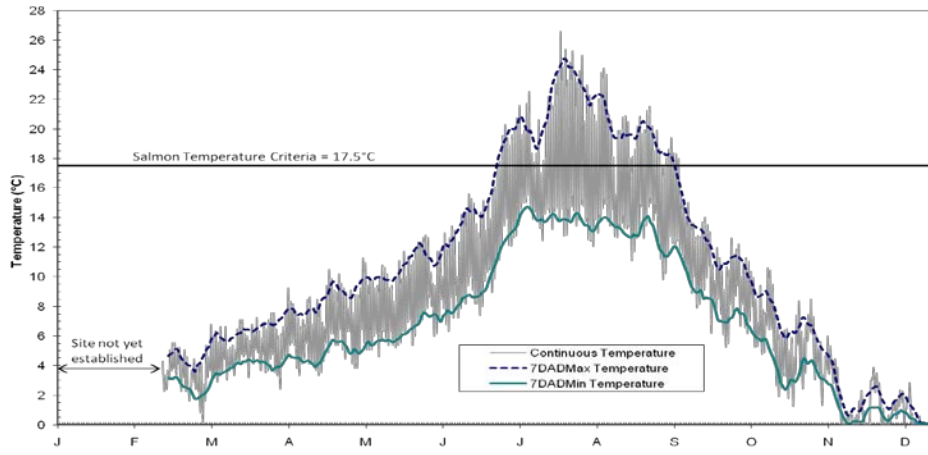


Figure F-12. 2007 continuous temperature profile for Peshastin Creek.

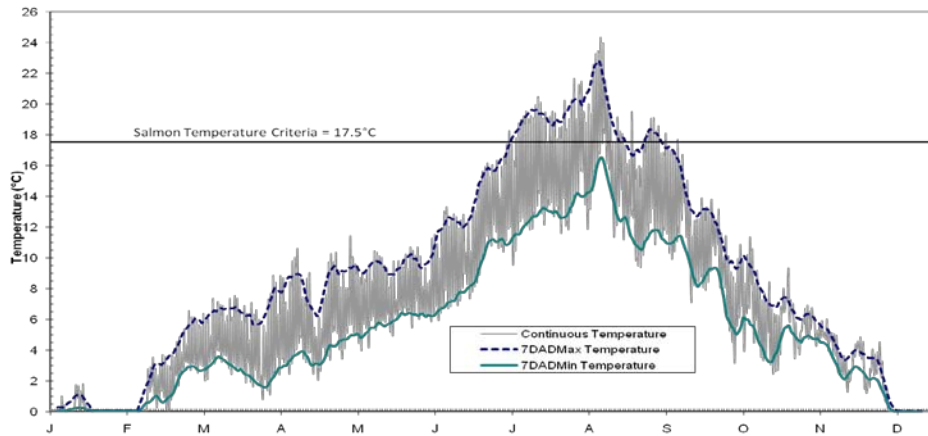


Figure F-13. 2008 continuous temperature profile for Peshastin Creek.

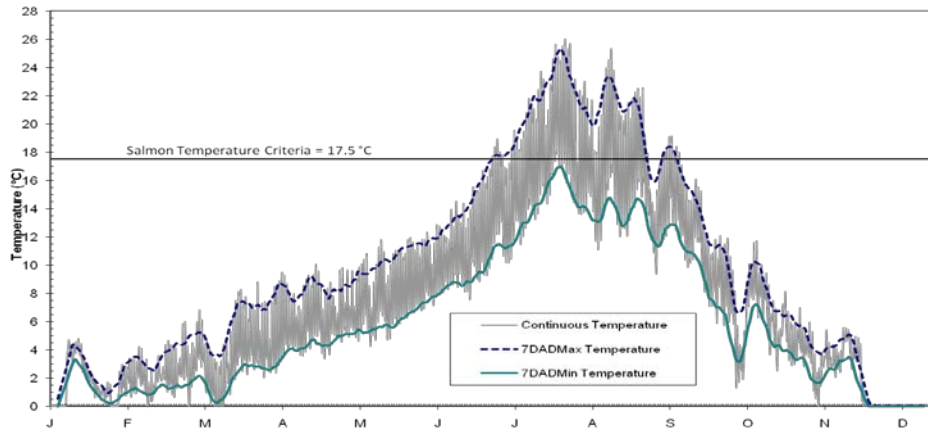


Figure F-14. 2009 continuous temperature profile for Peshastin Creek.

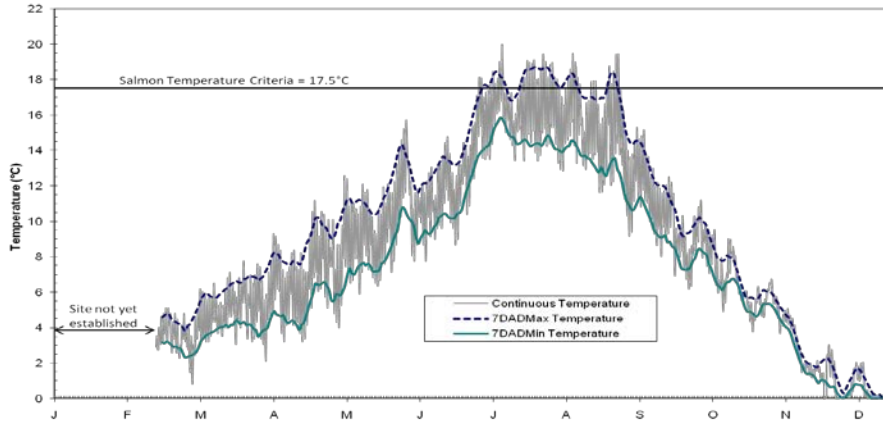


Figure F-15. 2007 continuous temperature profile for Mission Creek.

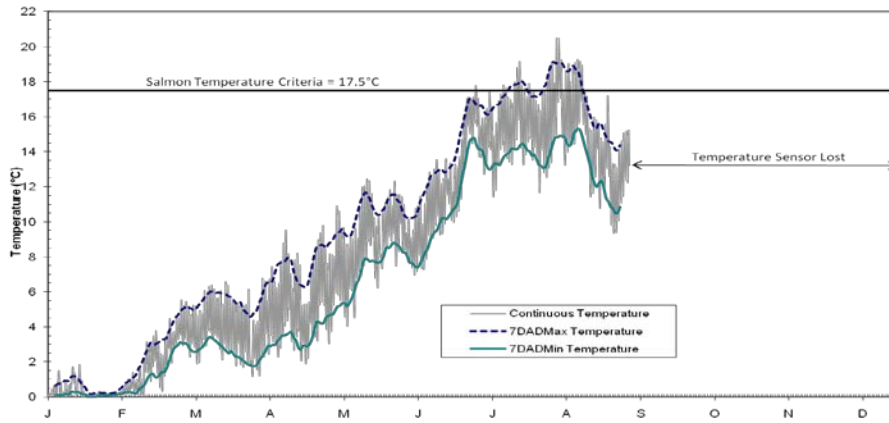


Figure F-16. 2008 continuous temperature profile for Mission Creek.

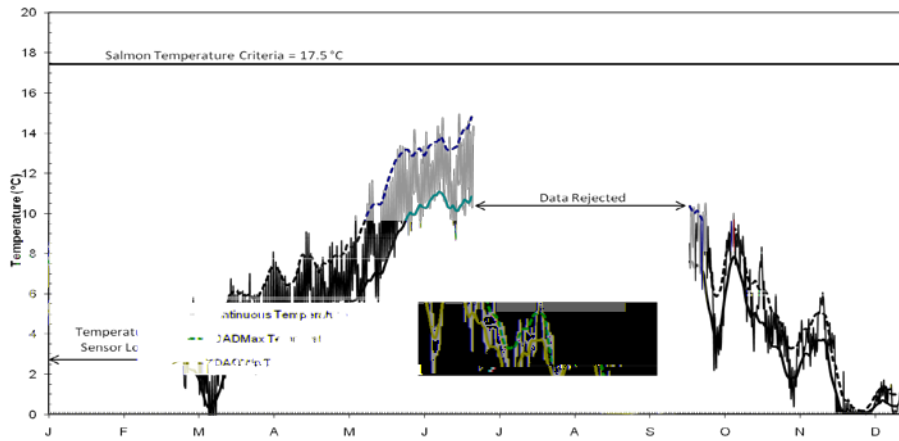


Figure F-17. 2009 continuous temperature profile for Mission Creek.

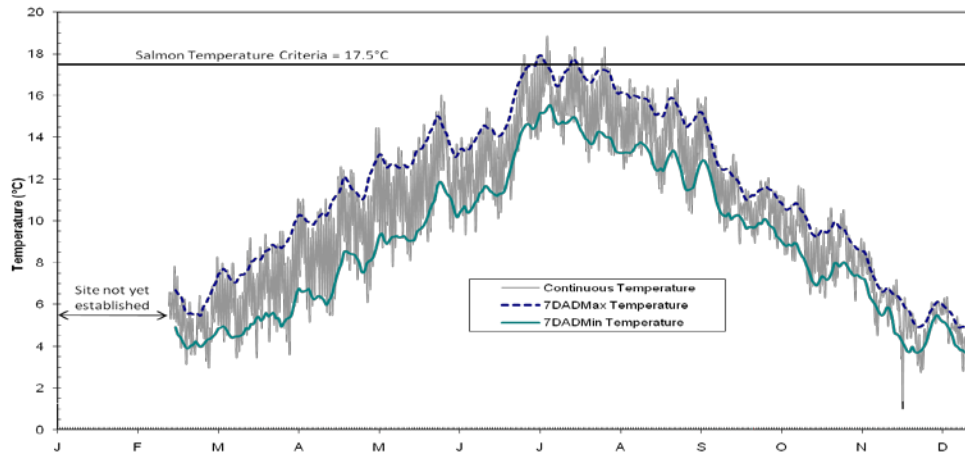


Figure F-18. 2007 continuous temperature profile for Brender Creek.

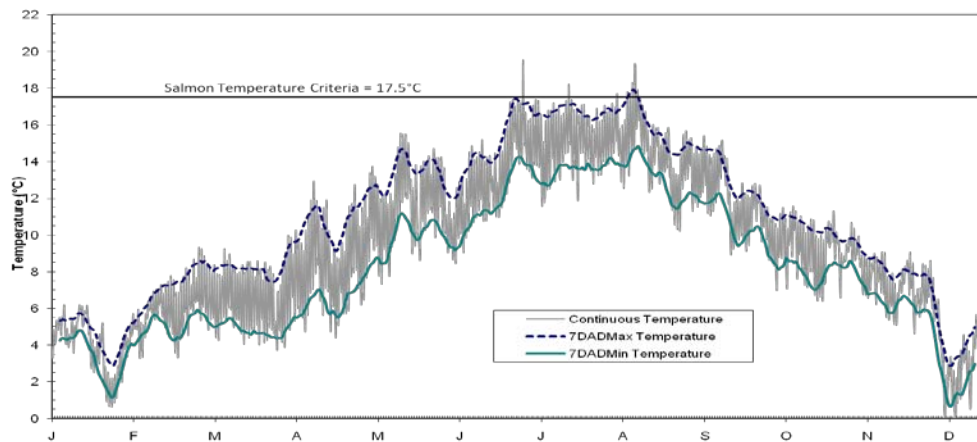


Figure F-19. 2008 continuous temperature profile for Brender Creek.

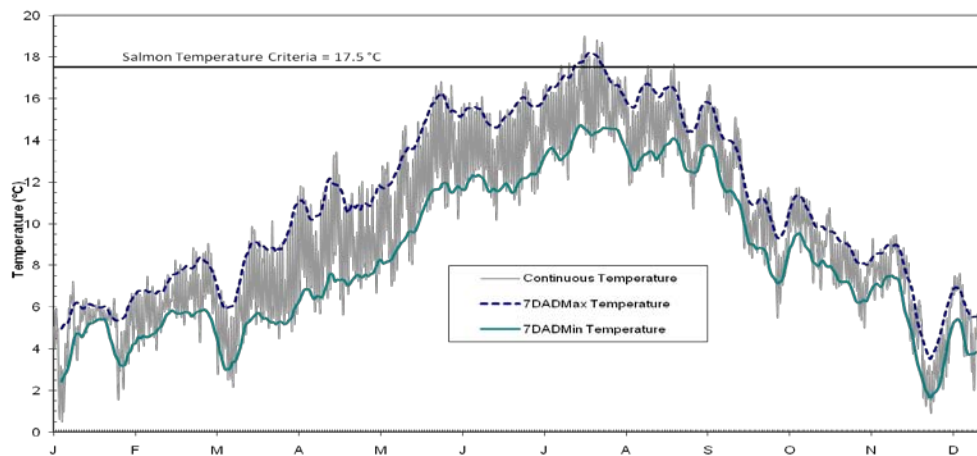


Figure F-20. 2009 continuous temperature profile for Brender Creek.

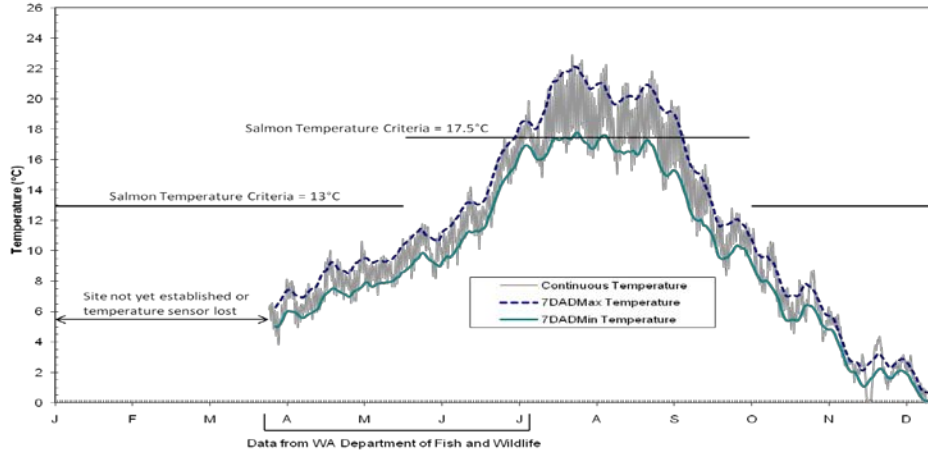


Figure F-21. 2007 continuous temperature profile for the Wenatchee River.

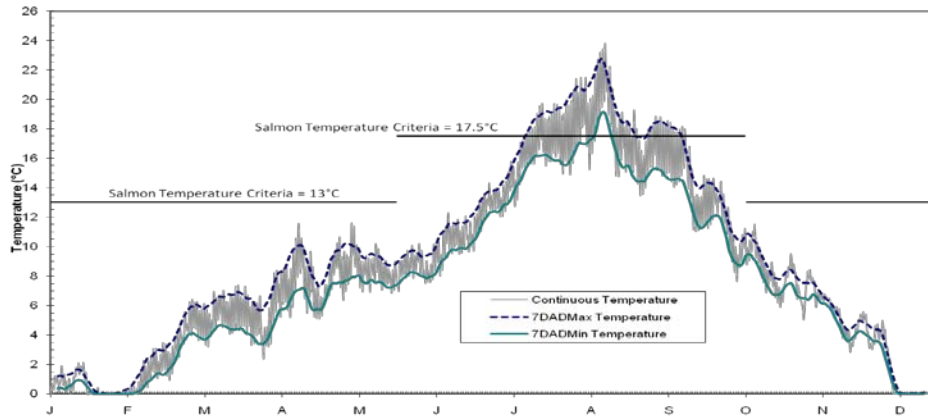


Figure F-22. 2008 continuous temperature profile for the Wenatchee River.

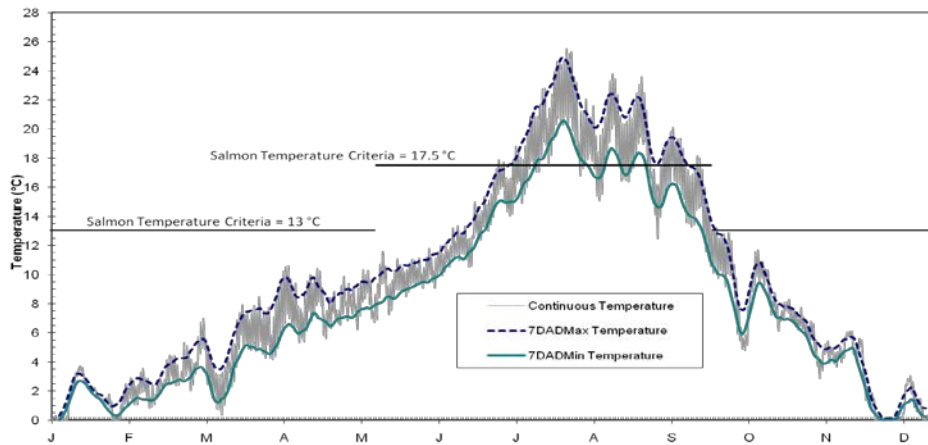


Figure F-23. 2009 continuous temperature profile for the Wenatchee River.

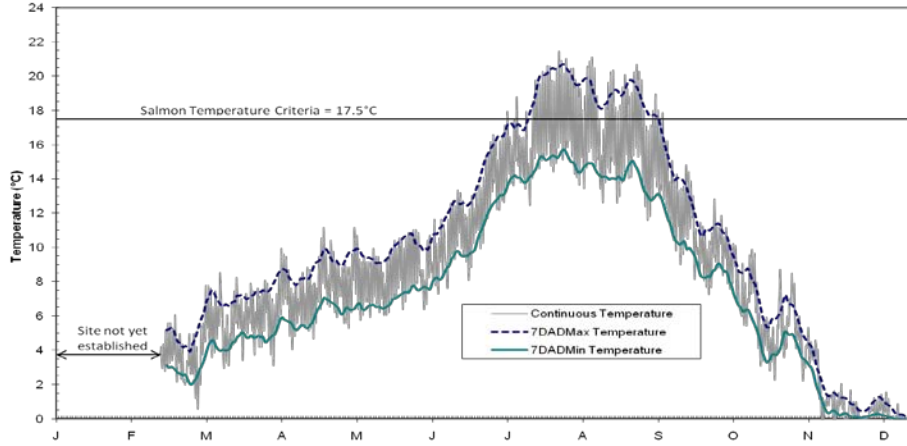


Figure F-24. 2007 continuous temperature profile for the Entiat River.

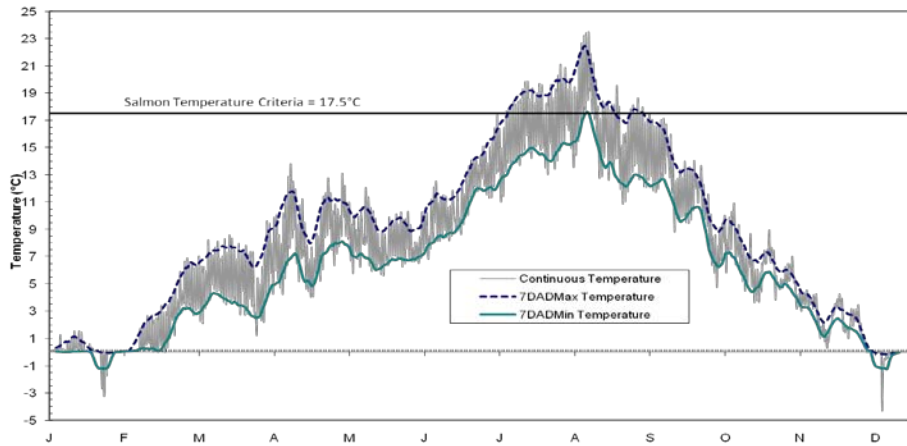


Figure F-25. 2008 continuous temperature profile for the Entiat River.

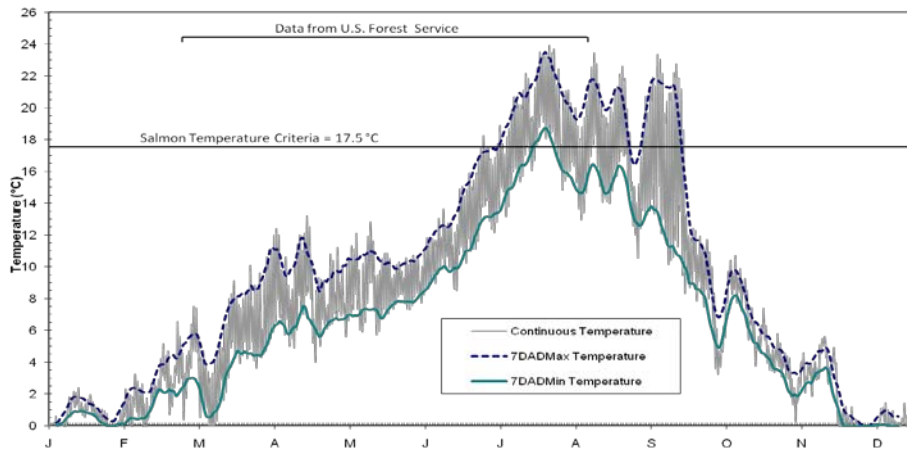


Figure F-26. 2009 continuous temperature profile for the Entiat River.

Appendix G. Flow, Precipitation, and Pesticide Detection Graphs

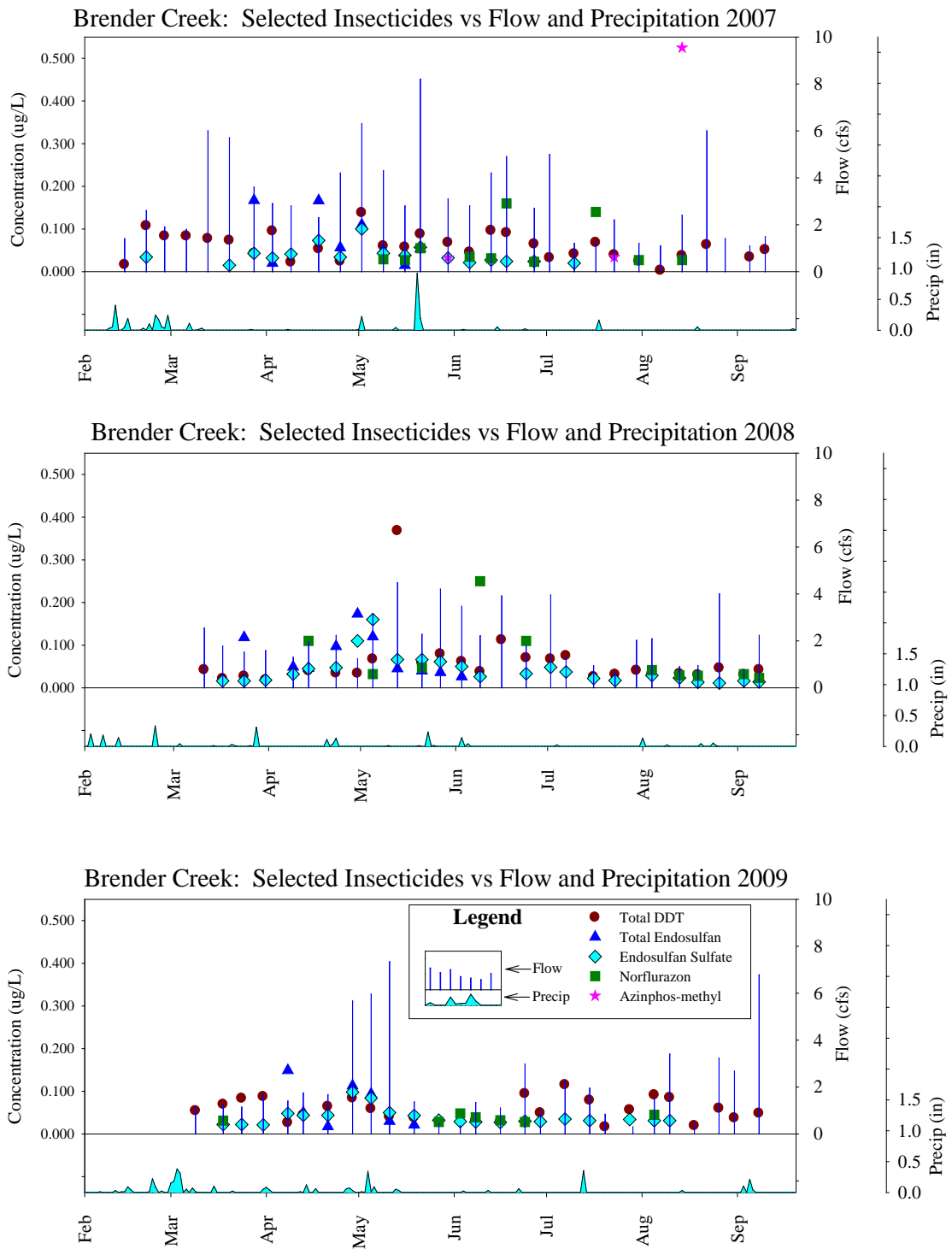


Figure G-1. Flow, precipitation, and most commonly seen insecticide concentrations for upstream Brender Creek, 2007-09.

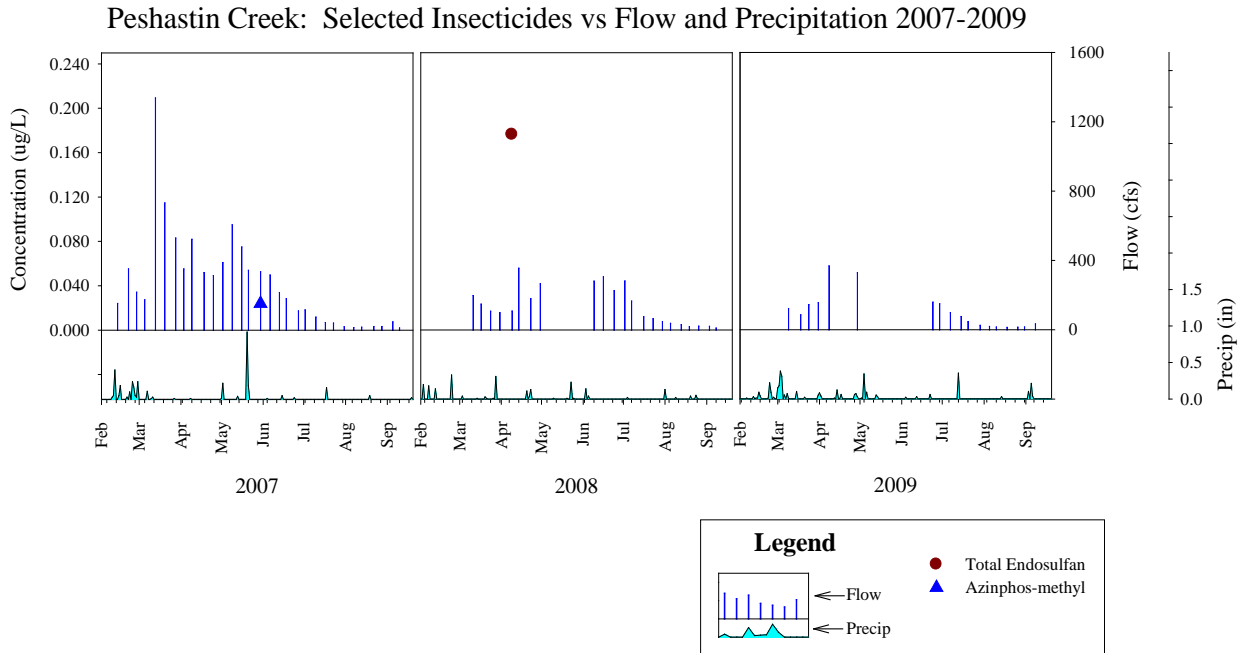


Figure G-2. Flow, precipitation, and most commonly seen insecticide concentrations for Peshastin Creek, 2007-09.

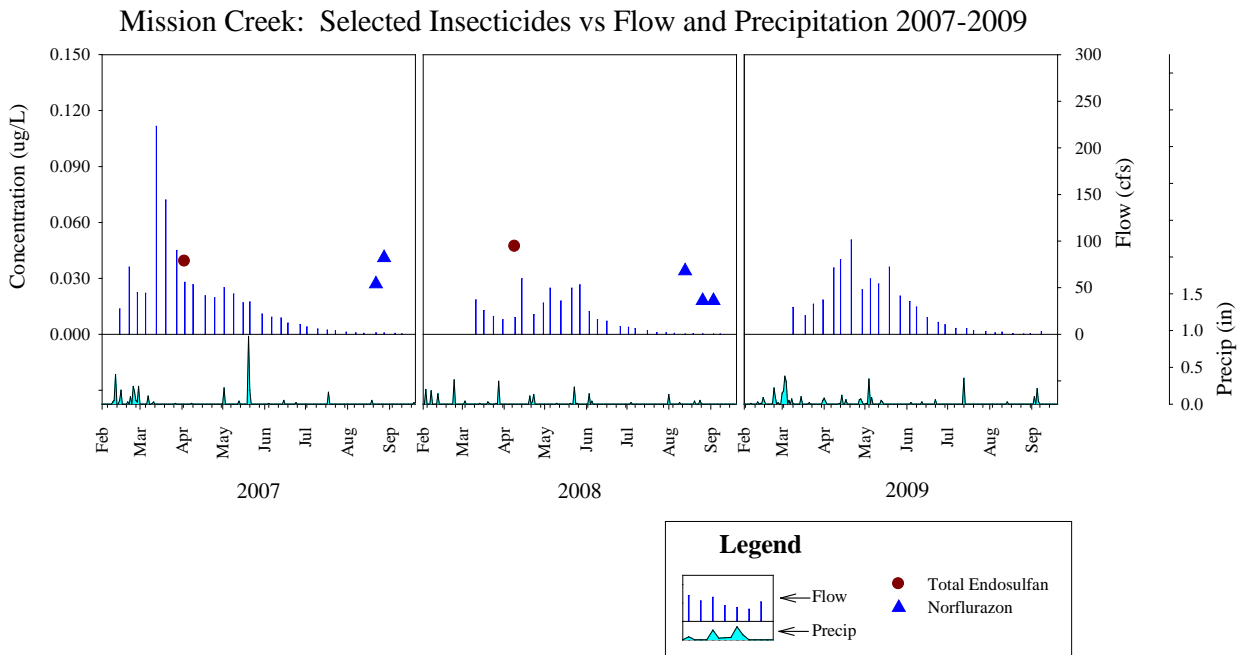


Figure G-3. Flow, precipitation, and most commonly seen insecticide concentrations for Mission Creek, 2007-09.

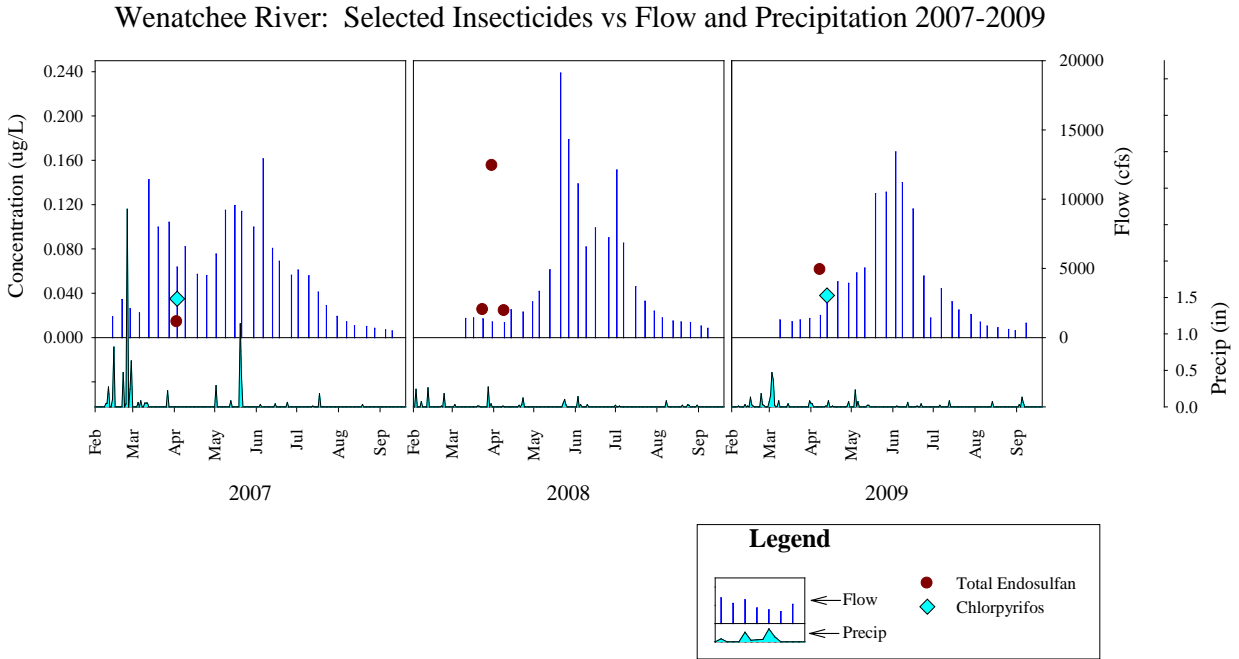


Figure G-4. Flow, precipitation, and most commonly seen insecticide concentrations for the Wenatchee River, 2007-09.

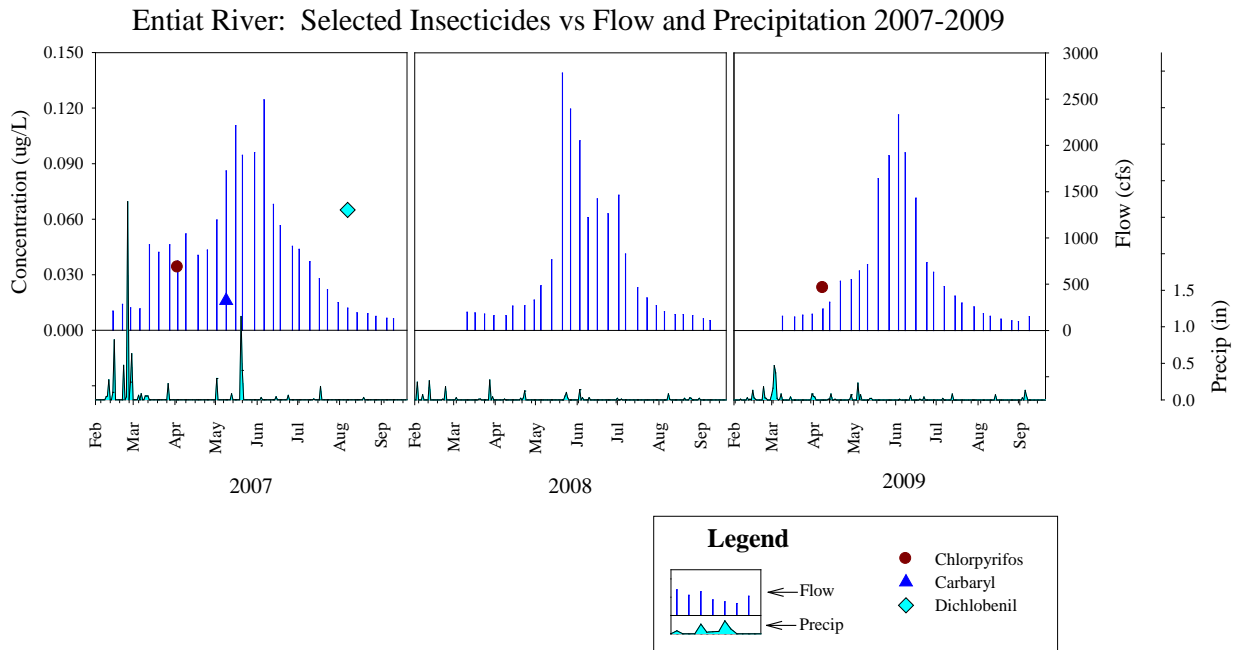


Figure G-5. Flow, precipitation, and most commonly seen herbicide concentrations for the Entiat River, 2007-09.