Surface Water Monitoring Program for Pesticides in Salmonid-Bearing Streams, 2010 Data Summary

A Cooperative Study by the Washington State Departments of Ecology and Agriculture

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Cover photo:

Left: Trellised apple orchard in the Brender Creek subbasin (photo by Dan Dugger).
Right: Dan Dugger, Department of Ecology employee, sampling for pesticides at Peshastin Creek (photo by Evan Newell).

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A Cooperative Study by the Washington State Departments of Ecology and Agriculture

by
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Table of Contents

Appendices C-E are in a separate electronic file on the web.

List of Figures and Tables

Page

Figures

Tables

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Abstract

Since 2003 the Washington State Departments of Agriculture and Ecology have been conducting a multi-year monitoring study to characterize pesticide concentrations in selected salmon-bearing streams during a typical pesticide-use period.

Monitoring is conducted in six basins:

- Thornton Creek in the Cedar-Sammamish basin and Longfellow Creek in the Green-Duwamish basin, representing urban land use.
- Lower Skagit-Samish basin, representing western Washington agriculture.
- Lower Yakima basin, representing eastern Washington irrigated agriculture.
- Wenatchee and Entiat basins, representing central Washington tree fruit agriculture.

This report summarizes data collected during the 2010 monitoring season. In 2010 surface water samples were analyzed for over 170 pesticides and pesticide degradates, as well as total suspended solids. Field measurements were collected for streamflow, temperature, pH, conductivity, and dissolved oxygen.

During 2010 only a few pesticide detections did not meet water quality criteria or standards:

- The urban sites and the Skagit-Samish sites met all available pesticide criteria.
- In the lower Yakima basins, one chlorpyrifos detection and one malathion did not meet a chronic pesticide criteria.
- In the Wenatchee-Entiat basins, the Wenatchee River and Mission Creek pesticide detections met all available criteria. One endosulfan detection in Peshastin Creek and in Brender Creek did not meet the endangered species level of concern for fish. The Entiat River had one detection of a legacy DDT degradate that did not meet chronic criteria. As in previous years, Brender Creek also had a number of detections of DDT and degradates that did not meet chronic criteria.

An intensive triennial review of pesticide results will be conducted after the 2011 monitoring season.

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Introduction

The Washington State Departments of Agriculture (WSDA) and Ecology (Ecology) are conducting a multi-year monitoring study to evaluate pesticide concentrations in surface waters. The study assesses pesticide presence in salmon-bearing streams during a typical pesticide-use season (e.g., March through October).

WSDA, the U.S. Environmental Protection Agency (EPA), the National Atmospheric and Oceanic Administration (NOAA) National Marine Fisheries Service, and the U.S. Fish and Wildlife Service (USFWS) use the data from this study to refine exposure assessments for pesticides that are registered for use in Washington State. Understanding the fate and transport of pesticides allows regulators to assess the potential effects of pesticides on endangered salmon species while minimizing the economic impacts to agriculture.

The purpose of this data report is to provide results from monitoring conducted during 2010 in six basins and to document any changes that occurred in the monitoring program during the year.

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Study Area

This pesticide monitoring project has been ongoing since 2003. As the project has progressed, additional sampling areas have been added.

Basins Monitored During 2010

The six basins monitored in 2010 are presented in Figure 1: two urban and four agricultural basins. The urban basins were chosen due to land-use characteristics, history of pesticide detections, and habitat use by salmon. The agricultural basins were chosen because they support several salmonid populations, produce a variety of agricultural commodities, and have a high percentage of cultivated areas.

Monitoring areas and timeframes are:

- Thornton Creek, located in the Cedar-Sammamish basin (WRIA[1] 8), represents an urban land-use area. Two to four sites have been sampled on this creek from 2003-2008. Starting in 2009 only one site, at the mouth of Thornton Creek, was sampled.
- Longfellow Creek, located in the Green-Duwamish basin (WRIA 9), represents an urban land-use area. Sampling started on this creek at one site in 2009.
- Four sub-basins of the lower Skagit-Samish basin (WRIA 3) were selected to represent western Washington agricultural land-use practices. The Samish River, Big Ditch Slough, Browns Slough, and Indian Slough have been sampled since 2006.
- Three sub-basins of the Lower Yakima basin (WRIA 37) were selected to represent eastern Washington irrigated crop-land agricultural practices. Marion Drain, Sulphur Creek Wasteway, and Spring Creek have been sampled since the start of the project in 2003.
- Four sub-basins of the Wenatchee basin (WRIA 45) and Entiat basin (WRIA 46) were selected to represent central Washington agricultural tree fruit practices. Peshastin Creek, Mission Creek, Brender Creek, and the Wenatchee River (WRIA 45) and the Entiat River (WRIA 46) have been sampled since 2007.

Site locations and duration of sampling during 2010 are described in Appendix B.

Detailed descriptions of sites, including basin description, site map, climate, agricultural land-use, and the salmon fishery, are included in the last triennial report (Sargeant et al., 2010) and the 2009 data report (Sargeant et al., 2011).

[1] Water Resource Inventory Area

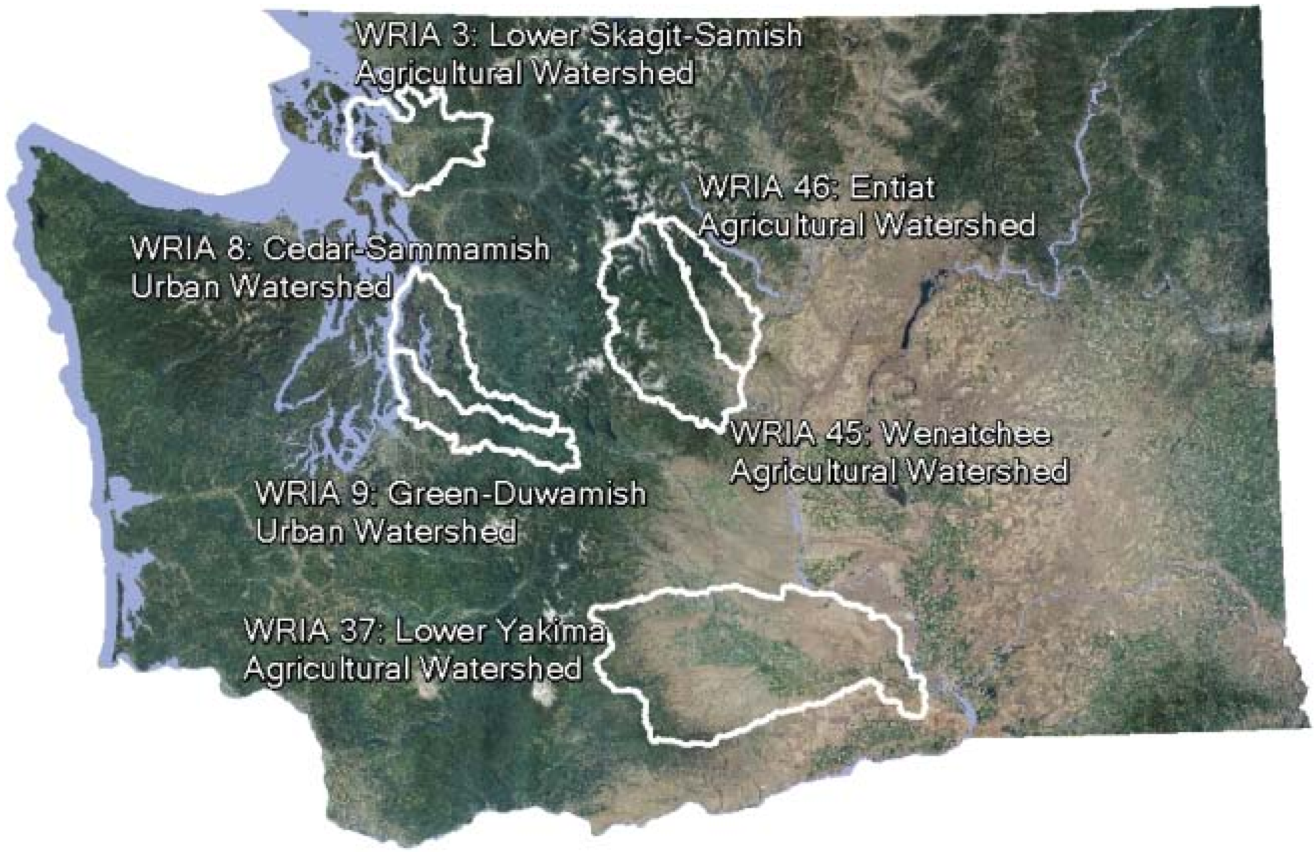

Figure 1. State map showing the six urban and agricultural basins monitored during 2010.

Study Design and Methods

Sampling was designed to address pesticide presence in salmonid-bearing streams during a typical pesticide-use period (e.g., March through September). The focus of monitoring is on currently registered pesticides, but laboratory analysis also included some historically used pesticides. Conventional water quality parameters were measured: total suspended solids (TSS), pH, conductivity, temperature, dissolved oxygen, and streamflow. The conventional parameters provide information to help better determine the factors influencing pesticide toxicity, fate and transport, and general water quality.

Detailed information on study design and methods are described in the Quality Assurance (QA) Project Plan (Johnson and Cowles, 2003), subsequent addendums (Burke and Anderson, 2006; Dugger et al., 2007; Anderson and Sargeant, 2009), and the triennial reports (Burke et al., 2006; Sargeant et al., 2010).

During 2010 samples were collected for analysis of over 170 pesticides and degradates including: 74 insecticides, 59 herbicides, 30 degradate pesticides, 10 fungicides, 2 synergistic compounds, and one wood preservative.

Sampling Sites and Sampling Frequency

In 2010, 27 sampling events were conducted. Sampling began the second week in March and continued through the second week in September at all sites except Marion Drain. As in previous years, Marion Drain sampling continued through the end of October (for a total of 34 sampling events) for organophosphate pesticides and TSS. The upstream Spring Creek site in the lower Yakima basin was sampled every other week for a total of 14 sampling events.

Field Procedures and Laboratory Analyses

A full description of field procedures and laboratory analysis is included in Sargeant et al. (2010). Field methods for grab sampling are a direct application or modification of United States Geological Survey (USGS) or EPA procedures. Surface water samples were collected by hand-compositing grab samples from quarter-point transects across each stream. In situations where streamflow was vertically integrated, a one-liter transfer container was used to dip and pour water from the stream into sample containers. Otherwise samples were collected using depth integrating equipment. Sample/transfer containers were delivered pre-cleaned by the manufacturer to EPA specifications (EPA, 1990). After collection, all samples were labeled and preserved according to the QA Project Plan (Johnson and Cowles, 2003).

Field meters were calibrated at the beginning of the field day according to manufacturers' specifications, using Ecology standard operating procedures (SOPs) (Swanson, 2007). Meters were post-checked at the end of the field day using known standards. Conventional parameters measured in the field were replicated once per sample day. Dissolved oxygen meter results were compared to grab samples that were analyzed by Winkler Titration for dissolved oxygen following Ecology SOPs (Ward, 2007). Two to three Winkler grab samples were obtained during each sample day. Continuous, 30-minute interval, temperature data were collected year-round in 2010. Temperature instruments were calibrated against a National Institute of Standards and Technology (NIST) primary reference (Wagner et al., 2000). Data quality objectives for field meters are described in Anderson and Sargeant (2009).

Discharge for sites other than Sulphur Creek Wasteway, Wenatchee River, Peshastin Creek, and Entiat River were measured using a Marsh-McBirney flow meter and top-setting wading rod, as described in the USGS method (Rantz et al., 1983). Discharge data for Sulphur Creek Wasteway were obtained from an adjacent U.S. Bureau of Reclamation gaging station, "SUCW – Sulphur Creek Wasteway at Holaday Road near Sunnyside". Wenatchee and Entiat River discharge data were obtained from USGS at the Wenatchee River at Monitor (Station 12462500) and Entiat River near Entiat (Station 12452990). Discharge data for Peshastin Creek were obtained from an Ecology gaging station located at Green Bridge Road (Station 45F070). Fifteen-minute discharges were available during the sampling period. The recorded flow closest to the actual sampling time was used in lieu of field measurements.

Ecology's Manchester Environmental Laboratory (MEL) analyzed all pesticide and TSS samples. Laboratory methods are presented in Table 1. A list of target analytes for this study is presented in Appendix C, Table C-3.

Table 1. Summary of laboratory methods, 2010.

Analyte	Analytical Methods[1]		
	Extraction	Analysis	Reference
Pesticides[2]	3510	GC/MS	8270
Herbicides	8151	GC/MS	8270 and 8251
Carbamates	3535M	LCMS/MS	8321A
Total Suspended Solids	n/a	Gravimetric	EPA 160.2

[1] All analytical methods refer to EPA SW 846, unless otherwise noted.
[2] Pesticides refers to all forms tested unless indicated otherwise.
GC: gas chromatograph.
MS: mass spectrometry.
LC: Liquid chromatography.
n/a: not applicable.

In 2010 MEL purchased a new instrument for carbamate analysis. With the new instrument, all analytes have at least one confirmation ion in addition to the quant ion. In addition, the new instrument improved sensitivity for all analytes. These improvements significantly reduced identification uncertainty, thus decreasing the potential for false positive and negatives.

Laboratory methods are discussed in the QA Project Plan (Anderson and Sargeant, 2009); previous QA Project Plan (Johnson and Cowles, 2003) and the QA Project Plan addendum (Burke and Anderson, 2006); and the SOP for the *Pesticides in Salmonid Streams Project* (Anderson and Sargeant, 2010).

Data Quality

Laboratory Data Quality

Performance of laboratory analyses is governed by QA and quality control (QC) protocols. The QA/QC protocol employs application of blanks, replicates, surrogates, and laboratory control samples, as well as matrix spike/matrix spike duplicates (MS/MSDs). Laboratory surrogate, blank, replicate, and control samples are analyzed as the laboratory component of QA/QC. Field blanks, replicates, and MS/MSDs integrate field and laboratory components. A summary of laboratory and field data quality are presented below. For a detailed discussion of 2010 data quality, refer to Appendix C.

At a minimum, during each week there was at least one replicate, one blank, and one MS/MSD covering at least one of the four laboratory analyses (PESTMS, HERBS, CARBAMLL, and TSS). QA/QC samples were concentrated during April, May, and June to cover the intensive application period for most pesticides. Sites were randomly selected for application of QA/QC samples.

From 2006-2009 there was an anomaly in the carbamate pesticide analytical method that caused false positive identification of 1-naphthol, aldicarb sulfone, aldicarb sulfoxide, and oxamyl (2009 only). In 2010 MEL purchased a new instrument that greatly improved the identification of all analytes included in the carbamate analysis. In addition sensitivity also improved. The improvements combined to significantly reduce identification uncertainty, thus decreasing the potential for false positives.

Because of the increased sensitivity of the new instrument, detections of select analytes in the carbamate analysis suite increased. One notable increase in detections was for the neonicotinoid insecticide, imidacloprid. In 2009 there were 15 detections of imidacloprid, and in 2010 there were 114 detections of imidacloprid; detections occurred in all of the project sampling areas.

Laboratory Blanks

Laboratory blank detections for 2010 are presented in Table 2. For all lab blank detections, any analyte found in associated samples below 5 times the lab blank detection were reported at the level detected but qualified as not detected at an estimated detection limit (UJ).

Table 2. Laboratory blank detections, 2010 (µg/L).

Analysis	Chemical	Analysis Date	Value
GCMS	2,4'-DDT	6/11/2010	0.015 J
	4,4'-DDD		0.012 J
	4,4'-DDE		0.007 J
	4,4'-DDT		0.018 J
	cis-Chlordane		0.002 J
	Mirex		0.012 J
	Trans-Chlordane		0.002 J
LCMS\MS	Imidacloprid	4/14/2010	0.001 J
		9/28/2010	0.002 J
	Carbaryl	6/11/2010	0.003 J
		7/23/2010	0.004 J

Field Blanks

Field blank detections indicate the potential for sample contamination in the field and laboratory and the potential for false detections due to analytical error.

In 2010 there were no field blank detections for the pesticide analysis. There was one TSS field blank detection of 3 mg/L on July 20, 2010 for the Samish River site. The reporting limit for TSS was 1 mg/L. All TSS values collected on that day (July 20, 2010) that are less than 9 mg/L will be qualified as estimates.

Replicate Results

Replicate sampling tests the reproducibility or precision of sampling results. During 2010 field replicate sampling frequency for pesticides was 7.6%, and for TSS was 7.7%. Precision between replicate pairs was calculated using relative percent difference (RPD).

Excluding TSS, 77 analytes were consistently identified, and 19 analytes were inconsistently identified in 97 replicate pairs. The average RPD of consistent field replicate pairs was low, 8.4%. This is an improvement over previous years and is likely due to improvements in the carbamate analysis instrumentation.

TSS was consistently detected in 33 replicate pairs. The average RPD of all replicates was 12.4%. A total of 81% of the replicate pairs were within the RPD criterion (20%).

Surrogates, Matrix Spikes, and Laboratory Control Samples

Surrogates are used to evaluate recovery for a group of compounds. The majority of surrogate recoveries fell within the control limits established by MEL. Sample results were qualified as estimates when surrogate recoveries did not meet MEL QC criteria.

MS/MSDs provide an indication of bias due to interferences from components of the sample matrix. The duplicate spike can be used to estimate analytical precision at the concentration of the spiked samples. The average recovery of the MS/MSD was 97%, and the average RPD between MS/MSD pairs was 7.8%. For most compounds, recovery and RPDs of MS/MSD pairs showed acceptable performance and were within defined limits for the project. Sample results were qualified as estimates if the MS/MSD recoveries did not meet MEL QC criteria.

Laboratory control samples (LCS) are analyte compounds spiked into deionized water at known concentrations and subjected to analysis. They are used to evaluate accuracy of pesticide residue recovery for a specific analyte. The average percent recovery for the LCS and the LCS duplicates was 92%, and the average RPD between the LCS and duplicate pairs was 12%. For most compounds, recovery and RPDs of LCS and LCS duplicates showed acceptable performance and were within limits for the project. Sample results were qualified as estimates if the LCS recoveries did not meet MEL QC criteria.

Field Data Quality

A detailed discussion of 2010 field data quality is included in Appendix C. In 2010 the field meter for the lower Yakima and Wenatchee-Entiat sites (eastside sites) met QC objectives including post-checks and Winkler comparisons for most sample events. On July 7, August 9 and 25, and October 20, conductivity measurements for the eastside sites were qualified as estimates due to meter post-checks not meeting QC limits.

At Indian Slough, a westside site, two replicate measures for dissolved oxygen and one for conductivity did not meet QC objectives. This site is influenced by incoming marine water; temperature, dissolved oxygen, and conductivity values vary by depth. Differences in the replicates were likely due to environmental factors and not due to data quality issues. Indian Slough dissolved oxygen and conductivity results for these days were qualified as estimates.

Two field audits were conducted in 2010. The purpose of the field audit is to ensure that sampling methodologies are consistent. Details of the audits are presented in Appendix C. The findings of the field audits include that both Ecology sampling teams are conducting field operations using consistent sampling methodologies that results in comparable data.

Data Analysis and Reporting Methods

The 2010 field and laboratory data were compiled and organized using Excel® spreadsheet software and Access® database software (Microsoft Corporation, 2007). Water quality results from field and laboratory work were also entered into Ecology's Environmental Information Management (EIM) database (www.ecy.wa.gov/eim).

Graphs, plots, mass balance calculations, and some statistical analyses were made using Excel® software. The following guidelines were used in reporting and analyzing data for this report.

Laboratory data were qualified as needed, and qualifiers are described in Table 3. A positive pesticide detection included unqualified values and values qualified with a J or E. Values qualified with NJ, U, or UJ were considered non-detects.

Table 3. Definitions of data qualifiers.

Qualifier	Definition
No qualifier	The analyte was detected at the reported concentration. Data are not qualified.
E	Reported result is an estimate because it exceeds the calibration range.
J	The analyte was positively identified; the associated numeric value is the approximate concentration of the analyte in the sample.
NJ	The analysis indicates the presence of an analyte that has been “tentatively identified,” and the associated numeric value represents its approximate concentration.
NAF	Not analyzed for.
NC	Not calculated.
REJ	The sample results are rejected due to serious deficiencies in the ability to analyze the sample and meet QC criteria. The presence or absence of the analyte cannot be verified.
U	The analyte was not detected at or above the reported sample quantitation limit.
UJ	The analyte was not detected at or above the reported sample quantitation limit. However, the reported quantitation limit is approximate and may or may not represent the actual limit of quantitation necessary to accurately measure the analyte in the sample.

MEL, 2000, 2008; EPA, 1999, 2007.

Comparison to Assessment Criteria and Water Quality Standards

Non-detect values (U, UJ, N, NJ) were not used for comparison to *assessment criteria* or *water quality standards*. When summing compound totals (such as total DDT, total endosulfan), the Toxic Studies Unit Guidance was used (Ecology, 2008). Non-detects (U, UJ) were assigned a value of zero (as in the guidance). Unlike the guidance, NJ values (tentatively identified compounds) were also assigned a value of zero.

Replicate Values

Field and laboratory replicates were obtained to determine data quality. Field and laboratory replicates were arithmetically averaged for comparisons to *assessment criteria* and *water quality standards*. For data analysis purposes, field and laboratory replicates were arithmetically averaged. If the sample or the replicate was a non-detect value while the other (either sample or replicate) was a detection, then the detected value was used.

When a laboratory replicate was performed on a field replicate, the laboratory replicate mean was calculated before the field replicate mean.

For select statistical analysis, NJ qualified data were used when detected pesticide values were not available. When this occurred, it is specified in the statistical test description.

EPA Assessment Criteria and Washington State Water Quality Standards

Assessment of pesticide effects on endangered salmonid species is evaluated by comparing detected pesticide concentrations against three criteria:

- EPA Federal Insecticide Fungicide and Rodenticide Act (FIFRA) Pesticide Registration Toxicity Criteria.
- EPA National Recommended Water Quality Criteria (NRWQC).
- Washington State Water Quality Standards for the Protection of Aquatic Life (WAC 173-201A).

The EPA and Washington State aquatic life criteria are based on evaluating the effects of a single chemical on a wide array of families, including at least one fish species from the family *Salmonidae* and a second species in the class *Osteichthys*. The criteria and standards do not account for the effects of multiple chemicals or pesticide mixtures on an organism. Many of the pesticides included in this study do not have EPA or Washington State criteria.

Aquatic life criteria, pesticide regulatory criteria, and toxicity (acute and chronic) results for fish, invertebrates, and aquatic plants are presented in Appendix D. Measured concentrations higher than criteria concentrations do not necessarily indicate that the water quality criteria have not been met. Numeric water quality criteria contain concentration values and duration of exposure components; both must be compared to the measured concentrations to assess compliance with the criteria.

In this report, EPA FIFRA Pesticide Registration Toxicity Criteria and EPA NRWQC will be referred to as *assessment criteria*. Washington State numeric water quality standards for pesticides will be referred to as *water quality standards*. For a description of these criteria and standards for pesticides, refer to Appendix D.

EPA Pesticide Registration Toxicity Criteria

EPA uses risk quotients (RQ) to assess the potential risk of a pesticide to non-target organisms. A RQ is calculated by dividing the environmental concentration by either an acute or chronic toxicity value, which gives an evaluation of exposure over toxicity. The resulting RQ is a unitless value that is compared to Levels of Concern (LOC). The LOC set by EPA are presented in Table 4. These LOC are used to assess the potential risk of a pesticide to non-target organisms.

The endangered species LOC (0.05 for aquatic species) is used as a comparative value to assess potential risk to threatened or endangered salmonids. The endangered species RQ can also be expressed as $1/20^{th}$ of the acute Lethal Concentration 50 (LC_{50}) for aquatic organisms. To assess the potential risk of a pesticide to salmonids, the LC_{50} for rainbow trout is commonly used as a surrogate species. Thus the endangered species LOC presented in subsequent tables are $1/20^{th}$ of the rainbow trout LC_{50}. When available, the endangered species LOC for specific salmonids is also presented.

Table 4. Risk quotient criteria for direct and indirect effects of pesticides on aquatic organisms.

Test Data	Risk Quotient	Presumption
Acute LC_{50}	>0.5	Potentially high acute risk.
	>0.1	Risk that may be mitigated through restricted use classification.
	>0.05	Endangered species may be affected acutely, including sublethal effects.
Chronic NOEC	>1	Chronic risk; endangered species may be affected chronically, including reproduction and effects on progeny.
Acute invertebrate LC_{50}	>0.5	May be indirect effects on T&E fish through food supply reduction.
Aquatic plant acute LC_{50}	>1	May be indirect effects on aquatic vegetative cover for T&E fish.

Turner, 2003.
NOEC: No observable effect concentration.
T&E: Threatened and endangered.

Acute toxicity is calculated by standardized toxicity tests using lethality as the measured criteria. A properly conducted test will use a sensitive (representative) species at a susceptible life stage (usually young, though not immature). The test also will subject the test species to a pesticide under a range of concentrations (minimum: no effect, 50% and 100% mortality). The dose response curve may be calculated, and the LC_{50} lethal concentration to cause mortality in 50% of test species will be derived. For fish, the lethality test is conducted over 96 hours at a constant concentration. Acute invertebrate toxicity is normally calculated over 48 hours, with the criteria being mortality or immobility (LC_{50}, or Effective Concentration - EC_{50} for immobility). Acute toxicity testing for aquatic plants is conducted over 96 hours; the criterion is based on reduction in aquatic plant growth (EC_{50}).

Chronic fish tests normally use reproductive effects, or effects to offspring, as the measured effect. The dose response curve is evaluated to determine a no observable effect concentration (NOEC). The chronic toxicity test is longer than the 96-hour acute test (21 day for fish, 14 days for invertebrates, 5 to 60 days for plants) to simulate exposure resulting from a persistent chemical or effect of repeated applications.

Toxicity values such as those used for pesticide registration are determined from continuous exposure over time (e.g., LC_{50} freshwater fish acute toxicity tests are run for 96 hours at a constant concentration). When comparing the monitoring data to either the aquatic life criteria or directly to the toxicity criteria, one must consider the duration of exposure as well as the numeric toxicity value. For pesticide registration criteria, it is not possible to determine if an aquatic life criterion has been met based solely on an individual sample because the sampling frequency is usually weekly. Weekly sampling does not allow for assessment of the temporal component of the criteria.

EPA National Recommended Water Quality Criteria

The NRWQC are established by the EPA Office of Water for the protection of aquatic life, as established under the federal Clean Water Act (33 U.S.C. 1251 et. seq.). The pesticide criteria established under the Act are based on vertebrate and invertebrate acute and chronic toxicological data. States often adopt the NRWQC as their promulgated (legal) standards. The NRWQC was updated in 2006, and those criteria are used in this report (EPA, 2006).

Washington State Water Quality Standards

Pesticides

Washington State water quality standards are established in the Washington Administrative Code (WAC), Chapter 173-201A. These standards include numeric pesticide criteria for the protection of aquatic life.

The aquatic life criteria are designed to protect for both short-term (acute) and long-term (chronic) effects of chemical exposure. The criteria are primarily intended to avoid direct lethality, and growth and reproductive effects, to fish and other aquatic life within the specified exposure periods. The chronic criteria for a number of the chlorinated pesticides are based on protection of fish-eating wildlife from adverse effects due to bioaccumulation.

The exposure periods assigned to the acute criteria are expressed as: (1) an instantaneous concentration not to be exceeded at any time or (2) a one-hour average concentration not to be exceeded more than once every three years on average. The exposure periods for the chronic criteria are either: (1) a 24-hour average not to be exceeded at any time or (2) a four-day average concentration not to be exceeded more than once every three years on average. For Clean Water Act section 303(d) listing purposes, measurements of instantaneous concentrations are assumed to represent the averaging periods specified in the water quality standards for both acute and chronic criteria, unless additional measurements are available to calculate averages (Ecology, 2006).

Because few water quality criteria for pesticides have been developed, the majority of comparisons to measured pesticide concentrations contained in this report are made using pesticide registration toxicity criteria.

Aquatic life criteria, pesticide regulatory criteria, and toxicity (acute and chronic) results for fish, invertebrates, and aquatic plants are presented in Appendix D.

Water Quality Standards for Temperature, Dissolved Oxygen, and pH

Washington State water quality standards for conventional water quality parameters are set forth in Chapter 173-201A of the WAC. Waterbodies are required to meet numeric water quality standards based on the beneficial uses of the waterbody. Conventional parameters including temperature, dissolved oxygen, and pH were measured in this study. Sargeant et al. (2010)

provides a full description of the water quality standards and also explains why parameters such as temperature, dissolved oxygen, and pH are important for fish health.

Numeric Water Quality Standards

Thornton Creek subbasin

Beneficial uses for Thornton Creek are *Core Summer Salmonid Habitat* and *Extraordinary Primary Contact Recreation*. The numeric water quality standards for temperature, dissolved oxygen, and pH in Thornton Creek are described in Table 5.

Table 5. Freshwater water quality standards for temperature, dissolved oxygen, and pH for *Core Summer Salmonid Habitat* use and *Extraordinary Primary Contact Recreation* use.

Parameter	Condition	Value
Temperature	Highest 7- DADMax	16º C. Thornton Creek also has *Supplemental Spawning and Incubation* criteria: during Sept 15 - May 15, highest 7-DADMax should not exceed 13° C.
Dissolved Oxygen	Lowest 1-day minimum	9.5 mg/L.
pH	--	Range within 6.5 – 8.5, with a human-caused variation within the above range of < 0.2 units.

DADMax: Daily average of the daily maximum temperature.

Longfellow Creek subbasin

Beneficial uses for Longfellow Creek include *Salmonid Spawning, Rearing, and Migration* habitat and *Primary Contact Recreation*. The numeric water quality standards for temperature, dissolved oxygen, and pH for Longfellow Creek (freshwater) are described in Table 6.

Skagit-Samish basin

Beneficial uses for the Samish River, Indian Slough, Big Ditch, and Browns Slough are *Salmonid Spawning, Rearing, and Migration Habitat* and *Primary Contact Recreation*. The Samish River, Indian Slough, and Big Ditch sites are freshwater and must meet the water quality standards described in Table 6. The site on Browns Slough is marine water and must meet the water quality standards described in Table 7.

Lower Yakima basin

Beneficial uses for Marion Drain, Sulphur Creek Wasteway, and Spring Creek are *Salmonid Spawning, Rearing, and Migration Habitat*. The freshwater water quality standard described in Table 6 applies to these sites.

Wenatchee-Entiat basins

Beneficial uses for the Mission Creek, Brender Creek, Wenatchee River, and Entiat River are *Salmonid Spawning, Rearing, and Migration*. The water quality standard described in Table 6 applies to these sites.

Table 6. Freshwater water quality standards for temperature, dissolved oxygen, and pH for *Salmonid Spawning, Rearing, and Migration Habitat* use and *Primary Contact Recreation* use.

Parameter	Condition	Value
Temperature	Highest 7- DADMax	17.5º C. The Wenatchee River site also has *Supplemental Spawning and Incubation* criteria: during Oct 1 - May 15, highest 7-DADMax should not exceed 13° C.
Dissolved Oxygen	Lowest 1-day minimum	8.0 mg/L
pH	--	Range within 6.5 – 8.5, with a human-caused variation within the above range of < 0.5 units.

Table 7. Marine water quality standards for temperature, dissolved oxygen, and pH for *Aquatic Life Excellent* use.

Parameter	Condition	Value
Temperature	Highest 7- DADMax	16°C (60.8°F).
Dissolved Oxygen	Lowest 1-day minimum	6.0 mg/L.
pH	--	Range within 7.0 – 8.5, with a human-caused variation within the above range of < 0.5 units.

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Results

Results from the 2010 monitoring season are summarized by basin in the following sections. All results for the 2010 season are available through Ecology's EIM system, www.ecy.wa.gov/eim/.

Pesticide calendars for 2010 are included in Appendix E. The calendars provide a chronological overview of concentrations and detections during 2010. The calendars also compare EPA Pesticide Registration Toxicity Criteria and EPA National Recommended Water Quality Criteria *(assessment criteria)* to numeric Washington State Water Quality Standards *(water quality standards).* Refer to Appendix D, Assessment Criteria and Water Quality Standards, in this report for information on assessment criteria development.

Western Washington

Cedar-Sammamish Basin (WRIA 8): Thornton Creek, 2010

Pesticide Detections and Concentrations

A total of 27 sampling events were conducted on Thornton Creek between March 8 and September 8, 2010. During this period, there were 58 detections of ten pesticides.

Of the 10 types of compounds detected, there were three insecticides (7 detections), six herbicides (42 detections), and a wood preservative (9 detections).

The number and types of pesticide detections are presented in Figure 2. The maximum number of pesticides detected during a sampling event was seven (Figure 2). The most frequently detected pesticides are described in Table 8.

Table 8. Most frequently detected pesticides for Thornton Creek, 2010.

Pesticide	Pesticide Type	Number of Detections
Dichlobenil	Herbicide	24
Pentachlorophenol	Wood Preservative	9
2,4-D	Herbicide	7

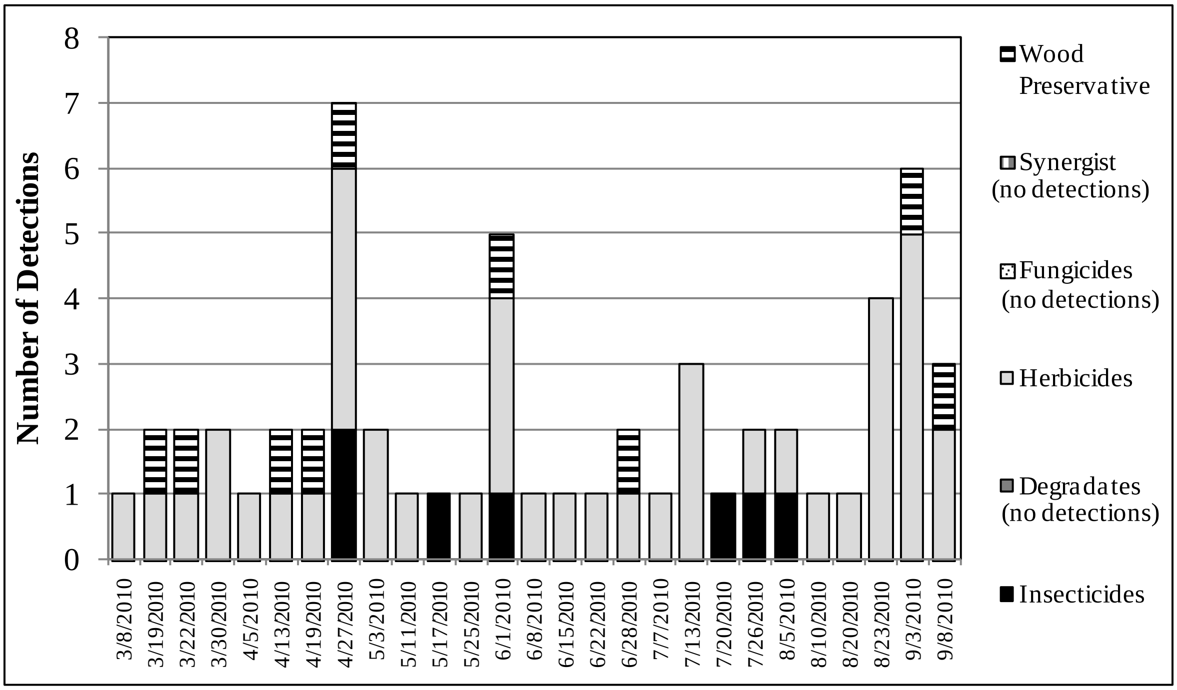

Figure 2. Pesticide detections by week and type for Thornton Creek, 2010.

Table 9 presents a pesticide calendar for Thornton Creek. This pesticide calendar is included as an example; the rest of the calendars are included in Appendix E. The calendar provides a chronological overview of concentrations and detections during 2010 and compares pesticide concentrations to *assessment criteria* and *water quality standards*. In the calendars, the number below the months indicate sample week. Numbers below Appendix E, Table E-1, present the color codes used to compare detected pesticide concentrations to assessment criteria.

In 2010, pesticide concentrations in Thornton Creek met (did not exceed) any available *assessment criteria* or *water quality standard* (Appendix D).

Table 9. Pesticide detections for Thornton Creek, 2010.

Pesticide results are in µg/L, and TSS results are in mg/L.

Month		**March**				**April**				**May**				**June**					**July**				**August**				**Sep**	
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
2,4-D	H				0.073				0.110	0.056										0.095						0.087	0.033	0.067
Carbaryl	I-C								0.005																			
Dichlobenil	H	0.017	0.008	0.013	0.027	0.011	0.009	0.009	0.044	0.014	0.010		0.008	0.021	0.012	0.014	0.014	0.007	0.001	0.002		0.012	0.012	0.016	0.012	0.015	0.008	
Diuron	H								0.039					0.053												0.028		
Imidacloprid	I-N											0.005		0.005							0.004	0.003	0.003					
MCPA	H																										0.031	
MCPP	H								0.050																		0.022	
Pentachlorophenol	WP		0.018	0.032			0.019	0.021	0.031					0.021				0.018									0.024	0.049
Propoxur	I-C								0.008																			
Triclopyr	H													0.035						0.063						0.064	0.150	0.210
Total Suspended Solids	NA	6.0	2.0	13.0	9.0	5.0	6.0	6.0	15.0	7.0	5.0	8.0	18.5	5.0	6.0	9.0	6.0	4.3	4.0	8.0	11.0	7.3	5.0	5.0	6.0	4.0	5.0	12.0

C: Carbamate, H: Herbicide, I: Insecticide, N: Neonicotinoid, NA: Not applicable, WP: Wood Preservative

Conventional Parameters

Conventional water quality parameters were measured in Thornton Creek. All summaries are based on point (discrete) measurements obtained during the time of sampling. Table 10 summarizes results for TSS, streamflow, pH, conductivity, and dissolved oxygen.

PH levels met water quality standards during 2010. Dissolved oxygen dropped below the 9.5 mg/L water quality standard nine times during July 13 – September 8, 2010.

Table 10. Mean, minimum, and maximum for discrete conventional parameter measurements for Thornton Creek, 2010.

Parameter	Number	Mean	Minimum	Maximum
Total Suspended Solids (mg/L)	27	7	2	19
Discharge (cfs)	27	8.0	4.0	20.0
pH (s.u.)	27	7.8	7.2	8.0
Conductivity (umhos/cm)	27	218	132	247
Dissolved Oxygen (mg/L)	27	9.9	8.3	11.6

In addition to discrete measurements for stream temperature, continuous (30-minute interval) measurements were collected year-round. During September 15 - May 15, the highest 7-Daily Average Daily Maximum (DADMax) should not exceed 13° C; during the rest of the year, the highest 7-DADMax should not exceed 16°C.

In 2010, stream temperatures did not meet (exceeded) the standard during the following periods:

- May 12-15, >13°C.
- July 6-August 27, >16°C.
- September 14, >16°C.
- September 15-October 13, >13°C.

Green-Duwamish Basin (WRIA 9): Longfellow Creek, 2010

Pesticide Detections and Concentrations

A total of 27 sampling events were conducted on Longfellow Creek between March 8 and September 8, 2010. During this period, there were 78 detections of 15 pesticides and degradates. The 15 compounds included five insecticides (13 detections), seven herbicides (58 detections), one insecticide degradate (1 detection), a fungicide (1 detection), and a wood preservative (5 detections).

The maximum number of pesticides detected during a sampling event was eight, and most of the pesticides detected were herbicide compounds (Figure 3). The most frequently detected pesticides were herbicides (Table 11).

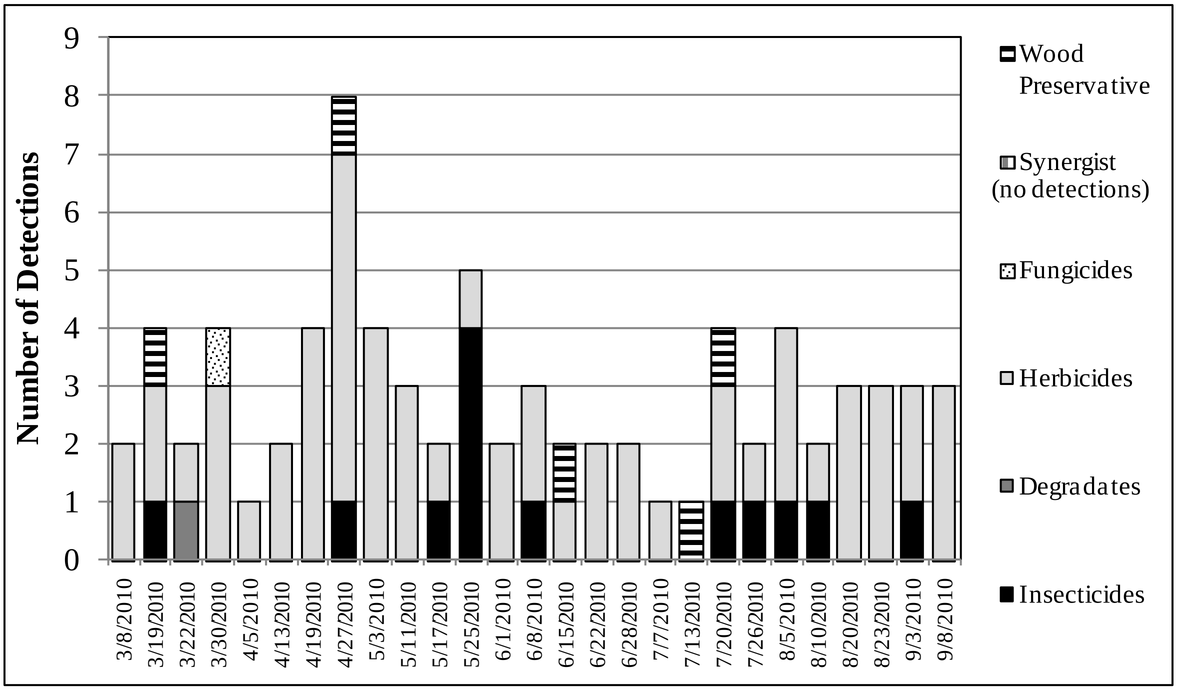

Figure 3. Pesticide detections by week and type for Longfellow Creek, 2010.

Table 11. Most frequently detected pesticides for Longfellow Creek, 2010.

Pesticide	Pesticide Type	Number of Detections
Dichlobenil	Herbicide	22
Triclopyr	Herbicide	19
2,4-D	Herbicide	12

Table 12 presents a pesticide calendar for Longfellow Creek. This calendar is included as an example; the rest of the calendars are included in Appendix E. The calendar provides a chronological overview of concentrations and detections during 2010 and compares pesticide concentrations to *assessment criteria* and *water quality standards*. In the calendars, the number below the months indicate sample week. Appendix E, Table E-1, presents the color codes used to compare detected pesticide concentrations to *assessment criteria* and *water quality standards.*

In 2010, pesticide concentrations in Longfellow Creek met (did not exceed) any available *assessment criteria* and *water quality standards* (Appendix D).

Table 12. Pesticide detections in Longfellow Creek, 2010.

Pesticide results are in µg/L, and TSS results are in mg/L.

Month		**March**				**April**				**May**				**June**					**July**				**August**				**Sep**	
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
2,4-D	H				0.057		0.036	0.032	0.540	0.150	0.068										0.024		0.030		0.130	0.042	0.038	0.086
Carbaryl	I-C								0.003																			
Carbofuran	I-C												0.003															
Dicamba I	H								0.076																			
Dichlobenil	H	0.017	0.011		0.054	0.027	0.017	0.021	0.210	0.078	0.026	0.008	0.010	0.024	0.010		0.014	0.006	0.002			0.013	0.011	0.015	0.012	0.011		0.017
Diuron	H								0.030																			
Imidacloprid	I-N		0.007									0.005	0.006		0.005						0.004	0.005	0.003	0.006			0.004	
MCPP	H								0.160	0.055																		
Metalaxyl	F				0.042																							
Methomyl	I-C												0.004															
Oxamyl	I-C												0.004															
Oxamyl oxime	D-C			0.013																								
Pentachlorophenol	WP		0.018						0.035							0.017				0.033	0.016							
Prometon	H							0.110																				
Triclopyr	H	0.034	0.033	0.031	0.080			0.049	0.140	0.092	0.049			0.070	0.053	0.031	0.036	0.031			0.031		0.037		0.080	0.052	0.048	0.150
Total Suspended Solids	NA	3.0	2.0	2.0	7.0	8.0	2.0	<3.0	17.0	6.0	5.3	4.0	4.0	4.0	4.0	4.0	3.0	4.0	2.0	9.0	2.0	3.0	2.0	4.0	2.0	9.0	2.0	3.0

C: Carbamate, D: Degradate, F: Fungicide, H: Herbicide, I: Insecticide, N: Neonicotinoid, NA: Not applicable, WP: Wood Preservative

Conventional Parameters

Conventional water quality parameters were measured in Longfellow Creek. Table 13 summarizes results for TSS, streamflow, pH, conductivity, and dissolved oxygen. All summaries are based on point (discrete) measurements obtained during the time of sampling.

All field data collected met the pH water quality standard (between 6.5-8.5 s.u.) and the 8.0 mg/L minimum dissolved oxygen water quality standard.

Table 13. Mean, minimum, and maximum for discrete conventional parameter measurements for Longfellow Creek, 2010.

Parameter	Number	Mean	Minimum	Maximum
Total Suspended Solids (mg/L)	27	4	2	17
Discharge (cfs)	27	1.6	0.7	5.8
pH (s.u.)	27	8.0	7.5	8.2
Conductivity (umhos/cm)	27	284	167	328
Dissolved Oxygen (mg/L)	27	10.3	9.2	11.9

In addition to discrete temperature measurements, continuous (30-minute interval) measurements were collected year-round. The temperature standard for Longfellow Creek is: the 7-DADMax should not exceed 17.5° C. During 2010 temperature did not meet (exceeded) the standard during the following periods:

- July 9-10, >17.5 °C.
- August 14-16, >17.5 °C.

Lower Skagit-Samish Basin (WRIA 3), 2010

Pesticide Detections and Concentrations

The lower Skagit-Samish sites were sampled for 27 consecutive weeks from March 8 to September 8, 2010. The lower Skagit-Samish sites are: upstream and downstream Big Ditch, Indian Slough, Browns Slough, and the Samish River. Browns Slough is classified as marine water; the other four sites are classified freshwater.

For the five Skagit-Samish sites combined, there were a total of 39 compounds detected: eight insecticides, 26 herbicides, two fungicides, one degradate, a wood preservative, and a synergistic compound. The most frequently detected pesticides found at the Skagit-Samish sites are described in Table 14.

Table 14. Most frequently detected pesticides for the Skagit-Samish sites, 2010.

Pesticide	Pesticide Type	Number of Detections
Dichlobenil	Herbicide	65
Bromacil	Herbicide	47
Imidacloprid	Insecticide	42
Triclopyr	Herbicide	38
Metolachlor	Herbicide	37
Pentachlorophenol	Wood preservative	24
Carbofuran	Insecticide	19

For the Skagit-Samish sites, Indian Slough had the greatest number of pesticide detections, with 145 detections. The upstream Big Ditch site had 140 detections and the downstream site 136 detections. Browns Slough had 76 detections and the Samish River nine detections. The Big Ditch site had 89 detections, Browns Slough had 46 detections, and the Samish River had 20 detections. The greatest number of pesticide detections during a sample event occurred on June 1, 2010, with 14 pesticide detections in both Indian Slough and Browns Slough.

Big Ditch

Two sites on Big Ditch were sampled in 2010. Water quality at the upstream site is influenced by industrial land use and stormwater, while the downstream site is influenced by agricultural land use. In 2010, 30 compounds were detected in Big Ditch: 22 at the upstream site and 26 at the downstream site. Eighteen of these compounds were found in common between the two sites.

At the upstream site, the maximum number of pesticides detected during a sampling event was 10; this occurred during three sample events (Figure 4). At the downstream site, the maximum number of pesticides detected during a sample event was 13 (Figure 5).

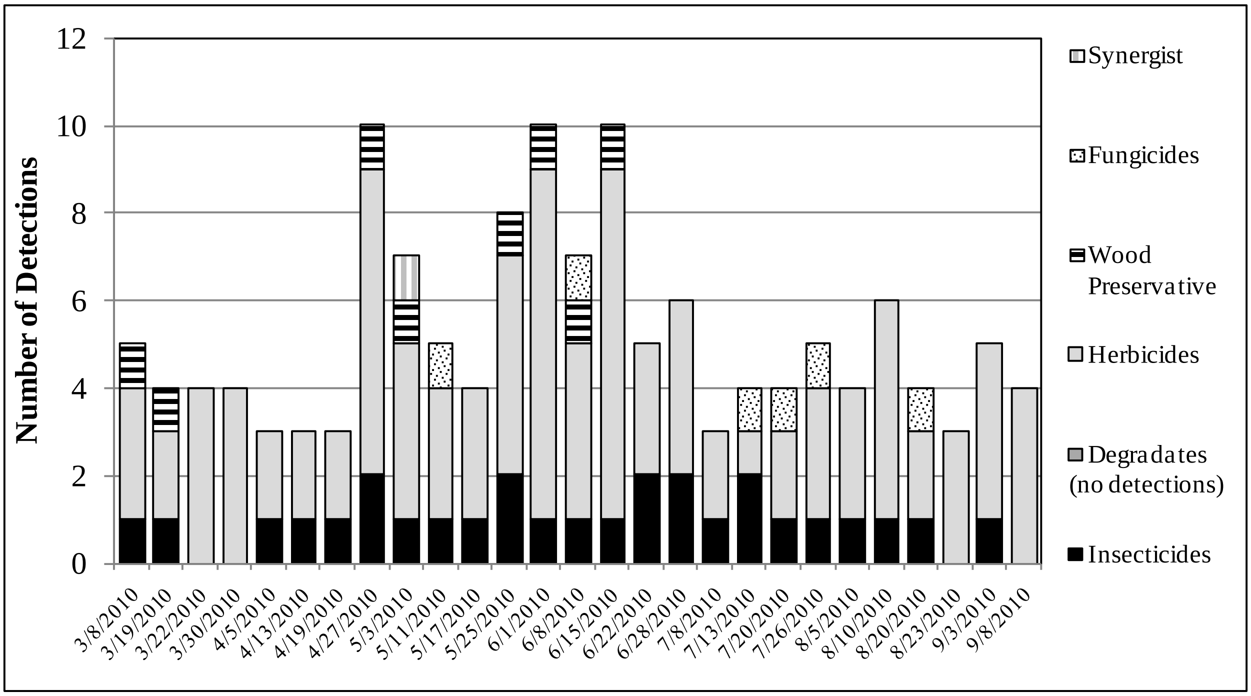

Figure 4. Pesticide detections by week and type for the upstream Big Ditch site, 2010.

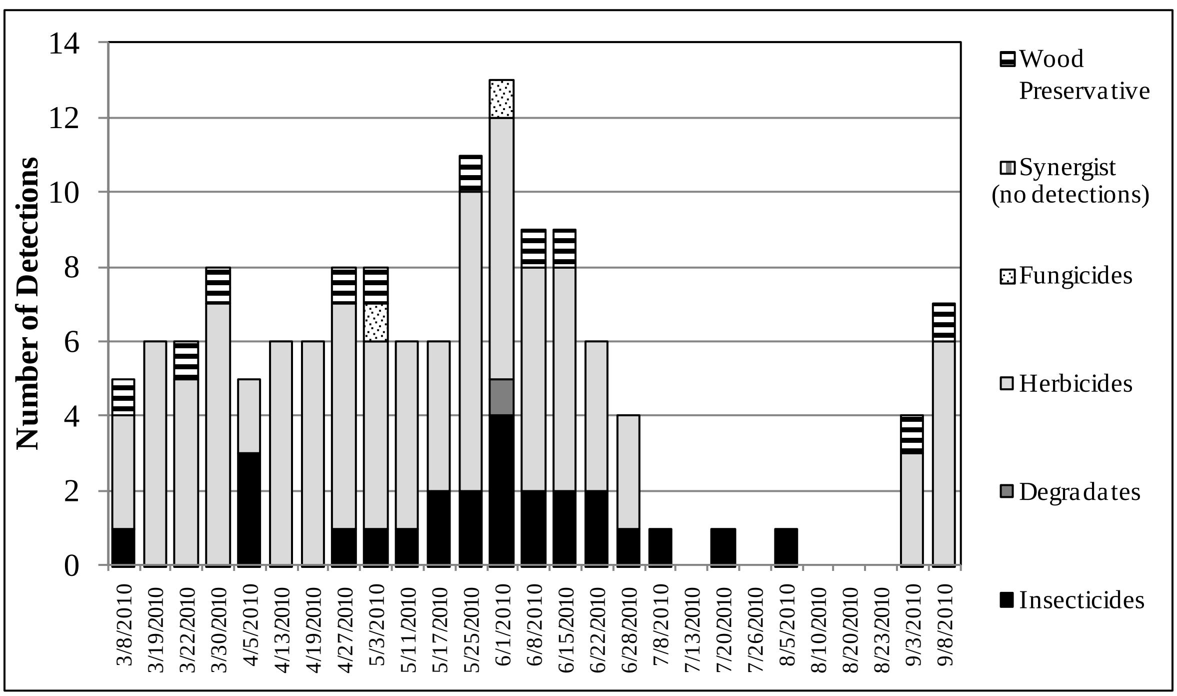

Figure 5. Pesticide detections by week and type for the downstream Big Ditch site, 2010.

Most of the pesticides detected were herbicide compounds. At the upstream site, pesticides were detected during every sampling event. At the downstream site, the greatest number of detections occurred from March through June. The most frequently detected pesticides during 2010 for both the upstream and downstream Big Ditch sites are presented in Table 15.

Table 15. Most frequently detected pesticides for the Big Ditch sites, 2010.

Pesticide	Pesticide Type	Number of Detections
Upstream Big Ditch		
Dichlobenil	Herbicide	25
Bromacil	Herbicide	22
Imidacloprid	Insecticide	22
Downstream Big Ditch		
Metolachlor	Herbicide	18
Dichlobenil	Herbicide	15
Imidacloprid	Insecticide	13

Tables E-4 and E-5 in Appendix E present the pesticide calendars for 2010 for the upstream and downstream Big Ditch sites, respectively. The calendars provide a chronological overview of concentrations and detections and compare pesticide concentrations to *assessment criteria* and *water quality standards*. Both Big Ditch sites met (did not exceed) any available *assessment criteria* or *water quality standard* (Appendix D).

Indian Slough

During 2010 there were 145 detections of 23 compounds. These 23 compounds were 17 herbicides, four insecticides, one fungicide, and a wood preservative. The number and types of pesticide detections are presented in Figure 6. The maximum number of pesticides detected during a sampling event was 13 (Figure 6). Of the 145 pesticide detections, 129 were herbicides.

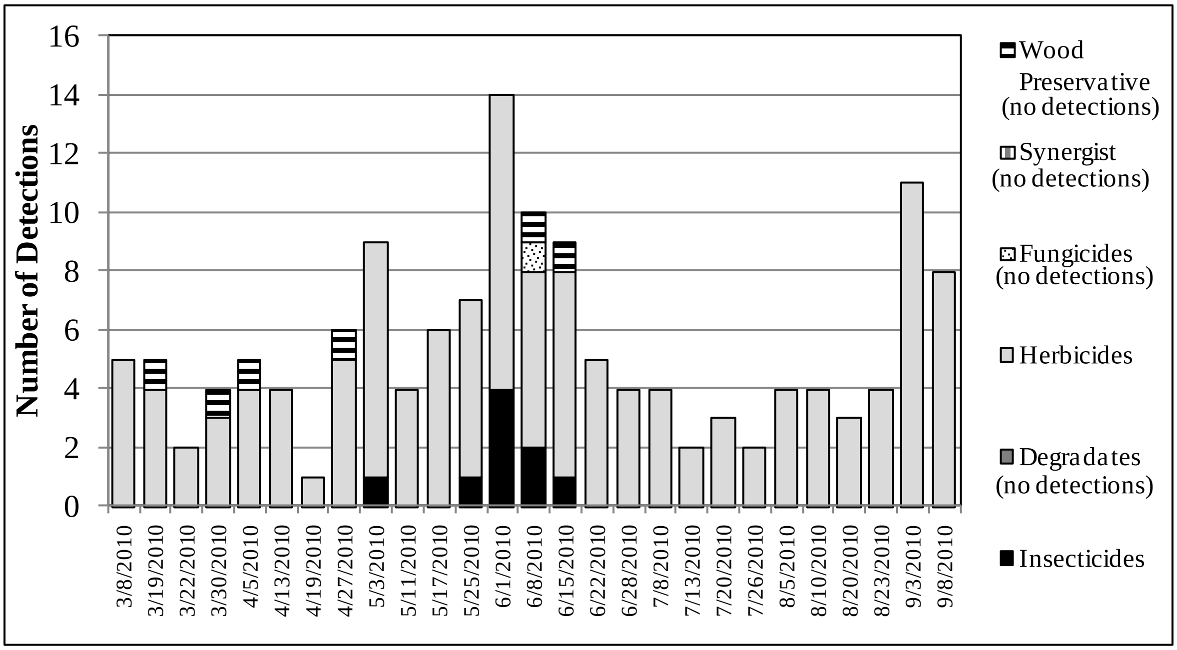

Figure 6. Pesticide detections by week and type for Indian Slough, 2010.

In Indian Slough the most frequently detected pesticides were herbicides (Table 16).

Table 16. Most frequently detected pesticides for Indian Creek, 2010.

Pesticide	Pesticide Type	Number of Detections
Bromacil	Herbicide	24
Dichlobenil	Herbicide	17
Hexazinone	Herbicide	15
Triclopyr	Herbicide	14
Diphenamid	Herbicide	12

During the 2006-09 monitoring, the herbicide diphenamid was detected 68 times in Indian Slough. Diphenamid has not been registered for use by EPA since 1991 (EPA, 2002). It is not known why diphenamid is detected so frequently in Indian Slough. Data quality for herbicide parameters is excellent, and detections are not likely due to field or laboratory error.

Appendix E, Table E-6, presents the pesticide calendar for 2010 for Indian Slough. The calendar provides a chronological overview of concentrations and detections and compares pesticide concentrations to *assessment criteria* and *water quality standards* (Appendix D). In 2010, pesticide concentrations in Indian Slough met (did not exceed) any available marine *assessment criteria* or *water quality standards* (Appendix D).

Browns Slough

Browns Slough is sampled downstream of a tidegate. Due to higher salinity at this site, marine *assessment criteria* and *water quality standards* are used for evaluating water quality. During

2010 there were 76 detections of 15 pesticides. The 15 pesticides were 12 herbicides, two insecticides, and a fungicide. The number and types of pesticide detections are presented in Figure 7. The maximum number of pesticides detected during a sampling event was 14 (Figure 7).

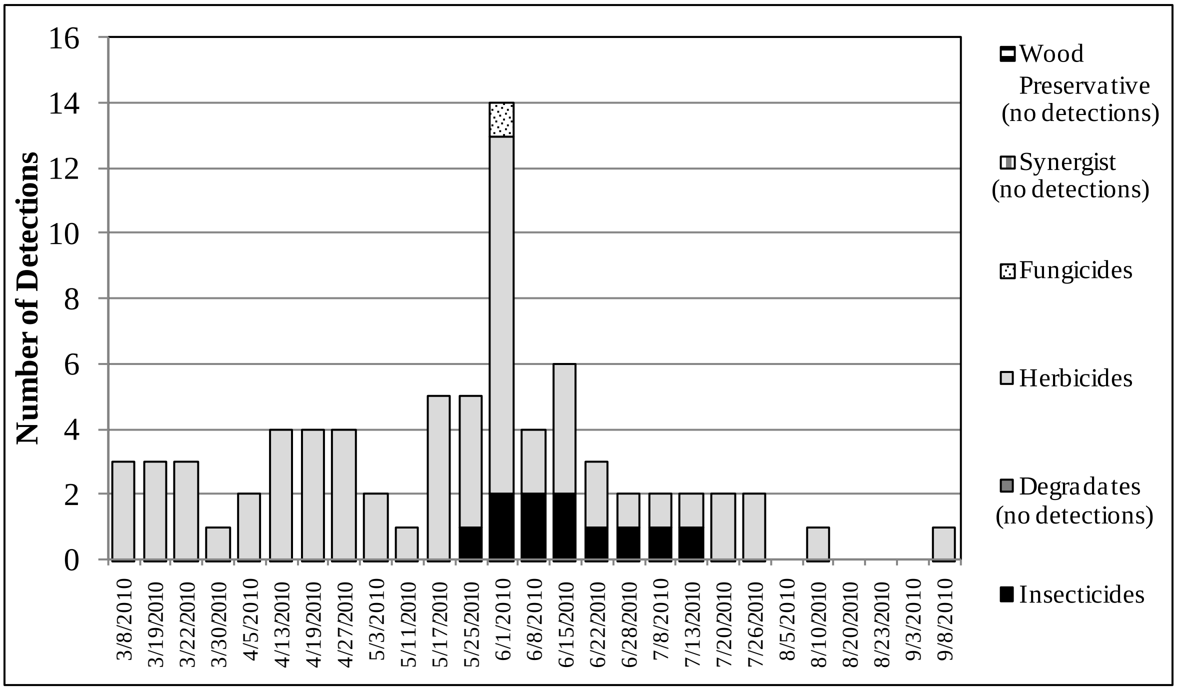

Figure 7. Pesticide detections by week and type for Browns Slough, 2010.

Most of the pesticides detected were herbicides (64 of 76). The most frequently detected pesticides are described in Table 17.

Table 17. Most frequently detected pesticides for Browns Slough, 2010.

Pesticide	Pesticide Type	Number of Detections
DCPA (dacthal)	Herbicide	20
Metolachlor	Herbicide	9
Carbofuran	Insecticide	6
Dichlobenil	Herbicide	6

Appendix E, Table E-7, presents the pesticide calendar for 2010 for Browns Slough. The calendar provides a chronological overview of concentrations and detections and compares pesticide concentrations to *assessment criteria* and *water quality standards*. In 2010, pesticide concentrations in Browns Slough met (did not exceed) any available marine *assessment criteria* or *water quality standards* (Appendix D).

Samish River

A total of 27 sampling events were conducted on the Samish River between March 8 and September 8, 2010. There were very few pesticide detections (9 detections total); these were four herbicides (eight detections) and an insecticide (one detection). The number and types of pesticide detections are presented in Figure 8. The maximum number of pesticides detected during a sampling event was two (Figure 8). The most commonly detected pesticide was the herbicide 2,4-D, with three detections.

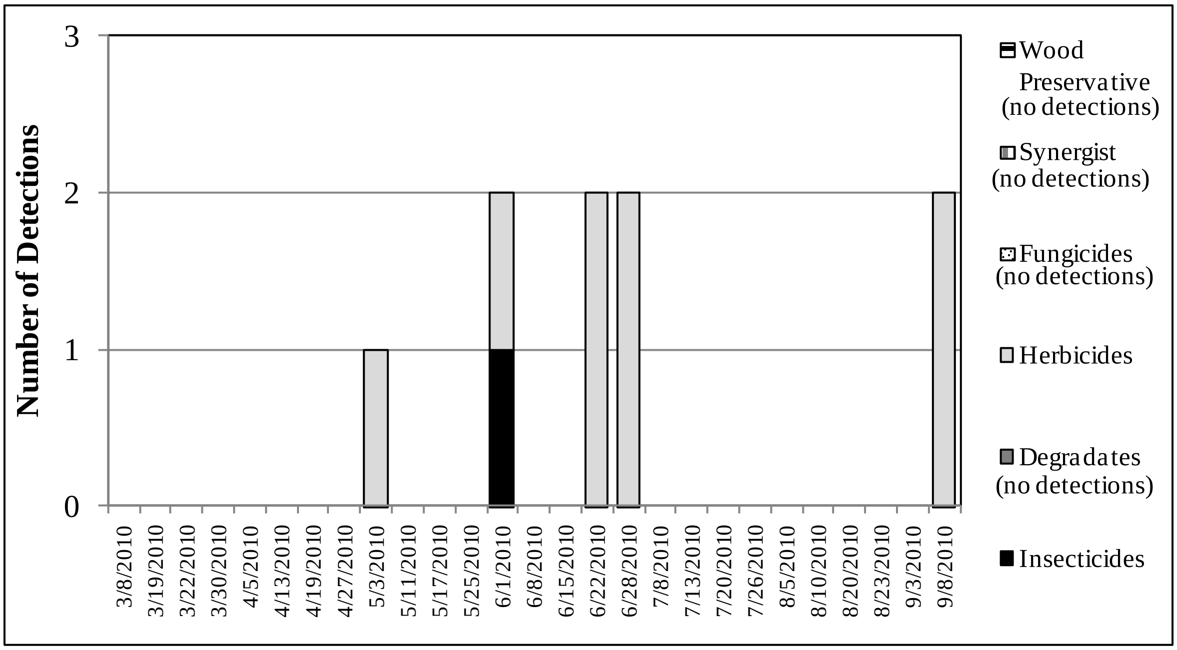

Figure 8. Pesticide detections by week and type for the Samish River, 2010.

Appendix E, Table E-8, presents a pesticide calendar for 2010 for the Samish River. The calendar provides a chronological overview of concentrations and detections and compares pesticide concentrations to *assessment criteria* and *water quality standards*. The Samish River met (did not exceed) any available *assessment criteria* or *water quality standards* (Appendix D).

Conventional Parameters

Conventional water quality parameters were measured at the Skagit-Samish sites. Table 18 summarizes results for TSS, streamflow, pH, conductivity, and dissolved oxygen. All summaries are based on point (discrete) measurements obtained during the time of sampling. Browns Slough is a marine site and must meet marine water quality standards; all the other Skagit-Samish sites must meet freshwater quality standards.

During 2010 dissolved oxygen levels did not meet the 8.0 mg/L minimum freshwater quality standard in upper Big Ditch (10 times), lower Big Ditch (6 times), and Indian Slough (17 times). The Samish River met dissolved oxygen water quality standards during all sampling events.

Browns Slough did not meet the 6.0 mg/L minimum marine water quality standard for four sampling events.

Upper Big Ditch, Indian Slough, and Samish River met pH water quality standards. Both Browns Slough (marine) and lower Big Ditch (freshwater) did not meet the pH standard once during the sample period with pH levels of 8.8 and 9.4 s.u., respectively.

Table 18. Mean, minimum, and maximum for discrete conventional parameter measurements for the Skagit-Samish sites, 2010.

Site	Total Suspended Solids (mg/L)	Flow (cfs)	pH (s.u.)	Conductivity (umhos/cm)	Dissolved Oxygen (mg/L)
Big Ditch (upstream)					
number	27	27	26	27	27
mean	7	2.3	7.0	319	8.5
minimum	3	0.5	6.8	213	6.1
maximum	16	6.2	7.3	448	10.3
Big Ditch (downstream)					
number	27	24	26	27	27
mean	7	13.3	7.6	475	10.6
minimum	< 1	2.4	6.8	50	6.1
maximum	25	34	9.4	925	16.0
Indian Slough					
number	27	27	26	27	27
mean	8	26.5	7.0	1040	7.4
minimum	2	0.7	6.6	268	4.4
maximum	22	56	7.5	7400	11.2
Brown Slough					
number	27	27	26	27	27
mean	7	4.7	7.6	10083	10.3
minimum	2	< 0.1	7.1	90	2.8
maximum	17	13	8.7	19106	20.1
Samish River					
number	27	27	26	27	27
mean	15	196	7.5	99	10.7
minimum	2	34	6.9	54	9.8
maximum	151	859	8.4	135	12.8

In addition to the discrete temperature measurements, continuous (30-minute interval) measurements were collected year-round. The temperature standard for the freshwater sites is that the 7-DADMax should not exceed 17.5° C; and for the marine water site is that the 7-DADMax should not exceed 16.0° C. Table 19 describes the periods that temperature did not meet (exceeded) the standard.

Table 19. Periods of water temperature exceedance for the Skagit-Samish sites, 2010.

Site	Periods When Temperature Did Not Meet Standards
Big Ditch (upstream) >17.5°C	Aug 12-21.
Big Ditch (downstream) >17.5°C	May 10-19, June 3-29, July 4-Sept 9, Sept 15-18.
Indian Slough >17.5°C	June 21-28, July 4-Sept 2, Sept 5-25.
Browns Slough >16.0°C	Apr 13-21, Apr 25-May 1, May 6-Oct 8.
Samish River >17.5°C	July 8-14, July 22-Aug 6, Aug 11-19.

Eastern Washington

Lower Yakima Basin (WRIA 37), 2010

Pesticide Detections and Concentrations

In the lower Yakima River basin, downstream Spring Creek, Marion Drain, and Sulphur Creek Wasteway were sampled for 27 consecutive weeks from March 10 to September 8, 2010. The upstream Spring Creek site was sampled every other week during the same period for a total of 14 sampling events. In Marion Drain, weekly sampling for organophosphates continued from September 13 through October 26, 2010. Historically, Marion Drain sampling for organophosphates has continued through the end of October.

For the four sites combined, there were a total of 368 detections of 35 types of pesticides and degradates. These 35 compounds were 12 insecticides, 21 herbicides, and two insecticide degradates. Marion Drain had the greatest number of detections, 162, and the greatest number of sampling events. Sulphur Creek Wasteway had 115 detections. The downstream Spring Creek site had 62 detections. The upstream Spring Creek site had 29 detections and the least number of sampling events.

The most frequently detected pesticides found at the lower Yakima sites are described in Table 20.

Table 20. Most frequently detected pesticides for the lower Yakima sites, 2010.

Pesticide	Pesticide Type	Number of Detections
2,4-D	Herbicide	67
Imidacloprid	Insecticide	49
Dicamba I	Herbicide	38
Terbacil	Herbicide	28
Carbaryl	Insecticide	24
Bromacil	Herbicide	19
Diuron	Herbicide	17
Bentazon	Herbicide	16

Spring Creek

Two sites on Spring Creek were sampled in 2010. The upstream site was sampled every two weeks, and the downstream site was sampled weekly. A total of 17 pesticide and degradate types were detected in Spring Creek: 11 herbicides, five insecticides, and one insecticide degradate. A total of 12 pesticides were detected in common between the upstream and downstream sites. A total of 14 pesticides were found upstream, and 15 were found downstream. The number and types of pesticide detections are presented in Figure 9 for the upstream site and Figure 10 for the downstream site. The maximum number of pesticides detected during a sampling event at the upstream site was five (Figure 9) and at the downstream site was six (Figure 10).

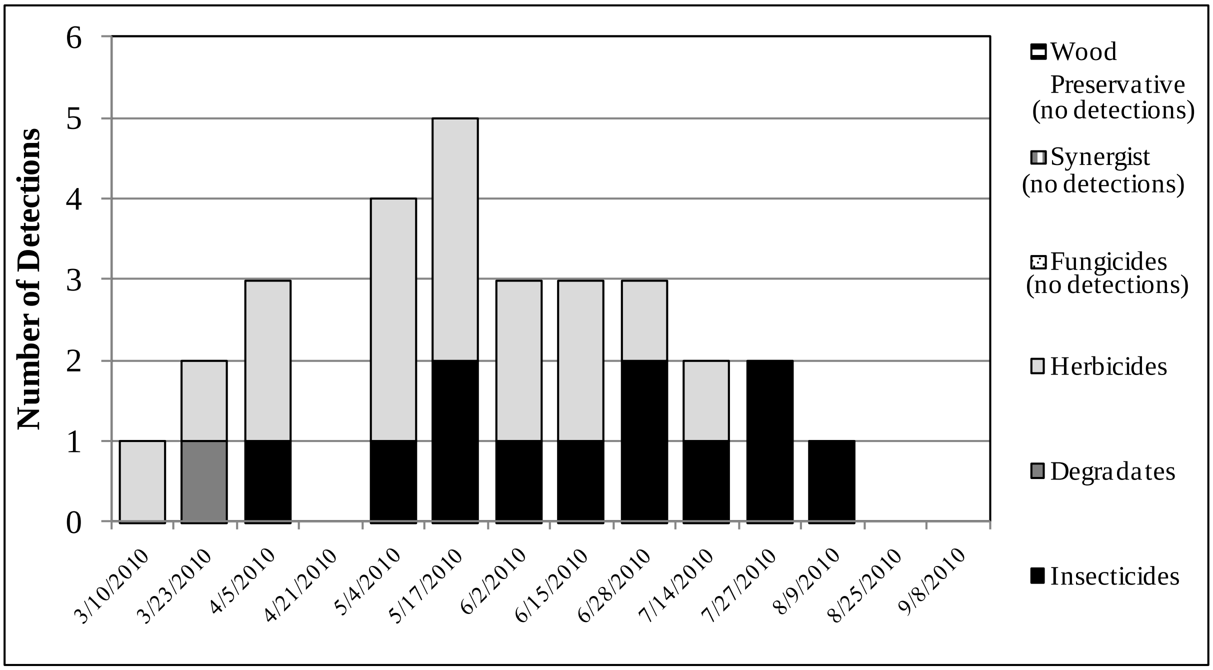

Figure 9. Pesticide detections by week and type for upstream Spring Creek, 2010.

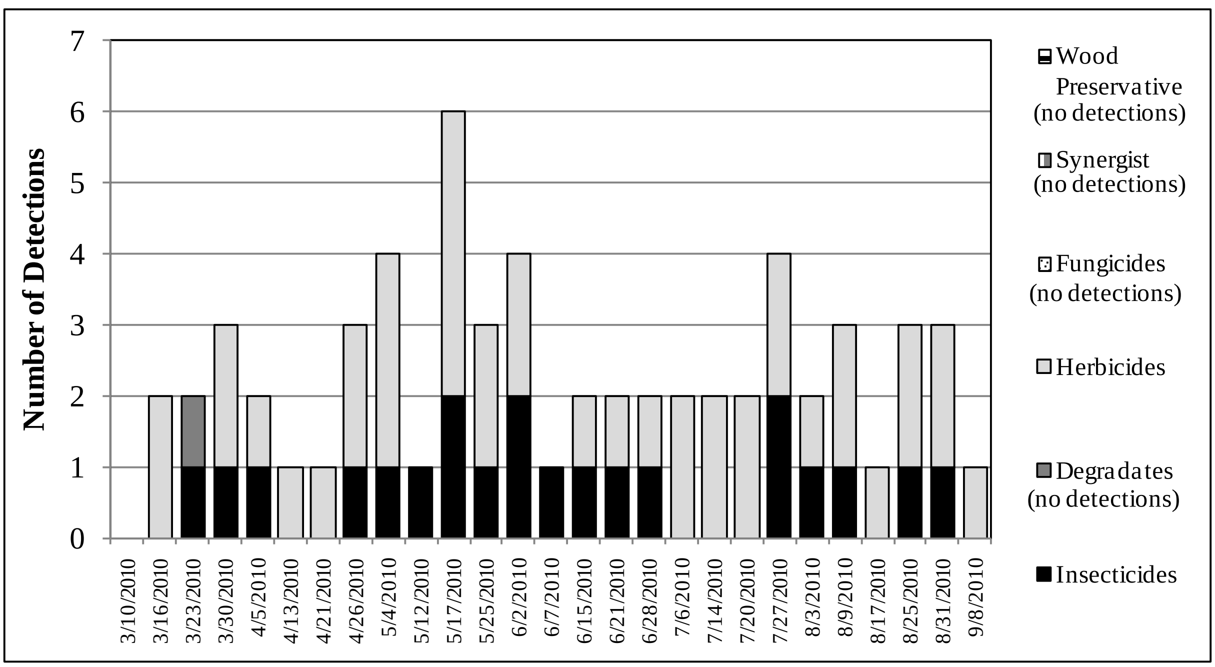

Figure 10. Pesticide detections by week and type for downstream Spring Creek, 2010.

The most frequently detected pesticides at the upstream and downstream sites were similar. Table 21 describes the most frequently detected pesticides for the upstream and downstream Spring Creek sites in 2010.

Table 21. Most frequently detected pesticides for the Spring Creek sites, 2010.

Pesticide	Pesticide Type	Number of Detections
Upstream Spring Creek		
Imidacloprid	Insecticide	7
2,4-D	Herbicide	5
Downstream Spring Creek		
2,4-D	Herbicide	19
Imidacloprid	Insecticide	11

Appendix E, Tables E-9 and E-10, present the pesticide calendars for 2010 for the upstream and downstream Spring Creek sites, respectively. The calendars provide a chronological overview of concentrations and detections and compare pesticide concentrations to *assessment criteria* and *water quality standards* (Appendix D).

At the downstream site, a March 30 detection of chlorpyrifos did not meet (exceeded) the chronic *water quality standard* and EPA's chronic invertebrate *criteria*. This single event met (did not exceed) the 21-day time component of the chronic invertebrate criteria.

Marion Drain

An additional seven weeks of sampling for organophosphates was conducted on Marion Drain after September 8, 2010. No pesticides were detected during the last two sampling events in late October. In addition only one pesticide, the insecticide imidacloprid, was detected between March 10 and April 13.

During 2010 there were 162 detections of 22 pesticides and a degradate. These 23 compounds were 14 herbicides, eight insecticides, and one insecticide degradate. The number and types of pesticide detections are presented in Figure 11. The maximum number of pesticides detected during a sampling event was 12 (Figure 11).

The most frequently detected pesticides in Marion Drain are described in Table 22.

Table 22. Most frequently detected pesticides for Marion Drain, 2010.

Pesticide	Pesticide Type	Number of Detections
Terbacil	Herbicide	25
2,4-D	Herbicide	20
Dicamba I	Herbicide	18
Imidacloprid	Insecticide	17
Pendimethalin	Herbicide	12

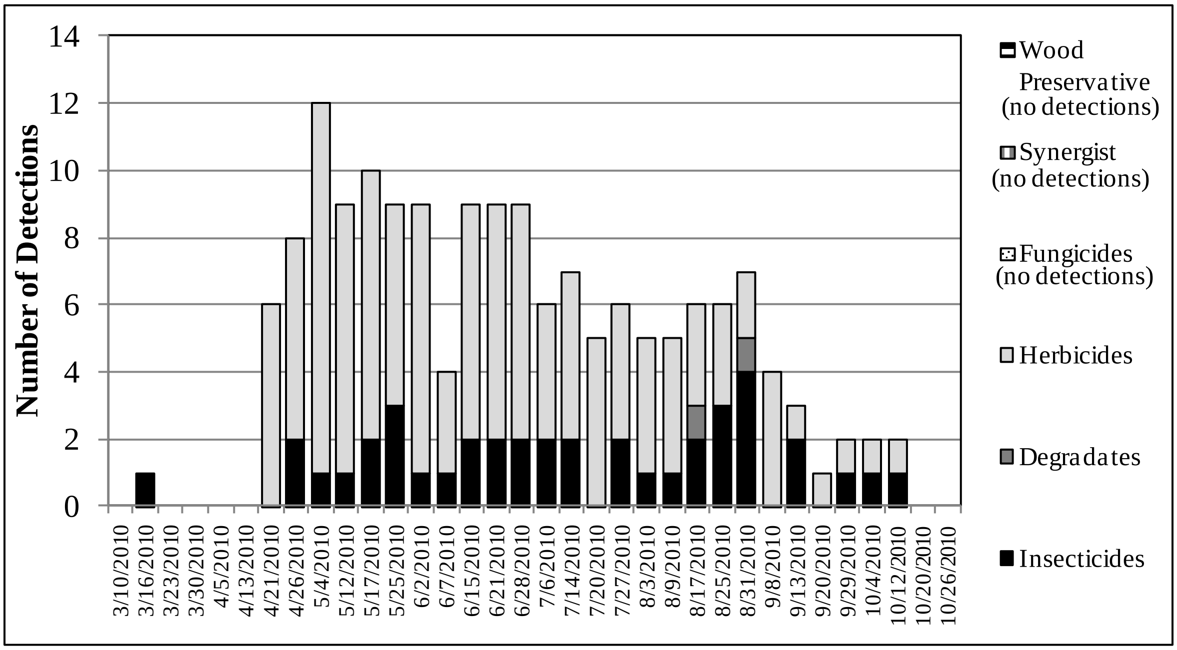

Figure 11. Pesticide detections by week and type for Marion Drain, 2010.

The pesticide calendar in Appendix E, Table E-11, provides a chronological overview of pesticide concentrations and detections during 2010 and compares concentrations to *assessment criteria* and *water quality standards* (Appendix D).

One detection of malathion on May 17 did not meet (exceeded) the registration criteria for chronic invertebrates. This single event met (did not exceed) the 21-day time component of the chronic invertebrate criteria.

Sulphur Creek Wasteway

During 2010 there were 115 detections of 21 pesticides and a degradate. These 22 compounds were 14 herbicides, seven insecticides, and one insecticide degradate. The number and types of pesticide detections are presented in Figure 12. The maximum number of pesticides detected during a sampling event was nine (Figure 12).

The the most frequently detected pesticides in Sulphur Creek Wasteway are described in Table 23.

Table 23. Most frequently detected pesticides for Sulphur Creek Wasteway, 2010.

Pesticide	Pesticide Type	Number of Detections
2,4-D	Herbicide	23
Imidacloprid	Insecticide	14
Bromacil	Herbicide	12
Dicamba I	Herbicide	12
Carbaryl	Insecticide	11

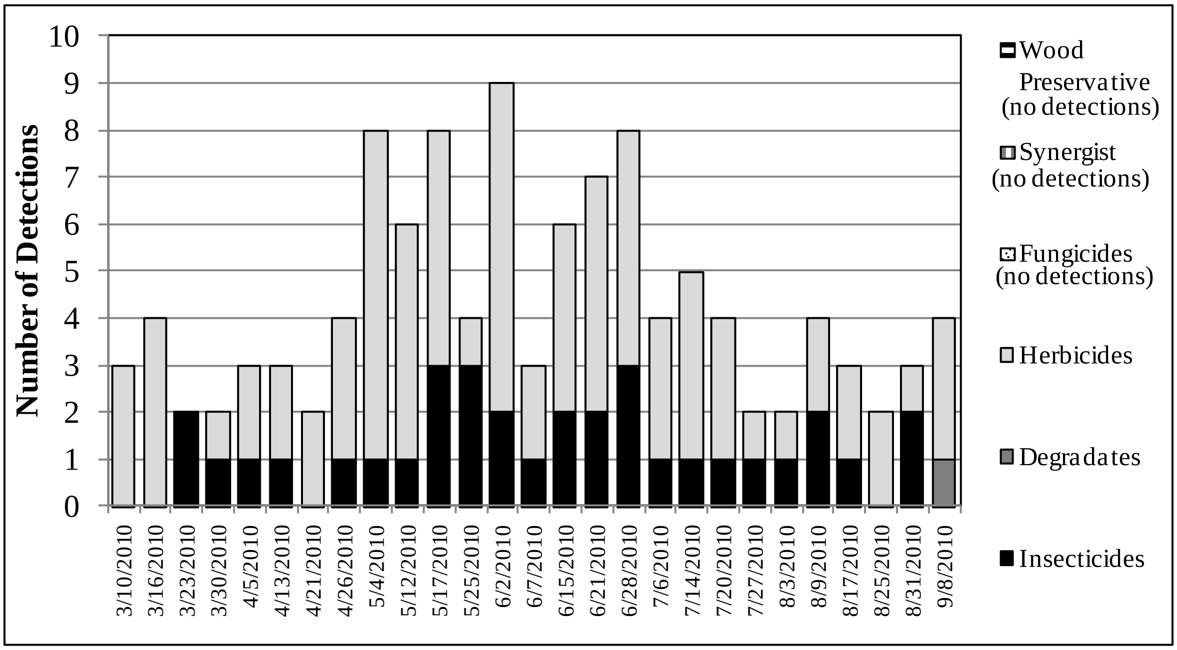

Figure 12. Pesticide detections by week and type for Sulphur Creek Wasteway, 2010.

The pesticide calendar in Appendix E, Table E-12, provides a chronological overview of pesticide concentrations and detections during 2010 and compares concentrations to *assessment criteria* and *water quality standards* (Appendix D).

In March there were two consecutive detections of chlorpyrifos that did not meet (were above) the chronic water quality standard and EPA's chronic invertebrate criteria. In addition, one of these detections also did not meet (was above) the acute water quality standard and EPA's acute invertebrate criteria.

Conventional Parameters

Conventional water quality parameters were measured at the four lower Yakima sites. Table 24 summarizes results for TSS, streamflow, pH, conductivity, and dissolved oxygen. All summaries are based on point (discrete) measurements obtained during the time of sampling. All sites must meet freshwater quality standards.

With the exception of the upstream Spring Creek site, all sites did not meet (exceeded) the pH water quality standard of 8.5 s.u. The downstream Spring Creek site exceeded the standard 17 times, Marion Drain five times, and Sulphur Creek Wasteway six times. Maximum pH values are described in Table 15.

All sites met the dissolved oxygen standard with the exception of the upstream Spring Creek site. On August 9, a dissolved oxygen value of 7.8 did not meet (fell below) the minimum water quality standard of 8.0 mg/L.

Table 24. Mean, minimum, and maximum for discrete conventional parameter measurements for the lower Yakima sites, 2010.

Site	Total Suspended Solids (mg/L)	Flow (cfs)	pH (s.u.)	Conductivity (umhos/cm)	Dissolved Oxygen (mg/L)
Spring Creek (upstream)					
Number	14	14	14	14	14
Mean	30	5.9	8.1	462	9.6
Minimum	7	2.3	7.8	334	7.8
maximum	143	11.3	8.3	656	11.2
Spring Creek (downstream)					
Number	27	27	27	27	27
Mean	9	12.0	8.7	407	10.3
Minimum	2	1.9	8.4	190	8.7
maximum	30	57.2	9.5	624	12.4
Marion Drain					
Number	34	33	34	34	34
Mean	13	158	8.1	262	12.0
Minimum	1	24.1	7.5	191	8.8
maximum	48	324	8.9	368	16.6
Sulphur Creek Wasteway					
Number	27	27	27	27	27
Mean	44	233	8.4	311	10.6
Minimum	7	51.4	8.1	193	9.2
maximum	251	493	8.8	775	12.1

In addition to the discrete temperature measurements, continuous (30-minute interval) measurements were collected year-round. The temperature thermistor for Sulphur Creek was lost; data for January 20-March 15 are not available.

The temperature standard for the freshwater sites is that the 7-DADMax should not exceed 17.5°C. None of the sites met temperature standards during all periods. Table 25 describes the periods that temperature did not meet the standard.

Table 25. Periods of water temperature exceedance for the lower Yakima sites, 2010.

Site	Periods When Temperature Did Not Meet Standards
Spring Creek (upstream) >17.5°C	Apr 17-28, May 11-Sept 24, Sept 28-Oct 5
Spring Creek (downstream) >17.5°C	May 17-21, June 9-19, June 23-Aug 30, Sept 16-22
Marion Drain >17.5°C	May 18, June 22-Sept 1, Sept 7-9, Sept 17-21, Oct 2-3
Sulphur Creek Wasteway >17.5°C	May 15-21, June 10-Sept 23, Oct 1-5

Wenatchee-Entiat Basins (WRIAs 45 and 46), 2010

Pesticide Detections and Concentrations

In the Wenatchee and Entiat basins, Peshastin, Mission, and Brender Creeks and the Wenatchee and Entiat Rivers were sampled for 27 consecutive weeks from March 9 to September 8, 2010. For the five sites combined, there was a total of 128 detections of 20 types of pesticides including degradates, a wood preservative, and a synergist. These 20 compounds were seven insecticides, seven herbicides, four degradates, a wood preservative, and a synergist.

The most frequently detected pesticides are described in Table 26. Very few pesticides were detected in the Wenatchee and Entiat Rivers and Peshastin and Mission Creeks. Detections presented in Table 26 are reflective of pesticide detections in Brender Creek (Table 27). Brender Creek had the greatest number of detections, 109. The other four sites had less than eight detections during the 2010 sampling period.

Table 26. Most frequently detected pesticides for the Wenatchee-Entiat sites, 2010.
The majority of detections included in this table are Brender Creek pesticide detections.

Pesticide	Pesticide Type[1]	Number of Detections
Endosulfan Sulfate	Endosulfan degradate	21
4,4'-DDT	Legacy Insecticide	16
4,4'-DDE	DDT (legacy) degradate	15
4,4'-DDD	DDT (legacy) degradate	10
Carbaryl	Insecticide	10

[1] Legacy pesticides are no longer allowed for use, and detections are due to historic applications.

Peshastin Creek

During 2010 very few pesticides were detected in Peshastin Creek. During 27 sample events, there were four detections: two herbicides, one insecticide, and one insecticide degradate. The maximum number of pesticides detected during a sampling event was one.

Appendix E, Table E-13, presents pesticide calendars for 2010 for Peshastin Creek. The calendars provide a chronological overview of concentrations and detections and compare pesticide concentrations to *assessment criteria* and *water quality standards* (Appendix D). On March 24, there was one detection of endosulfan that did not meet (was above) the endangered species level of concern (ESLOC) criteria for fish.

Mission Creek

During 2010 very few pesticide were detected in Mission Creek. During 27 sample events, there were three detections: one detection of the synergist compound, piperonyl butoxide, and two detections of the carbamate insecticide, carbaryl. The maximum number of pesticides detected during a sampling event was one.

Appendix E, Table E-14, presents the pesticide calendars for 2010 for Peshastin Creek. The calendars provide a chronological overview of concentrations and detections and compare pesticide concentrations to *assessment criteria* and *water quality standards* (Appendix D). Pesticide concentrations in Mission Creek met (did not exceed) any available *assessment criteria* or *water quality standards* (Appendix D).

Brender Creek

During 2010 there were 109 detections of 15 compounds: seven insecticides, three insecticide degradates, four herbicides, and a wood preservative. The number and types of pesticide detections are presented in Figure 13. The maximum number of pesticides detected during a sampling event was nine (Figure 13).

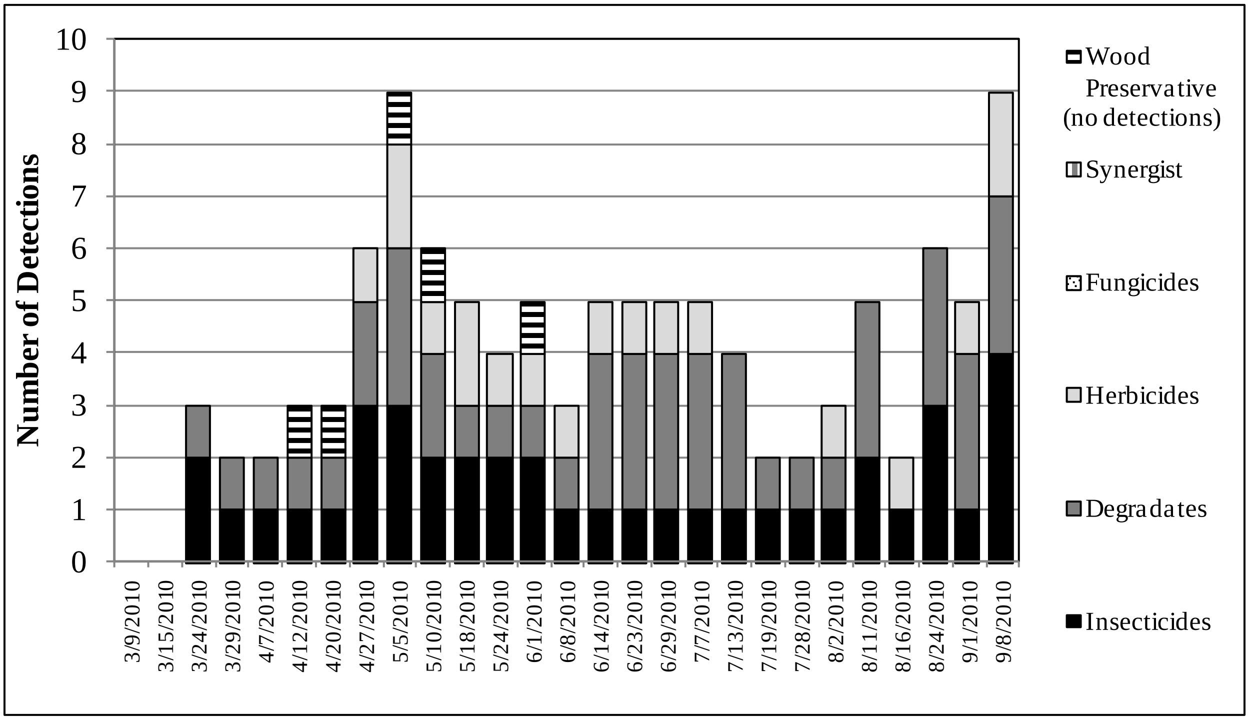

Figure 13. Pesticide detections by week and type for Brender Creek, 2010.

The most frequently detected pesticides in Brender Creek are described in Table 27.

Table 27. Most frequently detected pesticides for Brender Creek, 2010.

Pesticide	Pesticide Type	Number of Detections
Endosulfan Sulfate	Endosulfan degradate	21
4,4'DDT	Legacy[1] insecticide	15
4,4'DDE	DDT (legacy[1]) degradate	15
4,4'DDD	DDT (legacy[1]) degradate	10

[1] Legacy pesticides are no longer allowed for use, and detections are due to historic applications.

Appendix E, Table E-15, presents a pesticide calendar for 2010. The calendar provides a chronological overview of pesticide concentrations and detections and compares concentrations to *assessment criteria* and *water quality standards* (Appendix D).

There was one total endosulfan detection on March 24 that did not meet (was above) the ESLOC for fish and the chronic water quality standard.

One detection of chlorpyrifos on April 12 was above the acute and chronic water quality standard and EPA criteria.

On September 8, one detection of diazinon was above EPA's acute and chronic criteria.

DDT and DDT degradates were detected consistently throughout 2010. DDT and degradate detections were less than in previous years. In 2010 there were 18 sample events out of 27 where total DDT concentrations were above the chronic water quality standard.

Wenatchee River

During 2010 there were very few pesticides detected at the Wenatchee River site. During 27 sample events, there were five detections: three herbicides and two insecticides. The maximum number of pesticides detected during a sampling event was three on September 8.

Appendix E, Table E-16, presents the pesticide calendar for 2010 for the Wenatchee River. The calendars provide a chronological overview of concentrations and detections and compare pesticide concentrations to *assessment criteria* and *water quality standards*. During 2010 pesticide concentration met all *assessment criteria* and *water quality standards* (Appendix D).

Entiat River

During 2010 there were very few pesticides detected at the Entiat River site. During 27 sample events, there were five detections: three insecticides, one herbicide, and a synergist. The maximum number of pesticides detected during a sampling event was one.

Appendix E, Table E-17, present the pesticide calendars for 2010 for the Entiat River. The calendars provide a chronological overview of concentrations and detections and compare pesticide concentrations to *assessment criteria* and *water quality standards*.

On September 1, there was one detection of DDT that was above the chronic water quality standard and EPA criteria (Appendix D).

Conventional Parameters

Conventional water quality parameters were measured at all Wenatchee and Entiat sites. Table 17 summarizes results for TSS, streamflow, pH, conductivity, and dissolved oxygen. All summaries are based on point (discrete) measurements obtained during the time of sampling. All sites must meet freshwater quality standards.

Brender and Peshastin Creeks met the pH standard of 8.5 s.u., but the pH standard was not met nine times for the Wenatchee River, six times for the Entiat River, and twice for Mission Creek. Maximum pH values are described in Table 28. All sites met the dissolved oxygen standard.

Table 28. Arithmetic mean and range for conventional parameters (grabs) for the Wenatchee-Entiat sites, 2010.

Site	Total Suspended Solids (mg/L)	Flow (cfs)	pH (s.u.)	Conductivity (umhos/cm)	Dissolved Oxygen (mg/L)
Peshastin Creek					
number	27	27	27	27	27
mean	9	266	8.0	127	11.5
minimum	<1	18.3	7.6	80	9.3
maximum	55	887	8.4	199	13.3
Mission Creek					
number	27	27	27	27	27
mean	202	24.2	8.3	214	11.5
minimum	2	1.5	7.9	134	9.7
maximum	4180	87.8	8.7	270	13.6
Brender Creek					
number	27	27	27	27	27
mean	52	3.1	8.1	256	10.7
minimum	7	0.5	7.7	146	9.5
maximum	249	9.7	8.4	416	12.0
Wenatchee River					
number	27	27	27	27	27
mean	10	4485	8.2	54	11.9
minimum	2	766	7.1	31	9.9
maximum	70	13000	9.3	84	14.2
Entiat River					
number	27	27	27	27	27
mean	8	806	8.1	63	11.4
minimum	2	157	7.2	31	9.7
maximum	31	2440	9.0	111	12.7

Mean: Arithmetic mean.

In addition to the discrete temperature measurements, continuous (30-minute interval) measurements were collected year-round. The temperature thermistor for Brender Creek was lost; therefore, data for October 28-December 31, 2010 are not available.

The temperature standard for the freshwater sites is that the 7-DADMax should not exceed 17.5°C. The Wenatchee River has an additional temperature standard: the 7-DADMax should not exceed 13.0°C from October 1 – May 15. None of the sites met temperature standards during all periods. Table 29 describes the periods that temperature did not meet the standard.

Table 29. Periods of water temperature exceedance for the Wenatchee - Entiat sites, 2010.

Site	Periods When Temperature Did Not Meet Standards
Peshastin Creek >17.5°C	July 21-Aug 31
Mission Creek >17.5°C	July 28-Aug 12, Aug 14-23
Brender Creek >17.5°C	July 29-Aug 2
Wenatchee River >17.5°C	July 24-Sept 8
Wenatchee River >13.0°C	Oct 1-10
Entiat River >17.5°C	July 30-Aug 30

Summary for 2010

Findings

In 2010 the most commonly detected pesticide for the urban sites and the Skagit-Samish agricultural sites was the herbicide dichlobenil. At the urban sites, the herbicide 2,4-D was also frequently detected. For the Skagit-Samish sites, the herbicide bromacil and the neonicotinoid insecticide imidacloprid were also frequent detections.

For the lower Yakima sites, representing irrigated agriculture, the most commonly detected pesticides were the herbicide 2,4-D, followed by imidacloprid.

For the Wenatchee-Entiat sites, representing tree fruit agricultural, the most common detections were insecticide degradates, including the degradates for the legacy insecticide DDT and the endosulfan degradate.

Increases in imidacloprid detections were seen in 2010. This is likely due to increased sensitivity in the laboratory analysis because there were improvements in laboratory instrumentation during 2010.

Monitoring Program Changes

The following changes were made during the 2010 monitoring of the six basins across Washington State.

To improve data quality, Manchester Laboratory purchased a new instrument for carbamate analysis. The instrument enabled improvements to the identification criteria for all analytes in the carbamate analysis suite. All analytes now have at least one confirmation ion in addition to the quant ion. Sensitivity also improved with a drastic increase in the signal-to-noise ratio at the low end of the calibration curve. The maximum detection limit on the instrument demonstrates improved sensitivity for all analytes. These improvements have significantly reduced identification uncertainty, thus decreasing the potential for false positive and negatives.

Sampling for the following pesticides or degradates was added for all basins:

- Fenitrothion (fungicide).
- Phosmet oxygen analog (organophosphate phosmet degradate).
- Ronnel (organophosphate insecticide).

Planned Program Changes for 2011

Manchester Laboratory is planning to make the following changes during 2011:

1. Modify the carbamate analysis method used for this project. Eliminate the SE procedure and go to a DI method for the LCMS/MS carbamate analysis.

   In previous years, samples for the carbamate analysis underwent a sample extraction procedure before analysis. In 2010 the laboratory conducted a comparison study of two methods: the sample extraction (SE) procedure and a direct injection (DI) method before analysis. Results of the study showed that both methods were comparable (Weakland, 2011).

   Benefits of DI include higher spike recoveries for some analytes, closer to 100% recovery. During the SE process, there are losses which affect recovery rates. Another benefit of more consistent recoveries is less qualification of reported data and less rejected data.

   Changing to DI will also mean that solvents used during the SE process will no longer be needed. This will reduce the total solvent use at the laboratory, lessening toxic waste by-products.

2. Conduct a comparison study of DI versus the SE procedure for herbicide analysis.

   The laboratory will conduct a side-by-side study using both methods during the first four weeks of sampling. After the study is complete, the laboratory will compare results and make a recommendation about switching to DI for herbicide analysis.

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Appendices

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Appendix A. Glossary, Acronyms, and Abbreviations

Glossary

Analyte: Water quality constituent being measured (parameter).

Assessment criteria: Assessment criteria in this report are numeric criteria included in the EPA FIFRA Pesticide Registration Toxicity Criteria and endpoints; and the EPA National Recommended Water Quality Criteria (NRWQC).

Basin: Watershed. A drainage area in which all land and water areas drain or flow toward a central collector such as a stream, river, or lake at a lower elevation.

Bioaccumulation: Progressive increase in the amount of a substance in an organism or part of an organism which occurs because the rate of intake exceeds the organism's ability to remove the substance from the body.

Carbamate insecticide: N-methyl carbamate insecticides are similar to organophosphate insecticides in that they are nerve agents that inhibit cholinesterase enzymes. However they differ in action from the organophosphate compounds in that the inhibitory effect on cholinesterase is brief.

Clean Water Act: A federal act passed in 1972 that contains provisions to restore and maintain the quality of the nation's waters. Section 303(d) of the Clean Water Act establishes the TMDL program.

Conductivity: A measure of water's ability to conduct an electrical current. Conductivity is related to the concentration and charge of dissolved ions in water.

Degradate: Pesticide breakdown product.

Dissolved oxygen: A measure of the amount of oxygen dissolved in water.

Endosulfan: Endosulfan is an organochlorine insecticide that is registered for use on a number of agricultural commodities. In 2010, EPA signed an agreement with the registrants of endosulfan that will result in voluntary cancellation and phase out of all existing endosulfan uses in the United States. Under this agreement, all endosulfan uses will be phased out by July 2016. EPA is terminating uses of endosulfan to address its unacceptable risks to agricultural workers and wildlife (EPA, 2010).

Exceeded criteria: Did not meet criteria.

Grab sample: A discrete sample from a single point in the water column or sediment surface.

Herbicide: A substance used to kill plants or inhibit their growth.

Legacy pesticide: Banned pesticides no longer used but that persist in the environment.

Loading: The input of pollutants into a waterbody.

Marine water (seawater): Salt water.

Organophosphate pesticide: Organophosphate pesticides are derived from phosphoric acid and are highly neurotoxic, typically inhibiting cholinesterase.

Parameter: Water quality constituent being measured (analyte). A physical, chemical, or biological property whose values determine environmental characteristics or behavior.

Pesticide: A pesticide is any substance or mixture of substances intended for killing, repelling or mitigating any pest. Pests include nuisance microbes, plants, fungus, and animals.

pH: A measure of the acidity or alkalinity of water. A low pH value (0 to 7) indicates that an acidic condition is present, while a high pH (7 to 14) indicates a basic or alkaline condition. A pH of 7 is considered to be neutral. Since the pH scale is logarithmic, a water sample with a pH of 8 is ten times more basic than one with a pH of 7.

Point source: Sources of pollution that discharge at a specific location from pipes, outfalls, and conveyance channels to a surface water. Examples of point source discharges include municipal wastewater treatment plants, municipal stormwater systems, industrial waste treatment facilities, and construction sites that clear more than 5 acres of land.

Pollution: Contamination or other alteration of the physical, chemical, or biological properties of any waters of the state. This includes change in temperature, taste, color, turbidity, or odor of the waters. It also includes discharge of any liquid, gaseous, solid, radioactive, or other substance into any waters of the state. This definition assumes that these changes will, or are likely to, create a nuisance or render such waters harmful, detrimental, or injurious to (1) public health, safety, or welfare, or (2) domestic, commercial, industrial, agricultural, recreational, or other legitimate beneficial uses, or (3) livestock, wild animals, birds, fish, or other aquatic life.

Salmonid: Any fish that belong to the family *Salmonidae*. Basically, any species of salmon, trout, or char. www.fws.gov/le/ImpExp/FactSheetSalmonids.htm

Suspended sediment: Solid fragmented material (soil and organic matter) in the water column.

Synergistic: A synergistic effect occurs when the combined effects of two chemicals are greater than the predicted sum of each chemical's effects.

Thermistor: An electronic device that uses semiconductors to measure temperature. A data logger.

Total Maximum Daily Load (TMDL): Water cleanup plan. A distribution of a substance in a waterbody designed to protect it from exceeding water quality standards. A TMDL is equal to the sum of all of the following: (1) individual wasteload allocations for point sources, (2) the load allocations for nonpoint sources, (3) the contribution of natural sources, and (4) a Margin of Safety to allow for uncertainty in the wasteload determination. A reserve for future growth is also generally provided.

Total suspended solids (TSS): The suspended particulate matter in a water sample as retained by a filter.

Water quality standards: Washington State water quality standards.

Watershed: Basin. A drainage area in which all land and water areas drain or flow toward a central collector such as a stream, river, or lake at a lower elevation.

303(d) list: Section 303(d) of the federal Clean Water Act requires Washington State periodically to prepare a list of all surface waters in the state for which beneficial uses of the water – such as for drinking, recreation, aquatic habitat, and industrial use – are impaired by pollutants. These are water quality limited estuaries, lakes, and streams that fall short of Washington State surface water quality standards and are not expected to improve within the next two years.

7-DADMax or 7-day average of the daily maximum temperatures: The arithmetic average of seven consecutive measures of daily maximum temperatures. The 7-DADMax for any individual day is calculated by averaging that day's daily maximum temperature with the daily maximum temperatures of the three days prior and the three days after that date.

Acronyms and Abbreviations

DDD	Dichloro-diphenyl-dichloroethane
DDE	Dichloro-diphenyl-dichloroethylene
DDT	Dichloro-diphenyl-trichloroethane
Ecology	Washington State Department of Ecology
EIM	Environmental Information Management (Ecology)
EPA	United States Environmental Protection Agency
ESLOC	Endangered species level of concern (EPA)
FIFRA	Federal Insecticide Fungicide and Rodenticide Act
GCMS	Gas chromatograph coupled with mass spectrometer
LC_{50}	Lethal concentration to cause mortality in 50% of test species
LCMS	Liquid chromatograph coupled with mass spectrometer
LCS	Laboratory control sample
LOC	Level of concern
LPQL	Lower practical quantitation limit
MEL	Manchester Environmental Laboratory
MS	Mass spectrometer
MS/MSD	Matrix spike/matrix spike duplicate
n	Number
NAD	North American Datum
NRWQC	National Recommended Water Quality Criteria (EPA)
NIST	National Institute of Standards and Technology
NOAA	National Oceanic and Atmospheric Administration
NOEC	No observable effect concentration
QA	Quality assurance
QC	Quality control
RQ	Risk quotient
RPD	Relative percent difference
RSD	Relative standard deviation

SOP	Standard operation procedures
TMDL	(See Glossary above)
TSS	(See Glossary above)
TSU	Toxics Studies Unit
USFWS	United States Fish and Wildlife Service
USGS	United States Geological Survey
WAC	Washington Administrative Code
WRIA	Water Resource Inventory Area
WSDA	Washington State Department of Agriculture
7-DADMax	(See Glossary above)

Units of Measurement

°C	degrees centigrade
cfs	cubic feet per second
m	meter
mg	milligrams
mg/L	milligrams per liter (parts per million)
s.u.	standard units
umhos/cm	micromhos per centimeter

Appendix B. Monitoring Sites and Duration of Sampling

Table B-1. Station locations, duration of monitoring, and site location descriptions for 2010.

Site	Duration	Latitude	Longitude	Location Description
Cedar-Sammamish Watershed				
TC-3	Mar 8– Sept 8	47.6958	122.2757	Downstream of pedestrian footbridge near Mathews Beach Park.
Green-Duwamish Watershed				
LC-1	Mar 8– Sept 8	47.5625	122.367	Upstream of the culvert under the 12th Fairway on the West Seattle Golf Course.
Skagit-Samish Watershed				
BD-1	Mar 8 – Sept 8	48.3086	122.3473	Upstream side of bridge at Milltown Road.
BD-2	Mar 8 – Sept 8	48.3887	122.3329	Upstream side of bridge at Lenor Lane.
BS-1	Mar 8 – Sept 8	48.3406	122.4140	Downstream of tidegate on Fir Island Road.
IS-1	Mar 8 – Sept 8	48.4506	122.4651	Inside upstream side of tidegate at Bayview-Edison Road.
SR-1	Mar 8 – Sept 8	48.5209	122.4113	Upstream side of bridge at Thomas Road.
Lower Yakima Watershed				
MA-2	Mar 10 – Oct 26	46.3306	120.1989	Approximately 15 meters upstream of bridge at Indian Church Rd.
SP-2	Mar 10 – Sept 8	46.2583	119.7101	Downstream side of culvert on McCready Road.
SP-3	Mar 10 – Sept 8	46.2344	119.6845	Approximately 3 meters downstream of Chandler Canal overpass.
SU-1	Mar 10 – Sept 8	46.2509	120.0202	Downstream side of bridge at Holaday Road.
Wenatchee Watershed				
WE-1	Mar 9 – Sept 8	47.4721	120.3710	Upstream side of Sleepy Hollow bridge.
MI-1	Mar 9 – Sept 8	47.4893	120.4815	Above Woodring Canyon Road and Mission Creek Road.
PE-1	Mar 9 – Sept 8	47.5570	120.5825	Approximately 30 meters downstream of bridge at Saunders Road.
BR-1	Mar 9 – Sept 8	47.5211	120.4862	Upstream side of culvert at Evergreen Drive.
Entiat Watershed				
EN-1	Mar 9 – Sept 8	47.6633	120.2506	Upstream side of bridge at Keystone Road.

Datum in NAD 83.

Appendices C - E

Surface Water Monitoring Program for Pesticides in Salmonid-Bearing Streams, 2010

Publication No. 11-03-021-Appendices C-E

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Appendix C. Quality Assurance

Laboratory Data Quality

Data may be qualified if one or more analytical factors affect confidence in the prescribed data value. Manchester Environmental Laboratory (MEL) qualifies data according to the National Functional Guidelines for Organic Data Review (EPA, 1999, 2007). Definitions of data qualifiers are presented in Table C-1.

Table C-1. Data qualification.

Qualifier	Definition
(No qualifier)	The analyte was detected at the reported concentration. Data are not qualified.
E	Reported result is an estimate because it exceeds the calibration range.
J	The analyte was positively identified; the associated numerical value is the approximate concentration of the analyte in the sample.
NJ	The analysis indicates the presence of an analyte that has been "tentatively identified," and the associated numerical value represents its approximate concentration.
NAF	Not analyzed for.
NC	Not calculated.
REJ	The sample results are rejected due to serious deficiencies in the ability to analyze the sample and meet quality control criteria. The presence or absence of the analyte cannot be verified.
U	The analyte was not detected at or above the reported sample quantitation limit.
UJ	The analyte was not detected at or above the reported sample quantitation limit. However, the reported quantitation limit is approximate and may or may not represent the actual limit of quantitation necessary to accurately measure the analyte in the sample.

MEL, 2000, 2008; EPA, 1999, 2007.

Performance measures for quality assurance (QA) and quality control (QC) are presented in Table C-2. Lowest concentrations of interest for surface water grab samples are below reporting limits. Detections quantified below reporting limits are qualified as estimates.

Table C-2. Performance measures for quality assurance and quality control.

Analysis Method[1]	Analysis[2]	Field/Lab Replicates, MS/MSD[3], and Lab Control Samples	MS/MSD[3], Surrogates, and Lab Control Samples
		RPD[4]	% Recovery
GCMS	Pesticide-C-l	±40	30-130
	Pesticide-N	±40	30-130
	Pesticide-OP	±40	30-130
	Pesticide-Py	±40	30-130
GCMS-H	Herbicides	±50	40-130
LCMS/MS	Pesticide-C	±40	50-150
TSS	TSS	±20	80-120

[1] GCMS: Gas chromatography/mass spectroscopy, EPA method (modified) SW 846 3535M/8270M.
GCMS-H: Derivitizable acid herbicides by GCMS, EPA method (modified) SW 846 3535M/8270M.
LCMS/MS: Liquid chromatography/mass spectroscopy, EPA method (modified) SW 846 3535M/8321AM.
TSS: Total suspended solids, EPA method 2540D.
[2]C-l: chlorinated, N: nitrogen containing, OP: organophosphorus, Py: pyrethroid, C: carbamate.
[3]MS/MSD: Matrix spike and matrix spike duplicate.
[4]RPD: Relative percent difference.

Lower Practical Quantitation Limits

Lower practical quantitation limits (LPQLs) are the limits at which laboratories may report data without classifying the concentration as an estimate below the lowest calibration standard. The LPQL is determined by averaging the lower reporting values, per analyte, for all batches over each study period. LPQL data are presented in Table C-3.

Table C-3. Mean performance lower practical quantitation limits (LPQL) (ug/L).

Chemical	[1]Use	Parent	[2]Analysis Method	LPQL[3]		
				2008	2009	2010
1-Naphthol	D-C		LCMS\MS	0.053	0.050	0.049
2,3,4,5-Tetrachlorophenol	D-M		GCMS-H	0.063	0.063	0.063
2,3,4,6-Tetrachlorophenol	D-M		GCMS-H	0.063	0.063	0.063
2,4,5-T	H		GCMS-H	0.063	0.063	0.063
2,4,5-TP	H		GCMS-H	0.063	0.063	0.063
2,4,5-Trichlorophenol	D-M		GCMS-H	0.063	0.063	0.063
2,4,6-Trichlorophenol	D-M		GCMS-H	0.063	0.063	0.063
2,4-D	H		GCMS-H	0.063	0.063	0.063
2,4-DB	H		GCMS-H	0.063	0.063	0.063
2,4'-DDD	D-OC	DDT	GCMS	0.033	0.033	0.033
2,4'-DDE	D-OC	DDT	GCMS	0.033	0.033	0.033
2,4'-DDT	D-OC	DDT	GCMS	0.033	0.033	0.033
3,5-Dichlorobenzoic Acid	D-M		GCMS-H	0.063	0.063	0.063
3-Hydroxycarbofuran	D-C	Carbofuran	LCMS\MS	0.050	0.050	0.049
4,4'-DDD	D-OC	DDT	GCMS	0.033	0.034	0.033
4,4'-DDE	D-OC	DDT	GCMS	0.033	0.034	0.033
4,4'-DDT	I-OC		GCMS	0.033	0.034	0.033

Chemical	[1]Use	Parent	[2]Analysis Method	LPQL[3]		
				2008	2009	2010
4,4'-Dichlorobenzophenone	D		GCMS		0.101	0.100
4-Nitrophenol	D-H		GCMS-H	0.063	0.063	0.063
Acetochlor	H		GCMS		0.101	0.100
Acifluorfen	H		GCMS-H	0.063	0.063	0.063
Alachlor	H		GCMS	0.033	0.033	0.033
Aldicarb	I-C		LCMS\MS	0.100	0.100	0.096
Aldicarb Sulfone	D-C	Aldicarb	LCMS\MS	0.050	0.053	0.049
Aldicarb Sulfoxide	D-C	Aldicarb	LCMS\MS	0.020	0.054	0.020
Aldrin	I-OC		GCMS	0.033	0.033	0.033
Alpha-BHC	I-OC		GCMS	0.033	0.033	0.033
Atrazine	H		GCMS	0.033	0.034	0.033
Azinphos Ethyl	I-OP		GCMS	0.033	0.033	0.033
Azinphos Methyl	I-OP		GCMS	0.033	0.050	0.043
Benefin	H		GCMS	0.033	0.033	0.033
Bensulide	H		GCMS	0.033		
Bentazon	H		GCMS-H	0.063	0.063	0.063
Benthiocarb	H-C		GCMS	0.100	0.101	0.100
Beta-BHC	I-OC		GCMS	0.033	0.033	0.033
beta-Cypermethrin 65731-84-2 ([(1R)-1a(S*), 3a] isomer)	I-Py		GCMS		0.101	0.100
Bifenthrin	I-Py		GCMS		0.101	0.100
Bromacil	H		GCMS	0.033	0.033	0.033
Bromoxynil	H		GCMS-H	0.063	0.063	0.063
Butachlor	H		GCMS		0.304	0.303
Butylate	H		GCMS	0.033	0.033	0.033
Captan	F		GCMS	0.033	0.033	0.033
Carbaryl	I-C		LCMS/MS	0.020	0.020	0.020
Carbofuran	I-C		LCMS/MS	0.020	0.020	0.020
Carboxin	F		GCMS	0.034	0.044	0.051
Chlorothalonil	F		GCMS	0.033	0.033	0.033
Chlorpropham	H		GCMS	0.033	0.033	0.033
Chlorpyrifos	I-OP		GCMS	0.033	0.034	0.033
Chlorpyrifos O.A.	D-OP		GCMS		0.101	0.100
Cis-Chlordane	I-OC		GCMS	0.033	0.033	0.033
Cis-Nonachlor	I-OC		GCMS	0.033	0.051	0.051
Cis-Permethrin	I-Py		GCMS	0.050	0.051	0.051
Clopyralid	H		GCMS-H	0.063	0.063	0.063
Coumaphos	I-OP		GCMS	0.033	0.051	0.051
Cyanazine	H		GCMS	0.033	0.033	0.033
Cycloate	H		GCMS	0.033	0.033	0.033
DCPA	H		GCMS-H	0.063	0.063	0.063
DDVP	I-OP		GCMS	0.050	0.051	0.051
Delta-BHC	I-OC		GCMS	0.033	0.033	0.033
Deltamethrin	I-Py		GCMS	0.100	0.101	0.100
Diallate	H		GCMS	0.033	0.033	0.033

Chemical	[1]Use	Parent	[2]Analysis Method	LPQL[3]		
				2008	2009	2010
Diazinon	I-OP		GCMS	0.033	0.033	0.033
Diazoxon	D-OP	Diazinon	GCMS		0.101	0.100
Dicamba I	H		GCMS-H	0.063	0.063	0.063
Dichlobenil	H		GCMS	0.033	0.033	0.033
Dichlorprop	H		GCMS-H	0.063	0.063	0.063
Diclofop-Methyl	H		GCMS-H	0.063	0.063	0.063
Dieldrin	I-OC		GCMS	0.050	0.051	0.051
Dimethoate	I-OP		GCMS	0.033	0.033	0.033
Dinoseb	H		GCMS-H	0.063	0.063	0.063
Diphenamid	H		GCMS	0.033	0.033	0.033
Disulfoton	I-OP		GCMS	0.052	0.112	0.065
Disulfoton sulfone	I-OP		GCMS	0.100	0.101	0.100
Disulfoton sulfoxide	D-OP		GCMS		0.135	0.100
Diuron	H		GCMS	0.050	0.058	0.051
Endosulfan I	I-OC		GCMS	0.050	0.051	0.051
Endosulfan II	I-OC		GCMS	0.050	0.051	0.051
Endosulfan Sulfate	D-OC	Endosulfan	GCMS	0.033	0.034	0.033
Endrin	I-OC		GCMS	0.050	0.051	0.051
Endrin Aldehyde	D-OC	Endrin	GCMS	0.050	0.051	0.051
Endrin Ketone	D-OC	Endrin	GCMS	0.033	0.033	0.033
EPN	I-OP		GCMS	0.033	0.033	0.033
Eptam	H		GCMS	0.033	0.033	0.033
Ethalfluralin	H		GCMS	0.033	0.033	0.036
Ethion	I-OP		GCMS	0.033	0.033	0.033
Ethoprop	I-OP		GCMS	0.033	0.033	0.033
Fenamiphos	I-OP		GCMS	0.033	0.038	0.042
Fenamiphos Sulfone	D-OP		GCMS		0.101	0.100
Fenarimol	F		GCMS	0.033	0.033	0.033
Fenitrothion	I-OP		GCMS			0.050
Fensulfothion	I-OP		GCMS	0.033	0.033	0.033
Fenthion	I-OP		GCMS	0.048	0.033	0.033
Fenvalerate (2 isomers)	I-Py		GCMS	0.033	0.033	0.038
Fipronil	I-Pyra		GCMS		0.101	0.100
Fipronil Disulfinyl	D-Pyra		GCMS		0.101	0.100
Fipronil Sulfide	D-Pyra		GCMS		0.101	0.100
Fipronil Sulfone	D-Pyra		GCMS		0.101	0.100
Fluridone	H		GCMS	0.100	0.101	0.100
Fonofos	I-OP		GCMS	0.033	0.033	0.033
Heptachlor	I-OC		GCMS	0.033	0.033	0.033
Heptachlor Epoxide	D-OC	Heptachlor	GCMS	0.033	0.033	0.033
Hexachlorobenzene	F		GCMS	0.034	0.033	0.033
Hexazinone	H		GCMS	0.050	0.051	0.051
Imidacloprid	I-N		LCMS\MS	0.020	0.020	0.020
Imidan	I-OP		GCMS	0.033	0.068	0.038
Ioxynil	H		GCMS-H	0.063	0.063	0.063

Chemical	[1]Use	Parent	[2]Analysis Method	LPQL[3]		
				2008	2009	2010
Kelthane	I-OC		GCMS	0.314	0.304	0.303
lambda-Cyhalothrin	I-Py		GCMS		0.101	0.100
Lindane	I-OC		GCMS	0.033	0.033	0.033
Linuron	H		GCMS	0.050	0.051	0.051
Malathion	I-OP		GCMS	0.033	0.033	0.033
MCPA	H		GCMS-H	0.063	0.063	0.063
MCPP	H		GCMS-H	0.063	0.063	0.063
Metalaxyl	F		GCMS	0.033	0.033	0.033
Methidathion	I-OP		GCMS	0.293	0.304	0.303
Methiocarb	I-C		LCMS\MS	0.020	0.021	0.020
Methomyl	I-C		LCMS\MS	0.050	0.050	0.049
Methomyl oxime	D-C	Thiodicarb	LCMS\MS	0.020	0.020	0.020
Methoxychlor	I-OC		GCMS	0.033	0.051	0.051
Methyl Chlorpyrifos	I-OP		GCMS	0.033	0.033	0.033
Methyl Paraoxon	D-OP	Methyl parathion	GCMS	0.100	0.101	0.100
Methyl Parathion	I-OP		GCMS	0.033	0.033	0.033
Metolachlor	H		GCMS	0.033	0.033	0.033
Metribuzin	H		GCMS	0.033	0.033	0.033
Mevinphos	I-OP		GCMS	0.050	0.051	0.051
MGK-264	Sy		GCMS	0.033	0.051	0.051
Mirex	I-OC		GCMS	0.033	0.035	0.033
Monocrotophos	I-OP		GCMS	0.050	0.051	0.051
Naled	I-OP		GCMS	0.059	0.035	0.034
Napropamide	H		GCMS	0.050	0.051	0.051
Norflurazon	H		GCMS	0.033	0.034	0.033
Oryzalin	H		GCMS	0.100	0.114	0.133
Oxamyl	I-C		LCMS\MS	0.050	0.052	0.049
Oxamyl oxime	D-C	Oxamyl	LCMS\MS	0.020	0.020	0.020
Oxychlordane	D-OC	Chlordane	GCMS	0.033	0.033	0.033
Oxyfluorfen	H		GCMS	0.033	0.101	0.100
Parathion	I-OP		GCMS	0.033	0.033	0.033
Pebulate	H		GCMS	0.033	0.033	0.033
Pendimethalin	H		GCMS	0.033	0.034	0.033
Pentachlorophenol	WP		GCMS-H	0.063	0.063	0.063
Phenothrin	I-Py		GCMS	0.033	0.033	0.033
Phorate	I-OP		GCMS	0.299	0.291	0.303
Phorate O.A.	D-OP		GCMS		0.193	0.137
Phosmet O.A.	D-OP		GCMS			0.100
Picloram	H		GCMS-H	0.063	0.063	0.063
Piperonyl Butoxide	Sy		GCMS		0.101	0.100
Promecarb	I-C		LCMS\MS	0.020	0.020	0.020
Prometon	H		GCMS	0.033	0.033	0.033
Prometryn	H		GCMS	0.033	0.033	0.033
Pronamide	H		GCMS	0.033	0.033	0.033
Propachlor	H		GCMS	0.033	0.033	0.033

Chemical	[1]Use	Parent	[2]Analysis Method	LPQL[3]		
				2008	2009	2010
Propargite	I-SE		GCMS	0.033	0.051	0.051
Propazine	H		GCMS	0.033	0.033	0.033
Propoxur	I-C		LCMS\MS	0.050	0.050	0.049
Prothiofos	I-OP		GCMS		0.101	0.100
Resmethrin	I-Py		GCMS	0.050	0.036	0.033
Ronnel	I-OP		GCMS			0.050
Simazine	H		GCMS	0.033	0.033	0.033
Simetryn	H		GCMS	0.100	0.101	0.100
Sulfotepp	I-OP		GCMS	0.033	0.033	0.033
Sulprofos	I-OP		GCMS	0.033		0.050
Tebuthiuron	H		GCMS	0.033	0.033	0.033
Terbacil	H		GCMS	0.033	0.034	0.033
Tetrachlorvinphos	I-OP		GCMS	0.050	0.051	0.051
Thiodicarb	I-C		LCMS	0.020		
Tokuthion	I-OP		GCMS	0.050		
Total Suspended Solids			TSS		1.059	2.000
Tralomethrin	I-Py		GCMS	0.100	0.101	0.100
Trans-Chlordane	I-OP		GCMS	0.033	0.033	0.033
Trans-Nonachlor	I-OC		GCMS	0.033	0.051	0.051
trans-Permethrin	I-Py		GCMS		0.101	0.100
Triadimefon	F		GCMS	0.033	0.033	0.033
Triallate	H		GCMS	0.033	0.033	0.033
Trichloronat	I-OP		GCMS	0.050	0.051	0.051
Triclopyr	H		GCMS-H	0.063	0.063	0.063
Tricyclazole	F		GCMS		0.101	0.100
Trifluralin	H		GCMS	0.033	0.034	0.033

[1] C: Carbamate, D: Degradate, F: Fungicide, I: Insecticide, H: Herbicide, OC: Organochlorine, OP: Organophosphorus, Py: Pyrethroid, SE: Sulfite Ester, Sy: Synergist, WP: Wood Preservative.
[2] GCMS: Gas chromatography/mass spectroscopy, EPA method (modified) SW 846 3535M/8270M.
GCMS-H: Derivitizable acid herbicides by GCMS, EPA method (modified) SW 846 3535M/8270M.
LCMS\MS: Liquid chromatography/mass spectroscopy, EPA method (modified) SW 846 3535M/8321AM.
[3]Blank cells indicate no analysis for the compound in that year.

Quality Assurance Samples

QA samples were collected each year to assure consistency and accuracy of sample analysis.

For this project, QA samples included field replicates, field blanks, and matrix spike and matrix spike duplicates (MS/MSD). QA samples for the laboratory included split sample duplicates, laboratory control samples, surrogate spikes, and method blanks.

In 2010, 13.4% of the field samples obtained were for QA. In 2010 QA samples included 32 field replicates for carbamates, herbicides, and pesticide GCMS; and 33 replicates for TSS. QA also included 16 field blanks for TSS, herbicides, and pesticide GCMS; and 18 blanks for carbamates. There were also 16 MS/MSD samples for carbamates, herbicides, and pesticide GCMS.

Results for each QA sample method are outlined in the sections below.

Field Replicates

Results for pesticide field replicates are presented in Tables C-4 and C-5. Table C-4 presents the data value, data qualification (if assigned), and relative percent difference (RPD) between the results for compounds which were consistently identified in both the grab sample and replicate.

Table C-4. Pooled average %RPD of consistent field replicate pairs by analysis type.

Analysis	Pooled Average %RPD	Number of Replicate Pairs
Herbicides	10.5%	20
Carbamates	3.3%	16
Pesticide GCMS	9.3%	41
TSS	12.4%	33

Consistent identification refers to compounds which were identified in both the original sample and field replicate. Inconsistently identified replicate pairs are those in which the compound was identified in one sample but not the other. Inconsistently identified grab sample replicates are presented in Table C-5.

During 2010 field replicate sampling frequency for pesticides was 7.6% and for TSS was 7.7%. Precision between replicate pairs was calculated using percent RPD (%RPD). The percent (%RSD) is calculated by dividing the absolute value of the difference between the replicates by their mean, then multiplying by 100 for a percent value.

Excluding total suspended solids (TSS), there were 77 consistently identified analytes and 19 inconsistently identified analytes detected in 97 replicate pairs. The average %RPD for each of the analytical methods was excellent (Table C-4). Of the consistently identified replicate pairs only two of the 77 pairs exceeded the 40% RPD criterion (Table C-5). One of these replicate pairs (dichlobenil) had a %RPD of 100. This difference is likely because the results were very low and the RPD statistic has limited effectiveness in assessing variability at low levels (Mathieu, 2006).

Of the inconsistently identified replicate pairs, five of the 19 pairs exceeded the 40% RPD criterion (Table C-5). For the inconsistently identified replicate pairs, higher %RPD occurred because a tentative detection or estimated detection was compared to a less than reporting limit value. The tentative detections and estimated detections were all less than the reporting limit, and data for these replicates are of acceptable data quality.

TSS was consistently detected in 33 replicate pairs. The pooled %RPD of all replicates was 12.4%. A total of 81% of the replicates were within the 20% RPD criterion.

Table C-5. Detected pairs within field replicate results (µg/L).

Parameter	Sample	Q	Replicate	Q	RPD
2,4-D	0.041	J	0.041	J	0.0
	0.180		0.320		56.0
	0.040	J	0.039	J	2.5
	0.040	J	0.054	J	29.8
	0.027	J	0.026	J	3.8
	0.240	J	0.230	J	4.3
	0.038	J	0.045	J	16.9
	0.023	J	0.024	J	4.3
				Mean =	**14.7**
4,4'-DDE	0.038		0.037		**2.7**
4,4'-DDT	0.016	J	0.017	J	**6.1**
Bentazon	0.045	J	0.049	J	8.5
	0.063		0.082		26.2
				Mean =	**17.4**
Bromacil	0.036		0.038		5.4
	0.190	J	0.220	J	14.6
	0.070		0.066		5.9
	0.032	J	0.036		11.8
				Mean =	**9.4**
Carbaryl	0.015	J	0.014	J	**6.9**
Carbofuran	0.005	J	0.005	J	0.0
	0.100		0.094		6.2
	0.004	J	0.004	J	0.0
	0.006	J	0.006	J	0.0
				Mean=	**1.5**
Chlorpropham	0.025	J	0.023	J	**8.3**
DCPA	0.063	J	0.072		**13.3**
DDVP	0.070		0.067		**4.4**
Diazinon	0.120		0.120		**0.0**
Dicamba I	0.019	J	0.019	J	0.0
	0.014	J	0.013	J	7.4
	0.150		0.150		0.0
				Mean=	**2.5**
Dichlobenil	0.015	J	0.013	J	14.3
	0.010	J	0.010	J	0.0
	0.078		0.080	J	2.5
	0.006	J	0.006	J	0.0

Parameter	Sample	Q	Replicate	Q	RPD
	0.008	J	0.009	J	11.8
	0.037		0.037		0.0
	0.012	J	0.011	J	8.7
	0.015	J	0.015	J	0.0
	0.001	J	0.003	J	100.0
	0.011	J	0.013	J	16.7
				Mean=	**15.4**
Disulfoton sulfoxide	0.021	J	0.031	J	38.5
Diuron	0.110	J	0.120	J	8.7
	0.017	J	0.015	J	12.5
	3.6	E	3.6	E	0.0
				Mean=	**7.1**
Endosulfan Sulfate	0.036		0.038		**5.4**
Eptam	0.200		0.220		9.5
	0.027	J	0.028	J	3.6
	0.030	J	0.029	J	3.4
	0.063		0.074		16.1
				Mean=	**8.2**
Ethoprop	0.280		0.300		**6.9**
Imidacloprid	0.362		0.411		12.7
	0.005	J	0.005	J	0.0
	0.024		0.024		0.0
	0.020		0.019	J	5.1
	0.007	J	0.007	J	0.0
	0.005	J	0.005	J	0.0
	0.004	J	0.004	J	0.0
	0.005	J	0.005	J	0.0
	0.924		0.833		10.4
	0.009	J	0.008	J	11.8
				Mean=	**4**
MCPA	0.061	J	0.059	J	**3.3**
Methomyl	0.004	J	0.004	J	**0.0**
Metolachlor	0.054	J	0.056		3.6
	0.039		0.044		12.0
	0.008	J	0.008	J	0.0
	0.190		0.200		5.1
				Mean=	**5.2**
Metribuzin	0.210		0.210		**0.0**
Napropamide	0.480		0.400		**18.2**

Parameter	Sample	Q	Replicate	Q	RPD
Pendimethalin	0.076		0.074		**2.7**
Pentachlorophenol	0.016	J	0.016	J	**0.0**
Terbacil	0.098		0.095		3.1
	0.035	J	0.036	J	2.8
	0.090		0.100		10.5
	0.500		0.510		2.0
				Mean=	**4.6**
Triclopyr	0.160		0.190		17.1
	0.042	J	0.043	J	2.4
	0.089		0.083		7.0
	0.030	J	0.032	J	6.5
				Mean=	**8.2**
Trifluralin	0.022	J	0.023	J	**4.4**

Inconsistent replicate detections are an indicator of sampling uncertainty. Table C-6 compares inconsistent replicate detections to the LPQL for non-detections in the paired replicate. Most inconsistent detections were found at concentrations near or below the LPQL.

Table C-6. Inconsistent field replicate detections compared to the lower practical quantitation limit (LPQL) (µg/L).

Parameter	Sample		Replicate		RPD
Bentazon	0.052	J	0.044	NJ	16.7
Bromacil	0.037		0.034	U	8.5
Carbaryl	0.020	UJ	0.016	J	22.2
	0.020	U	0.005	J	120
			Mean=		71.1
Carbofuran	0.006	J	0.020	U	108
Dicamba I	0.015	J	0.016	NJ	6.5
	0.011	J	0.011	NJ	0.0
			Mean=		3.2
Diuron	0.230	J	0.200	NJ	14.0
	0.150		0.170	NJ	12.5
	0.025	J	0.028	NJ	11.3
	0.067	J	0.059	NJ	12.7
	Mean=				12.6
Imidacloprid	0.005	J	0.020	U	120
	0.005	J	0.020	U	120
			Mean=		120
MCPA	0.023	NJ	0.024	J	4.3
Pentachlorophenol	0.015	NJ	0.015	J	0.0
	0.021	J	0.068	U	106
	0.020	J	0.021	NJ	4.9
			Mean=		36.8
Triclopyr	0.035	J	0.035	NJ	0.0
Trifluralin	0.015	J	0.034	U	77.6

Laboratory Duplicates

MEL used laboratory split sample duplicates to ensure consistency of TSS analyses. In 2010 there were 32 laboratory replicate pairs. The pooled average RPD was 12.8%, the maximum RPD was 29%. Six out of 32 replicate pairs exceeded the 20% RPD criteria. For these replicates, results were low, and the RPD statistic has limited effectiveness in assessing variability at low levels (Mathieu, 2006).

Field Blanks

Field blank detections indicate the potential for sample contamination in the field and laboratory and the potential for false detections due to analytical error.

In 2010 there were no field blank detections for the pesticide analysis. There was one TSS field blank detection of 3 mg/L on July 20, 2010 for the Samish River site. The reporting limit for TSS was 1 mg/L. All TSS values analyzed that day (July 20, 2010) that are less than 9 mg/L will be qualified as estimates.

Laboratory Blanks

MEL uses laboratory blanks to assess the precision of equipment and the potential for internal laboratory contamination. If lab blank detections occur, the sample LPQL may be increased, and detections may be qualified as estimates.

Laboratory blank detections for 2010 are presented in Table C-7. For all lab blank detections, any analytes found in associated samples below 5 times the lab blank detection were reported at the level detected, but qualified as not detected at an estimated detection limit (UJ).

Table C-7. Laboratory blank detections (µg/L).

Analysis	Chemical	Analysis Date	Value
GCMS	2,4'-DDT	06/11/2010	0.015 J
	4,4'-DDD	06/11/2010	0.012 J
	4,4'-DDE	06/11/2010	0.007 J
	4,4'-DDT	06/11/2010	0.018 J
	cis-Chlordane	06/11/2010	0.002 J
	Mirex	06/11/2010	0.012 J
	Trans-Chlordane	06/11/2010	0.002 J
LCMS\MS	Imidacloprid	4/14/2010	0.001 J
		9/28/2010	0.002 J
	Carbaryl	6/11/2010	0.003 J
		7/23/2010	0.004 J

Surrogates

Surrogates are compounds that are spiked into field samples at the laboratory. They are used to check recovery for a group of compounds. For instance, triphenyl phosphate is a surrogate for organophosphorus insecticides (Table C-8).

In 2010 MEL discontinued use of 4,4'-DDE-d8 and gamma-BHC-d6 as surrogates for the pesticide GCMS analysis. MEL could no longer purchase these standards from any supplier. The 4,4'-DDE-d8 standard was replaced with a carbon 13 labeled version, 4,4'-DDE-12C13. Atrazine-D5 and triflurin-D14 labeled surrogates were also added to support pesticide GCMS chemistries.

High pesticide surrogate recovery requires related detections to be qualified as estimates. Low pesticide surrogate recovery requires all related data to be qualified as estimates.

Table C-8. Pesticide surrogates.

Surrogate Compound	Surrogate for...
2,4,6-tribromophenol	Acid-derivitizable herbicides
2,4-dichlorophenylacetic acid	
Carbaryl C13	Carbamate pesticides
4,4'-DDE-13C12	Chlorinated pesticides
Decachlorobiphenyl (DCB)	
Atrazine-D5	Chlorinated and nitrogen pesticides
1,3-dimethyl-2-nitrobenzene	Nitrogen pesticides
Trifluralin-D-14	
Chlorpyrifos-d10	Organophosphorus pesticides
Triphenyl phosphate	

The majority of surrogate recoveries fell within the QC limits established by MEL for all compounds except for the compounds and dates described in Table C-9.

Table C-9. Surrogate compounds that did not meet data quality control limits.

Surrogate Compound	Surrogate for	Result Date	Percent Recovery
Atrazine-D5	Chlorinated and nitrogen pesticides	04/23/2010	161%
Chlorpyrifos-d10	Organophosphorus pesticides	04/23/2010	139%
Triphenyl phosphate	Organophosphorus pesticides	04/23/2010	142%
Carbaryl C13	Carbamate pesticides	05/08/2010	39%
1,3-Dimethyl-2-nitrobenzene	Nitrogen pesticides	05/11/2010	0%

On April 23 surrogate recoveries were high for the pesticide GCMS analysis (Table C-9). All positive sample results for pesticide GCMS sampled on April 12 and 13 were qualified as estimated and may be biased high. For the May 8, carbamate analysis, the method blank surrogate recovery was low; the method blank results were qualified as estimates. For the May 11 pesticide GCMS analysis, there were low recoveries for nitrogen pesticide surrogates; affected samples were qualified as estimates.

Matrix Spike/Matrix Spike Duplicates (MS/MSD)

MS/MSD results reflect the process of sample duplication (field), analyte degradation, matrix interaction (sample/standard), extraction efficiency, and analyte recovery. This measure is the best overall indicator of accuracy and reproducibility of the entire sampling process.

Table C-10 presents the mean, minimum, and maximum percent recovery for the MS/MSD for the three types of analysis as well as the RPD between for the MS/MSD for 2010.

In 2010, the average recovery for all three analyses and the average RPD was good, showing acceptable performance for most compounds.

Table C-10. Mean, minimum, and maximum percent recovery for MS/MSD and MS/MSD RPD.

Analysis	MS\MSD Recovery			%RPD for MS\MSD		
	Mean	Minimum	Maximum	Mean	Minimum	Maximum
LCMS\MS	74%	36%	99.4%	10%	5%	35%
GCMS-Herbicides	82%	24%	299%	9%	5%	19%
GCMS-Pesticides	108%	5%	238%	7%	2%	31%

The percent recoveries for the LCMS\MS analysis were excellent, within the target range of 50-150% recovery, except for methomyl oxime which had low recoveries (34-37%). Methomyl oxime was not detected at any site in 2010.

The percent recoveries for the GCMS-Herbicide analysis were also good, with most analyte recoveries between the acceptable range of 40-130%. Acifluorfen had high recoveries (298-300%). Acifluorfen was not detected in any of the 2010 samples. Picloram had low recoveries (24%). The upstream Big Ditch site had two detections of picloram; both detections were qualified as estimates due to low recoveries.

The percent recoveries for the GCMS-Pesticides included many analytes, and generally were good. Percent recoveries for resmethrin were low (4-5%). Resmethrin was not detected in any 2010 samples.

Fifteen analytes exceeded the acceptable range of 30-130% recoveries. Table C-11 presents the MS/MSD average percent recoveries for these compounds. Chlorothalonil, terbacil, ethoprop, and diuron were the only compounds detected during 2010. Detections of these compounds were qualified as estimates if MS/MSD recoveries were low for that day.

Table C-11. Analytes that exceeded the % MS/MSD recovery range of > 130%.

Analyte	MS/MSD Average % Recovery	Number of Detections
Tetrachlorvinphos	139	0
Ethalfluralin	140	0
Chlorothalonil	143	1
Fenarimol	147	0
Terbacil	150	29
Cyanazine	150	0
Ethoprop	151	5
Fenamiphos	158	0
Diuron	160	62
Methyl Paraoxon	160	0
Fluridone	166	0
Azinphos Ethyl	178	0
Mevinphos	192	0
Coumaphos	219	0
Azinphos Methyl	238	0

Laboratory Control Samples

Laboratory control samples (LCS) are analyte compounds spiked into deionized water at known concentrations and subjected to analysis. They are used to evaluate accuracy of pesticide residue recovery for a specific analyte. Detections may be qualified based on low LCS recovery and/or high RPD between paired LCS.

Table C-12 presents the mean, minimum, and maximum percent recovery for the LCS and paired LCS for the three types of analysis, as well as the RPD between the LCS and the paired LCS for 2010.

Table C-12. Mean, minimum, and maximum percent recovery for LCS and paired LCS and the LCS and paired LCS RPD.

Analysis	LCS Recovery			%RPD for LCS\LCS-Duplicate		
	Mean	Minimum	Maximum	Mean	Minimum	Maximum
LCMS\MS	70%	23%	95%	13%	7%	33%
GCMS-Herbicides	72%	37%	196%	18%	7%	108%
GCMS-Pesticides	106%	36%	155%	9%	3%	45%

In 2010, the average recovery for all three analyses and the average RPD was good, showing acceptable performance for most compounds.

The percent recoveries for the LCMS\MS analysis were good. Four compounds (1-naphthol, aldicarb, methomyl oxime, and oxamyl oxime) had average recovery rates just below the target range of 50-150%. Methomyl oxime recoveries averaged 23%, while the other compounds averaged 41-48% recovery. If LCS recoveries were low and that compound was detected, the result was qualified as an estimate.

The percent recoveries for the GCMS-Herbicide analysis were also good with most analyte recoveries between the acceptable range of 40-130%. Acifluorfen had high recoveries (195-197%); but acifluorfen was not detected in any of the 2010 samples.

The percent recoveries for the GCMS-Pesticides included many analytes, and generally percent recoveries were good. Nine analytes slightly exceeded the acceptable range of 30-130% recoveries. These analytes were: azinphos methyl (147%), beta-BHC (135%), chlorothalonil (142%), coumaphos (155%), fluridone (136%), methyl paraoxon (138%), mevinphos (133%), napropamide (133%), and tetrachlorvinphos (132%). Of these analytes only two were detected once in Indian Slough: chlorothalonil and napropamide. The chlorothalonil detection was qualified as an estimate, and the napropamide detection needed no qualification because recoveries were within acceptable limits that day.

Field Data Quality

Quality Control Procedures

Field meters were calibrated at the beginning of the field day according to manufacturers' specifications, using Ecology's standard operating procedures (Swanson, 2007). Meters were post-checked at the end of the field day using known standards. Conventional parameters measured in the field were replicated once per sample day. Dissolved oxygen meter results were compared to Winkler laboratory titration results from grab samples. Two to three Winkler grab samples were obtained during each sample day.

Results for 2010

In 2010 the field meter for the lower Yakima sites and Wenatchee-Entiat sites (eastside sites) met QC objectives including post-checks and Winkler comparisons (Table C-13) except the conductivity post-checks for July 7, August 9 and 25, and October 20. Conductivity data for these days did not meet post-check standards, and data were qualified as estimates.

The field meter for the urban sites and the lower Skagit-Samish (westside sites) met QC objectives including post-checks and Winkler comparisons (Table C-13) except two dissolved oxygen measurements for Indian Slough. Two meter and Winkler dissolved oxygen results had an 11.3 and 14.5% RSD on June 15 and 28, respectively. This slightly exceeds the QC objective of ≤ 10% RSD. In addition, a replicate conductivity reading for Indian Slough on August 20 had a 42.5% RSD, exceeding the QC objective of ≤ 10% RSD.

Table C-13. Quality control results (%RSD) for field meter and Winkler replicates.

Replicate Meter Parameter	Westside		Eastside	
	Average	Maximum	Average	Maximum
Winkler and meter DO	1.6%	14.5%	1.4%	5.5%
Replicate Winkler's for DO	0.6%	2.5%	0.3%	1.8%
Meter DO	0.6%	5.5%	n/a	n/a
Meter conductivity	2.1%	42.5%	2.1%	6.9%
Meter pH	0.6%	4.2%	0.9%	2.4%
Meter flow	5.2%	29.0%	4.5%	32.6%

DO= dissolved oxygen.

At times the Indian Slough site is influenced by incoming marine water. When this occurs, temperature, dissolved oxygen, and conductivity values can vary greatly by depth. Thus, it is difficult to obtain consistent meter readings at the Indian Slough site. It is likely that environmental factors are the cause of the differences in the dissolved oxygen and conductivity replicates. Field QC objectives were met. Indian Slough dissolved oxygen and conductivity results for these days will be qualified as estimates.

Four replicate flow results exceeded data QC objectives, three for the eastside sites and one for the westside sites (Table C-14). Flow replicates were during low-flow conditions when the RSD statistic produces higher variability. Flow results for these days are acceptable.

Table C-14. Streamflow results where the %RSD exceeded the quality control of 10% RSD.

Site	Date	Flow (cfs)	Replicate flow (cfs)	Difference in %RSD
Westside Meter				
BD-2	09/03/10	0.8	0.6	21%
Eastside Meter				
BR-1	3/9/10	0.5	0.3	33%
BR-1	6/14/10	3.6	2.9	14%
MI-1	8/16/10	2.1	1.6	17%

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Appendix D. Assessment Criteria and Water Quality Standards

EPA pesticide assessment documents were reviewed to determine the most comparable and up-to-date toxicity guidelines for freshwater (Table D-1) and marine species (Table D-2).

EPA Toxicity Criteria

Rainbow trout are a surrogate for freshwater endangered and threatened species. *Daphnia magna* (invertebrate) and *Selenastrum capricornutum* (green algae also called *pseudokirchneria subcapitata*) represent components of the aquatic food web that may be affected by pesticide use. Alternative species are used only if no data are available for rainbow trout, *Daphnia magna*, or *Selenastrum capricornutum*.

Marine toxicity criteria were evaluated for detections at sites with estuarine influence. These sites were all in the Skagit-Samish watersheds and included lower Big Ditch, Browns Slough, and Indian Slough. Criteria were generated for marine species including (1) sheepshead minnow and tidewater silverside for fish; (2) pink shrimp, Eastern Oyster, Grass Shrimp, *Acartia tonsa* (copepod), and Mysid shrimp for invertebrates; and (3) *Isochrysis galbana*, and *Skeletonema costatum* for aquatic plants.

EPA classifies a laboratory study as 'core' if it meets guidelines appropriate for inclusion in pesticide registration. Usually a core designation may be made if the study is appropriately designed, monitored, and conditions controlled, and duration of exposure is consistent with other studies. Core study criteria are used in the assessment table. Keeping with pesticide review precedent, the most toxic, acceptable criteria from core studies are used.

Water Quality Standards and Assessment Criteria

The most recent versions of Washington State water quality standards and EPA National Recommended Water Quality Criteria (NRWQC) were applied for this report. The NRWQC remained largely unchanged from the 2003 update through 2008.

The toxic standards for Washington State waters were also used. These remain essentially unchanged following the 1997 rule and 2003 updates (Washington Administrative Code (WAC), Chapter 173-201A).

Table D-1. Freshwater toxicity and regulatory guideline values. All values reported in ug/L.

Chemical	[1]Freshwater Toxicological and Reregistration Criteria													Freshwater Standards and Criterion			
	Fisheries					Invertebrate				Plant				[2]WAC		[3]NRWQC	
	Acute	Chronic	ESLOC	Spp.	Ref	Acute	Chronic	Spp.	Ref	Acute	Chronic	Spp.	Ref	Acute	Chronic	CMC	CCC
1-Naphthol	1400		70	RT	10	700		DM	10	1100		SC	10				
2,4-D (Acids, Salts, Amines)[m]	101000	14200	5050	RT; FM	1	25000	16050	DM	1	3880	1440	ND	1				
2,4-D (BEE Ester)[m]	428		21.4	BS	1	4970	200	DM	1	1020	538	ND	1				
2,4'-DDD																	
2,4'-DDT																	
3-Hydroxycarbofuran	362	5.7	18.1	RT	54; 60	2.23	0.75	CD	54								
	88		4.4	BG	54	29	9.8/27	DM	60								
4,4'-DDD																	
4,4'-DDE														1.1[a,b]	0.001[a,c]	1.1[a]	0.001[a]
4,4'-DDT														1.1[a,b]	0.001[a,c]	1.1[a]	0.001[a]
4-Nitrophenol																	
Alachlor	2100	187	105	RT	2	1550	110	DM	2	1.64	0.35	SC	2				
Aldicarb	560	78	28	RT;FM	3	410	20	DM	3								
Aldicarb Sulfone	42000	78	2100	RT;FM	3	280	20	DM	3								
Aldicarb Sulfoxide	7140	78	357	RT-A; FM-C	3	696	20	DM	3								
Atrazine	5300	65	265	RT; BT	4	6900	140	DM	4	49		SC	4				
Azinphos Methyl	2.9	0.23	0.145	RT	5	1.1	0.25	DM	5								0.01
	3.2		0.16	Coho	5												
Bentazon	>100000		>5000	RT	6	>100000		DM	6	4500		SC	6				
Bromacil	36000		1800	RT	7	121000		DM	7	6.8		SC	7				
Bromoxynil	50	18/ 39	2.5	RT-A; FM-C	8	11	2.5/5.9	DM	8	80		SC	8				
Carbaryl	1200		60	RT	9	5.6	1.5	DM	10	1100	370	SC	10				
	2400		120	Chinook	10												
	2400		120	Coho	10												
Carbofuran	362	5.7	18.1	RT	54; 60	2.23	0.75	CD	54								
	88		4.4	BG	54	29	9.8/27	DM	60								
Chlorothalonil	42.3	3	2.12	RT; FM	46	68	39	DM	46	190		SC	46				
Chlorpropham	5700		285	RT	47	3700		DM	47								
Chlorpyrifos	3	0.57	0.15	RT; FM	11; 12	0.1	0.04	DM	11					0.083d	0.041e	0.083	0.041

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Table D-1 (continued). Freshwater toxicity and regulatory guideline values.

Chemical	[1]Freshwater Toxicological and Reregistration Criteria													Freshwater Standards and Criterion			
	Fisheries					Invertebrate				Plant				[2]WAC		[3]NRWQC	
	Acute	Chronic	ESLOC	Spp.	Ref	Acute	Chronic	Spp.	Ref	Acute	Chronic	Spp.	Ref	Acute	Chronic	CMC	CCC
cis-Permethrin[n]	2.9;17	0.30/ 0.41	0.145	RT;CS-A FM-C	58	0.039	0.039/ 0.084	DM	58								
	0.79		0.0395	BG	58												
Clopyralid	1968000	N/A	98400	RT	59, 64	113000	N/A	DM	59, 64	6900		SC	59				
Cycloate	4500		225	RT	48	24000		DM	48								
DCPA	6600	N/A	330	RT	56	27000	N/A	DM	56	>12380		SC	56				
Diazinon	90	0.8	4.5	RT; BT	13; 14	0.8	0.17	DM	13	3700		SC	13			0.17	0.17
Dicamba I	28000		1400	RT	15	34600	16400	DM	15	3700	5	SC; AFA	15				
Dichlobenil	4930	330	246.5	RT	16; 17	6200	560	DM	17	1500	160	SC	17				
Dimethoate	6200	430	310	RT	18	3320	40	DM	18								
Diphenamid	97000		4850	RT	59	58000		DM	59								
Disulfoton Sulfone	9200		460	RT	20, 66	35.2	0.14/ 0.27	DM	20, 66								
Diuron	1950	26.4	97.5	RT; FM	21; 22	1400	200	DM	22	2.4		SC	22				
Endosulfan I	0.8	0.1	0.04	RT	23	166	2	DM	23					0.22[b,f]	0.056[c,f]	0.22[i]	0.056[i]
Endosulfan II	0.8	0.1	0.04	RT	23	166	2	DM	23					0.22[b,f]	0.056[c,f]	0.22[i]	0.056[i]
Endosulfan Sulfate	2.2		0.11	ND	23	580		DM	23								
Endrin Aldehyde																	
Eptam	14000		700	ND	24	6500		ND	24	1360		SC	24				
Ethoprop	1020	180	51	RT; FM	25	44	0.8	DM	25								
Fenarimol	2100	430	105	RT	67	6800	113	DM	67		100	SC	67				
Hexachlorobenzene	1000	3.68	50	CH-A; RT-C	59, 26	30	16	DM	26	30		SC	26				
Hexazinone	180000	17000	9000	RT; FM	27; 28	151600	20000	DM	27	7	4	SC	27				
	317000		15850	Chinook	27												
	246000		12300	Coho	27												
	317000		15850	Sockeye	27												
Imidacloprid	>83000	1200/ 2500	4150	RT	61	69	1800/ 3600	CT-A; DM-C	61								
						85200		DM	59								
Linuron	3000	<42	150	RT	49	120		DM	50	67		SC	49				

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Table D-1 (continued). Freshwater toxicity and regulatory guideline values.

Chemical	[1]Freshwater Toxicological and Reregistration Criteria													Freshwater Standards and Criterion			
	Fisheries					Invertebrate				Plant				[2]WAC		[3]NRWQC	
	Acute	Chronic	ESLOC	Spp.	Ref	Acute	Chronic	Spp.	Ref	Acute	Chronic	Spp.	Ref	Acute	Chronic	CMC	CCC
Malathion	4.1	21	0.205	RT	30	1	0.06	DM	30								0.1
	170		8.5	Coho	31												
MCPA	1150	916	57.5	RT	32	280	77	DM	32	250	32	SC	32				
MCPP	93000	N/A	4650	RT	65	91000	50800/ 102700	DM	65	14	9	SC	65				
Metalaxyl	132000	9100	6600	RT; FM	51	29000	1270	DM	51	140000		SC	51				
Methiocarb	436		21.8	RT	C	19		DM	C								
Methomyl	860	57/ 117	43	RT-A; FM-C	57	5	>0.4	DM	57								
Methomyl Oxime																	
Metolachlor	3900	780	195	ND	33	25100		DM	33								
Metribuzin	77000		3850	RT	52	4200	1290	DM	52	11.9	8.9	NP	51				
Napropamide	6400	1100	320	RT	53	14300	1100	DM	53	3400		SC					
Norflurazon	8100	770/ 1500	405	RT	34	15000	1000/ 2600	DM	34	9.7	3.2	SC	34-A 59-C				
Oryzalin	3260		163	RT	D	1400		DM	D								
Oxamyl	4200	770/ 1500	210	RT	62	180	1000/ 4200	CP-A; DM-C	62	120	4.6	SC	62				
Oxamyl Oxime																	
Oxyfluorfen	250	38/74	12.5	RT-A; FM-C	35, 36	80	13/28	DM	35, 36	0.29	0.1	SC	35, 36				
Pendimethalin	138	6.3	6.9	RT; FM	37	280	14.5	DM	37	5.4	3	SC	37				
Pentachlorophenol	15	11	0.75	RT	38	450	240	DM	38	50		SC	38	8.2 to 41.0[d,g]	5.2-25.9[e,h]	7.9-107.6[j]	6.1-82.6[k]
Picloram	5500	N/A	275	RT	53	34400	N/A	DM	53								
Promecarb																	
Prometon	12000	9500	600	RT-A; FM-C	68	25700	3500/ 6800	DM	68	98	32	SC	68				
Propargite	118	16	5.9	RT; FM	40	74	9	DM	40	66.2	5	SC	40				
Propoxur	3700		185	RT	63	11		DM	63								
Simazine	70500	1200	3525	RT; FM	41	1100		DM	41	100		SC	41				

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Table D-1 (continued). Freshwater toxicity and regulatory guideline values.

Chemical	[1]Freshwater Toxicological and Reregistration Criteria													Freshwater Standards and Criterion			
	Fisheries					Invertebrate				Plant				[2]WAC		[3]NRWQC	
	Acute	Chronic	ESLOC	Spp.	Ref	Acute	Chronic	Spp.	Ref	Acute	Chronic	Spp.	Ref	Acute	Chronic	CMC	CCC
Tebuthiuron	143000	9300	7150	RT; FM	42	297000	21800	DM	42	50	13	SC	42				
Terbacil	46200		2310	RT	43	65000		DM	43	18	4	SC	43				
Triadimefon	4100	41/ 116	205	RT	55	1600	52/119	DM	55	100/1710		SC	55				
Triclopyr	650		32.5	RT	44	12000		DM	44	2300	2	SC; NP	44				
Trifluralin	41	1.14	2.05	RT	45	560	2.4	DM	45	7.52	5.37	SC	45				

*Values are not analytically qualified. Non-asterisk values have been J-qualified as estimates, normally below the practical quantitation limit.
[1]Criteria identified in EPA reregistration and review documents or peer reviewed literature. References listed separately.
Time component of standards are explained in body of report.
ESLOC refers to Endangered Species Level of Concern.
Species abbreviated in table: RT-Rainbow Trout, CS-Coho Salmon, CH-Chinook salmon, FM- Fathead Minnow, BT-Brook Trout, BS-Bluegill Sunfish, ND-Not Described, DM-Daphnia magna, CD-Ceriodaphnia dubia, SC-Selenastrum capricornutum (aka; Pseudokirchneria subcapitata), Anabaena flos-aquae, and Navicula pelliculosa, SM-sheepshead Minnow, CT-Chironomus tentans (midge).
[2]WAC: Promulgated standards according to Chapter 173-201A WAC.
[3]EPA National Recommended Water Quality Criteria (EPA-822-R-02-047).
CMC: Criteria Maximum Concentration; estimate of the highest concentration of a material in surface water to which an aquatic community can be exposed briefly without resulting in an unacceptable effect.
CCC: Criteria Continuous Concentration; estimate of the highest concentration of a material in surface water to which an aquatic community can be exposed indefinitely without resulting in an unacceptable effect.
a-Criteria applies to DDT and its metabolites (ΣDDT).
b-An instantaneous concentration not to be exceeded at any time.
c-A 24-hour average not to be exceeded.
d-A 1-hour average concentration not to be exceeded more than once every three years on average.
e-A 4-day average concentration not to be exceeded more than once every three years on average.
f-Chemical form of endosulfan is not defined in WAC 173-201A. Endosulfan sulfate may be applied in this instance.
g≤ e[1.005(pH)-4.830], pH range of 6.9 to 9.5 shown.
h≤ e[1.005(pH)-5.29], pH range of 6.9 to 9.5 shown.
i-Value refers to ∑α and β-endosulfan.
j≤ e[1.005(pH)-4.869], pH range of 6.9 to 9.5 shown.
k≤ e[1.005(pH)-5.134], pH range of 6.9 to 9.5 shown.
m-There are many forms of 2,4-D that include acids, salts, amines, and esters all of which have unique toxicity values. The criteria presented are in acid equivalents and are intended to provide a range of possible effects. Toxicity values for each form of 2,4-D are available in the referenced document.
n-Assessment criteria for permethrin are based on a formulation of cis and trans-permethrin isomers. Manchester Laboratory analysis includes only the cis-permethrin isomer, the more toxic of the two; and cis-permethrin concentrations are compared to the assessment criteria for permethrin.

Table D-2. Marine toxicity and regulatory guideline values for three estuarine sites. All values are reported in ug/L

Chemical	EPA Marine Toxicological and Registration Criteria													Marine Standards and Criterion			
	Fisheries					Invertebrate				Plant				[2]WAC		[3]NRWQC	
	Acute	Chronic	ESLOC	Spp.	Ref	Acute	Chronic	Spp.	Ref	Acute	Chronic	Spp.	Ref	Acute	Chronic	CMC	CCC
1-Naphthol	1200		60	SM	10	2100		EO	10								
2,4-D (Acids, Salts, Amines) [m]	>80,000 (175,000 definitive)	no data	4000	TS	1	57000	no data	EO	1								
2,4-D (BEE Ester)[m]	no data	555		SM	1	1800	no data	EO	1								
2,4'-DDD																	
2,4'-DDT																	
3-Hydroxycarbofuran	33	2.6	1.65	AS; SM	60	4.6	0.4	PS; MS	60								
4,4'-DDD																	
4,4'-DDE														0.13[a]	0.001[b]		
4,4'-DDT														0.13[a]	0.001[b]		
4-Nitrophenol																	
Alachlor																	
Aldicarb																	
Aldicarb Sulfone																	
Aldicarb Sulfoxide																	
Atrazine	2000	2542	100	SM	4	94	80	AT; M	4	22		IG	4				
Azinphos Methyl																	
Bentazon	136		6.8	SM	6	>132.5; >109		PS; EO	6								
Bromacil	162		8.1	SM		12.9; 130		M; EO	7								
Bromoxynil																	
Carbaryl	2600		130	SM	10	32; >2		PS; EO	10								
Carbofuran	33	2.6	1.65	AS; SM	60	4.6	0.4	PS; MS	60								
Chlorothalonil	32		1.6	SM	46	154; 3.6	1.2	PS; EO; M	46								
Chlorpropham																	
Chlorpyrifos	270	0.38	13.5	SM; TS	11	2.4	<0.0046	PS; M	11					0.011[c]	0.0056[d]	0.011[G]	0.0056[G]
cis-Permethrin[n]	2.2	0.83	0.11	AS; SM	58	0.019	0.011	M	58								
Clopyralid																	
Cycloate																	

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Table D-2 (continued). Marine toxicity and regulatory guideline values for three estuarine sites.

Chemical	EPA Marine Toxicological and Registration Criteria													Marine Standards and Criterion			
	Fisheries					Invertebrate				Plant				[2]WAC		[3]NRWQC	
	Acute	Chronic	ESLOC	Spp.	Ref	Acute	Chronic	Spp.	Ref	Acute	Chronic	Spp.	Ref	Acute	Chronic	CMC	CCC
DCPA	>1000		50	SM	56	620		EO	56	>11000		SkC	56				
Diazinon						4.2	0.23	M	13							0.82	0.82
Dicamba I	>180000		>9000	SM	15												
Dichlobenil	14000		700	SM	16	>1000; 2500		PS; EO	16								
Dimethoate																	
Diphenamid																	
Disulfoton Sulfone																	
Diuron	6700		335	SM	22		270	M	22								
Endosulfan I														0.034[a]	0.0087[b]	0.034[a]	0.0087[b]
Endosulfan II														0.034[a]	0.0087[b]	0.034[a]	0.0087[b]
Endosulfan Sulfate																	
Endrin Aldehyde																	
Eptam																	
Ethoprop																	
Fenarimol																	
Hexachlorobenzene																	
Hexazinone																	
Imidacloprid	163000		8150	SM	61	37	>0.6/1.3	MS	61								
Linuron	890		44.5	SM	49	4500; 890		M; EO									
Malathion																	
MCPA	>4100	4100	>205	SM	32	150000	115000	EO	32	300	15	SkC	32				
MCPP																	
Metalaxyl						25700; 4600		M; EO	51								
Methiocarb																	
Methomyl	1160		58	SM	57	>140000; 230		EO; M	57								
Methomyl Oxime																	
Metolachlor	7900	1000	395	ND	33												

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Table D-2 (continued). Marine toxicity and regulatory guideline values for three estuarine sites.

Chemical	EPA Marine Toxicological and Registration Criteria													Marine Standards and Criterion			
	Fisheries					Invertebrate				Plant				[2]WAC		[3]NRWQC	
	Acute	Chronic	ESLOC	Spp.	Ref	Acute	Chronic	Spp.	Ref	Acute	Chronic	Spp.	Ref	Acute	Chronic	CMC	CCC
Metribuzin	85000		4250	SM	52	48300; 49800		M; EO	52	8.7	5.8	SkC	52				
Napropamide	14000		700	SM	53	4200; 1400		M; EO									
Norflurazon																	
Oryzalin																	
Oxamyl	2600		130	SM	62	0.4		EO	62								
Oxamyl Oxime																	
Oxyfluorfen																	
Pendimethalin																	
Pentachlorophenol	240		12	SM	38	48		PO	38	27		SkC	38	13.0[c]	7.9[d]		
Picloram																	
Promecarb																	
Prometon	47300		2365	SM	68	18000		MS	68								
Propargite																	
Propoxur																	
Simazine	>4300		215	SM	41	113000; >3700		PS; EO	41	600		SkC	41				
Tebuthiuron						62000		PS	42	31		SkC	42				
Terbacil																	
Triadimefon																	
Triclopyr	450		22.5	TS	44	2470		GS	44	1170	209	SkC	44				
Trifluralin	190		9.5	SM	45	638.5		GS	45	28		SkC	45				

*Values are not analytically qualified. Non-asterisk values have been J-qualified as estimates, normally below the practical quantitation limit.
[1]Criteria identified in EPA reregistration and review documents or peer reviewed literature. References listed separately.
Time component of standards are explained in body of report.
ESLOC refers to Endangered Species Level of Concern
Species abbreviated in table: ND-Not determined, AS-Atlantic silverside, IS-Inland silverside, TS-Tidewater silverside, PS-Pink Shrimp, EO-Eastern Oyster, AT-Acartia tonsa (copepod), M-Mysid, IG-Isochrysis galbana, LG-Lemna gibba, CT-Chironomus tentans (midge), GS - Grass Shrimp, SkC - Skeletonema costatum, PO-Pacific Oyster
[2]WAC: Promulgated standards according to Chapter 173-201A WAC.
[3]EPA National Recommended Water Quality Criteria (EPA-822-R-02-047).
CMC: Criteria Maximum Concentration; estimate of the highest concentration of a material in surface water to which an aquatic community can be exposed briefly without resulting in an unacceptable effect.

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CCC: Criteria Continuous Concentration; estimate of the highest concentration of a material in surface water to which an aquatic community can be exposed indefinitely without resulting in an unacceptable effect.

a-Criteria applies to DDT and its metabolites (ΣDDT).

b-An instantaneous concentration not to be exceeded at any time.

c-A 24-hour average not to be exceeded.

d-A 1-hour average concentration not to be exceeded more than once every three years on average.

e-A 4-day average concentration not to be exceeded more than once every three years on average.

f-Chemical form of endosulfan is not defined in WAC 173-201A. Endosulfan sulfate may be applied in this instance.

g≤ e[1.005(pH)-4.830], pH range of 6.9 to 9.5 shown.

h≤ e[1.005(pH)-5.29], pH range of 6.9 to 9.5 shown.

i-Value refers to ∑α and β-endosulfan.

j≤ e[1.005(pH)-4.869], pH range of 6.9 to 9.5 shown.

k≤ e[1.005(pH)-5.134], pH range of 6.9 to 9.5 shown.

m-There are many forms of 2,4-D that include acids, salts, amines, and esters all of which have unique toxicity values. The criteria presented are in acid equivalents and are intended to provide a range of possible effects. Toxicity values for each form of 2,4-D are available in the referenced document.

n-Assessment criteria for permethrin are based on a formulation of cis- and trans-permethrin isomers. Manchester Laboratory analysis includes only the cis-permethrin isomer, the more toxic of the two; and cis-permethrin concentrations are compared to the assessment criteria for permethrin.

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Appendix E. Pesticide Calendars

To determine if water quality concentrations were healthy for aquatic life, monitoring data were compared to EPA pesticide registration toxicity criteria and EPA National Recommended Water Quality Criteria (NRWQC), referred to as *assessment criteria* in this report. Data were also compared to numeric Washington State water quality standards, referred to as *water quality standards*. Refer to Appendix D, *Assessment Criteria and Water Quality Standards*, in this report for information on assessment criteria development.

Table E-1 presents the color codes used to compare detected pesticide concentrations to assessment criteria. In the calendars, the number below the months indicate sample week.

Table E-1. Color codes for comparison to assessment criteria in the pesticide calendars.

	Each square represents the period when a sample was taken. If blank then no pesticide residue was detected.
	Analysis not completed.
	Pesticide residue detected. Assessment criteria not available.
	Detection of pesticide residue, concentration below regulatory or toxicological criteria or standard.
	Magnitude of detection above an EPA[1] acute or chronic invertebrate registration criteria.
	Magnitude of detection above an WAC2 or NRWQC[3] acute or chronic regulatory standard.
	Magnitude of detection above an EPA[1] acute or chronic invertebrate registration criteria.

[1] EPA: United States Environmental Protection Agency
[2] WAC: Washington Administrative Code
[3] NRWQC: EPA's National Recommended Water Quality Criteria

Detection of a pesticide concentration above an assessment criteria does not indicate exceedance of (not meeting) the regulatory criteria. The temporal component of the criteria must also be exceeded. The Washington State Department of Agriculture (WSDA) advises pesticide user groups and other stakeholders on the results of this study and determines if assessment criteria are exceeded. If an exceedance is determined, WSDA advises stakeholders of appropriate measures to reduce pesticide concentrations.

For additional information on pesticide assessment criteria, contact the WSDA, Natural Resources Assessment Section, toll free at (877) 301-4555, #6 or (360) 902-2067, or e-mail: nras@agr.wa.gov. Their web site is http://agr.wa.gov/PestFert/natresources/SWM/.

Cedar-Sammamish Basin

Thornton Creek

In 2010 there were 58 detections of 10 pesticides in Thornton Creek (Table E-2).

No detections were above assessment criteria or water quality standards.

Table E-2. Thornton Creek 2010.

Month		**March**				**April**				**May**				**June**					**July**				**August**				**Sep**	
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
2,4-D	H				0.073				0.110	0.056										0.095						0.087	0.033	0.067
Carbaryl	I-C								0.005																			
Dichlobenil	H	0.017	0.008	0.013	0.027	0.011	0.009	0.009	0.044	0.014	0.010		0.008	0.021	0.012	0.014	0.014	0.007	0.001	0.002		0.012	0.012	0.016	0.012	0.015	0.008	
Diuron	H								0.039					0.053												0.028		
Imidacloprid	I-N											0.005		0.005							0.004	0.003	0.003					
MCPA	H																										0.031	
MCPP	H								0.050																		0.022	
Pentachlorophenol	WP		0.018	0.032			0.019	0.021	0.031					0.021				0.018									0.024	0.049
Propoxur	I-C								0.008																			
Triclopyr	H													0.035						0.063						0.064	0.150	0.210
Total Suspended Solids	NA	6.0	2.0	13.0	9.0	5.0	6.0	6.0	15.0	7.0	5.0	8.0	18.5	5.0	6.0	9.0	6.0	4.3	4.0	8.0	11.0	7.3	5.0	5.0	6.0	4.0	5.0	12.0

C: Carbamate, H: Herbicide, I: Insecticide, N: Neonicotinoid, NA: Not applicable, WP: Wood Preservative

Green-Duwamish Basin

Longfellow Creek

In 2010 there were 78 detections of 15 pesticides and degradates in Longfellow Creek (Table E-3).

No detections were above assessment criteria or water quality standards.

Table E-3. Longfellow Creek 2010.

Month		March				April				May				June					July				August				Sep	
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
2,4-D	H				0.057		0.036	0.032	0.540	0.150	0.068										0.024		0.030		0.130	0.042	0.038	0.086
Carbaryl	I-C								0.003																			
Carbofuran	I-C												0.003															
Dicamba I	H								0.076																			
Dichlobenil	H	0.017	0.011		0.054	0.027	0.017	0.021	0.210	0.078	0.026	0.008	0.010	0.024	0.010		0.014	0.006	0.002			0.013	0.011	0.015	0.012	0.011		0.017
Diuron	H								0.030																			
Imidacloprid	I-N		0.007									0.005	0.006		0.005						0.004	0.005	0.003	0.006			0.004	
MCPP	H								0.160	0.055																		
Metalaxyl	F				0.042																							
Methomyl	I-C												0.004															
Oxamyl	I-C												0.004															
Oxamyl oxime	D-C			0.013																								
Pentachlorophenol	WP		0.018						0.035							0.017				0.033	0.016							
Prometon	H							0.110																				
Triclopyr	H	0.034	0.033	0.031	0.080			0.049	0.140	0.092	0.049			0.070	0.053	0.031	0.036	0.031			0.031		0.037		0.080	0.052	0.048	0.150
Total Suspended Solids	NA	3.0	2.0	2.0	7.0	8.0	2.0	<3.0	17.0	6.0	5.3	4.0	4.0	4.0	4.0	4.0	3.0	4.0	2.0	9.0	2.0	3.0	2.0	4.0	2.0	9.0	2.0	3.0

C: Carbamate, D: Degradate, F: Fungicide, H: Herbicide, I: Insecticide, N: Neonicotinoid, NA: Not applicable, WP: Wood Preservative

Skagit-Samish Basins

Big Ditch

A total of 30 pesticides and degradates were detected in Big Ditch in 2010. Of these, 22 were found at the upper Big Ditch site (Table E-4). A total of 26 pesticides and degradates were found at the lower Big Ditch site (Table E-5).

During 2010 no detections at either Big Ditch site were above pesticide assessment criteria or water quality standards.

Comparison of Upper Big Ditch to Lower Big Ditch

In 2010 the upper and lower sites on Big Ditch were sampled weekly on the same day. During the year, 18 pesticides were detected in common between the two sites: 2,4-D, bromacil, carbaryl, carbofuran, chlorpropham, dicamba, dichlobenil, diuron, eptam, imidacloprid, MCPA, mecoprop (MCPP), metalaxyl, Methiocarb, Metolachlor, pentachlorophenol, prometon, and triclopyr. Four compounds were detected only at the upper site: oxamyl, picloram, tebuthiuron, and the synergist piperonyl butoxide. Eight compounds were detected only at the lower site: 3-hydroxycarbofuran, atrazine, bentazon, cycloate, ethoprop, fipronil, linuron, and trifluralin.

Table E-4. Upper Big Ditch 2010.

Month		**March**				**April**				**May**				**June**					**July**				**August**				**Sep**	
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
2,4-D	H			0.063	0.045		0.160		0.120	0.170			0.052	0.170		0.235								0.058				0.073
Bromacil	H	0.045	0.050		0.055	0.043		0.059			0.048	0.037	0.035	0.060	0.021	0.061	0.068	0.058	0.042	0.033	0.050	0.057	0.062	0.051	0.050	0.055	0.057	
Carbaryl	I-C								0.005																			
Carbofuran	I-C												0.003															
Chlorpropham	H																						0.038					
Dicamba I	H								0.026					0.016		0.150												
Dichlobenil	H	0.029	0.022	0.067	0.046	0.013	0.022	0.012	0.056	0.097	0.011	0.009	0.010	0.062	0.020	0.022	0.015	0.007	0.002		0.006	0.011	0.011	0.015	0.021	0.012	0.007	
Diuron	H								0.032				0.017	0.062	0.041	0.074								0.041			0.130	0.089
Eptam	H										0.027																	
Imidacloprid	I-N		0.017			0.009	0.012	0.016	0.072	0.095	0.093	0.387	0.079	0.023	0.133	0.016	0.012	0.018	0.215	0.095	0.035	0.303	0.033	0.009	0.879		0.005	
MCPA	H								0.041							0.060												
MCPP	H								0.048	0.120				0.026														0.040
Metalaxyl	F										0.060				0.049					0.250	0.190	1.000			0.083			
Methiocarb	I-C	0.003																										
Metolachlor	H															0.041												
Oxamyl	I-C																0.003	0.004		0.003								
Pentachlorophenol	WP	0.025	0.019						0.032	0.025			0.021	0.024	0.021	0.020												
Picloram	H											0.061						0.120										
Piperonyl Butoxide	Sy									0.120																		
Prometon	H	0.130		0.046										0.040														
Tebuthiuron	H																	0.054				0.035				0.036	0.047	
Triclopyr	H			0.040	0.051				0.077	0.070			0.063	0.110	0.042	0.043	0.030							0.043				0.066
Total Suspended Solids	NA	12.0	3.0	12.0	5.0	3.0	3.0	4.0	8.5	15.0	3.5	6.0	7.0	9.0	4.0	4.0	4.0	8.0	7.0	5.0	4.0	11.5	9.0	6.5	16.0	8.0	5.5	7.0

C: Carbamate, F: Fungicide, H: Herbicide, I: Insecticide, N: Neonicotinoid, NA: Not applicable, Sy: Synergist, WP: Wood Preservative

Table E-5. Lower Big Ditch 2010.

Month		**March**				**April**				**May**				**June**					**July**				**August**				**Sep**	
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
2,4-D	H		0.057	0.086	0.098			0.077		0.110			0.140	0.160	0.110	0.033												0.041
3-Hydroxycarbofuran	D-C													0.004														
Atrazine	H		0.054														0.059											
Bentazon	H				0.056																							
Bromacil	H											0.022																
Carbaryl	I-C					0.012																						
Carbofuran	I-C					0.067						0.005	0.005	0.584	0.018	0.008	0.004											
Chlorpropham	H	0.770	1.500	0.690	0.260	0.056	0.250	0.067	0.024																			
Cycloate	H													0.073														
Dicamba I	H	0.053	0.026													0.026												
Dichlobenil	H		0.024	0.009	0.037		0.012		0.010	0.052	0.006	0.007	0.006	0.032	0.016	0.011	0.009										0.006	0.009
Diuron	H						1.500	1.300	0.230	3.400	0.115	0.160	0.100	1.100	0.098	0.290		0.012										
Eptam	H						0.080	0.081			0.210		0.024															
Ethoprop	I-OP													0.200														
Fipronil	I-Pyra													0.037														
Imidacloprid	I-N								0.034	0.166	0.055	0.055	0.024	0.024	0.023	0.014	0.027	0.022	0.007		0.003		0.008					
Linuron	H																											0.014
MCPA	H			0.250	0.110		0.270		0.029				0.300	0.092	0.034													
MCPP	H							0.026	0.022																			
Metalaxyl	F									0.096				0.110														
Methiocarb	I-C	0.002				0.060																						
Metolachlor	H	0.036	0.045	0.036	0.065	0.028	0.049	0.027	0.056	0.066	0.042	0.110	0.060	0.190	0.074	0.081	0.040	0.024										0.029
Pentachlorophenol	WP	0.022		0.022	0.031				0.021	0.029			0.020		0.022	0.019											0.026	0.026
Prometon	H												0.046														0.034	0.042
Triclopyr	H				0.064					0.058			0.089	0.092	0.052	0.031	0.086	0.034									0.026	0.040
Trifluralin	H										0.015																	
Total Suspended Solids	NA	7.0	13.0	10.0	10.0	6.0	9.0	9.0	11.0	12.0	8.5	25.0	8.0	11.0	9.0	6.0	4.3	4.0	5.0	4.0	2.0	2.0	2.0	<1.0	2.0	2.0	2.0	1.0

C: Carbamate, F: Fungicide, H: Herbicide, I: Insecticide, N: Neonicotinoid, NA: Not applicable, OP: Organophosphate, Pyra: Pyrethroid, WP: Wood Preservative

Indian Slough

A total of 23 pesticides and degradates were detected in Indian Slough in 2010 (Table E-6). No detections were above assessment criteria or water quality standards.

Table E-6. Indian Slough 2010.

Month		March				April				May				June					July				August				Sep	
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
2,4-D	H	0.120	0.049						0.051	0.250		0.040	0.044	0.440					0.073					0.043			3.000	1.600
Bentazon	H		0.035																									
Bromacil	H	0.037				0.031	0.037	0.040	0.082	0.120	0.035	0.029	0.035	0.205	0.140	0.080	0.060	0.047	0.032	0.027	0.036	0.041	0.042	0.038	0.035	0.041	0.650	0.310
Carbaryl	I-C													0.015														
Carbofuran	I-C									0.004			0.004	0.033	0.006	0.004												
Chlorothalonil	F														0.024													
Chlorpropham	H									0.110																		
Dicamba I	H																							0.019			0.200	0.073
Dichlobenil	H	0.009	0.022	0.026	0.039	0.011	0.009		0.018	0.130	0.006	0.006		0.037	0.075	0.011	0.007						0.009				0.007	0.026
Diphenamid	H					0.017	0.022				0.006	0.007	0.005			0.025	0.026	0.022			0.014		0.017			0.014	0.022	
Diuron	H								0.038	0.280				3.600	0.260	0.440		0.012									0.310	1.000
Eptam	H											0.036	0.022	0.069														
Ethoprop	I-OP													0.290														
Hexazinone	H	0.079		0.085		0.069	0.110			0.084	0.073	0.060	0.058	0.110	0.061	0.120	0.120	0.065	0.045		0.050							
Imidacloprid	I-N													0.020	0.007													
MCPP	H																										0.330	0.140
Metolachlor	H				0.038									0.195	0.018	0.043							0.029		0.015	0.015	0.028	0.079
Metribuzin	H													0.210														
Napropamide	H													0.440														
Pentachlorophenol	WP		0.023		0.028	0.019			0.023						0.023	0.019												
Prometon	H									0.036																	0.055	
Tebuthiuron	H																					0.040		0.039	0.040	0.045	0.049	
Triclopyr	H	0.089	0.029		0.043				0.053	0.175			0.036	0.230	0.083	0.037	0.040		0.062	0.033							0.530	0.640
Total Suspended Solids	NA	14.3	9.8	9.0	7.0	8.5	7.0	9.8	6.0	11.0	9.0	7.8	7.0	10.0	6.0	6.0	7.3	9.0	6.8	6.0	3.0	22.0	4.0	4.0	4.0	2.0	3.0	5.0

C: Carbamate, H: Herbicide, I: Insecticide, N: Neonicotinoid, NA: Not applicable, OP: Organophosphate, WP: Wood Preservative

Browns Slough

A total of 15 pesticides and degradates were detected in Browns Slough in 2010 (Table E-7). No detections were above assessment criteria or water quality standards.

Table E-7. Browns Slough 2010 – Freshwater and Marine Criteria.

Month		March				April				May				June					July				August				Sep	
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
2,4-D	H											0.063		0.043							0.047	0.370						
Bentazon	H	0.096	0.084											0.250		0.110												
Carbofuran	I-C												0.004	0.097	0.023	0.015	0.006	0.004										
DCPA	H	0.100	0.091	0.075	0.120	0.091	0.250	0.230	0.047	0.100	0.041	0.046	0.050	0.110	0.072	0.200	0.045	0.098	0.051	0.049				0.032				
Dicamba I	H																				0.022	0.160						
Dichlobenil	H	0.013	0.014	0.008			0.010							0.009														0.010
Diuron	H								0.031					0.190		0.042												
Eptam	H							0.037				0.050	0.030	0.034														
Imidacloprid	I-N													0.020	0.008	0.004			0.007	0.008								
MCPA	H							0.410	0.066			0.066	0.033	0.044														
Metalaxyl	F													0.064														
Metolachlor	H							0.130	0.021	0.015		0.011	0.008	0.590	0.028	0.046	0.035											
Simazine	H			0.037		0.072	0.034							0.031														
Terbacil	H													0.056														
Triclopyr	H						0.055							0.042														
Total Suspended Solids	NA	4.0	7.0	17.3	8.0	8.0	6.0	15.0	5.0	6.0	6.0	6.0	7.0	5.0	5.0	5.0	4.0	6.0	6.0	6.5	5.0	2.0	5.0	8.0	4.0	8.0	4.0	10.3

C: Carbamate, F: Fungicide, H: Herbicide, I: Insecticide, N: Neonicotinoid, NA: Not applicable

Samish River

A total of five pesticides were detected in the Samish River in 2010 (Table E-8). No detections were above assessment criteria or water quality standards.

Table E-8. Samish River 2010.

Month		**March**				**April**				**May**				**June**					**July**				**August**				**Sep**	
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
2,4-D	H																0.022	0.031										0.120
Dicamba I	H																0.010	0.013										
Dichlobenil	H									0.019				0.010														
Ethoprop	I-OP													0.054														
Triclopyr	H																											0.050
Total Suspended Solids	NA	8.0	7.0	10.5	17.0	10.5	11.0	8.0	12.0	151.0	12.0	10.0	6.0	51.0	18.0	13.0	9.0	5.0	5.0	2.0	3.5	5.0	3.0	5.0	4.0	4.0	7.0	8.0

H: Herbicide, I: Insecticide, NA: Not Applicable, OP: Organophosphate

Lower Yakima Basin

Spring Creek

A total of 17 pesticides and degradates were detected in Spring Creek in 2010. Of these, 14 were detected at the upper Spring Creek site (Table E-9) and 15 were detected in the lower Spring Creek site (Table E-10).

The upstream Spring Creek site met pesticide assessment criteria and water quality standards. At the lower Spring Creek site, one detection of chlorpyrifos on March 30 was above the chronic water quality standard and NRWQC for fish and the EPA chronic invertebrate criteria. Because the chlorpyrifos concentration exceeded criteria and standards only once, it is unlikely that the time component of the chronic water quality standard and NRWQC was exceeded.

Comparison of Upper Spring Creek to Lower Spring Creek

In 2010 the upper Spring Creek site was sampled every other week and the lower site was sampled weekly. During the year, 12 pesticides and degradates were detected in common between the two sites: 2,4-D, atrazine, bentazon, carbaryl, chlorpyrifos, diazinon, dicamba I, dichlobenil, diuron, imidacloprid, MCPA, and oxamyl oxime.

Carbofuran and oryzalin were detected only at the upstream site. Three pesticides were detected only at the lower site: bromacil, norflurazon, and prometon.

Table E-9. Upper Spring Creek 2010.

Month		March		April		May		June			July		August		Sep
Chemical	Type	11	13	15	17	19	21	23	25	27	29	31	33	35	37
2,4-D	H					0.040	0.050		0.037	0.038	0.050				
Atrazine	H							0.028	0.027						
Bentazon	H	0.051	0.047												
Carbaryl	I-C					0.027	0.024								
Carbofuran	I-C									0.005					
Chlorpyrifos	I-OP			0.020											
Diazinon	I-OP											0.120			
Dicamba I	H					0.019	0.017								
Dichlobenil	H			0.010											
Diuron	H					0.150		0.045							
Imidacloprid	I-N						0.005	0.007	0.005	0.006	0.004	0.006	0.004		
MCPA	H						0.025								
Oryzalin	H			1.000											
Oxamyl oxime	D-C		0.019												
Total Suspended Solids	NA	7.0	10.0	23.0	29.0	16.0	143.0	46.0	19.0	50.0	17.0	22.0	15.0	7.5	10.0

C: Carbamate, D: Degradate, H: Herbicide, I: Insecticide, N: Neonicotinoid, NA: Not Applicable, OP: Organophosphate

Table E-10. Lower Spring Creek 2010.

Month		March				April				May				June					July				August					Sep
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
2,4-D	H				0.031	0.031		0.100	0.032	0.041		0.047	0.027			0.022	0.051	0.047	0.130	0.038	0.029	0.046	0.041		0.065	0.097	0.050	0.110
Atrazine	H													0.027												0.012		
Bentazon	H		0.035				0.032																					
Bromacil	H																				0.030	0.029		0.026			0.024	
Carbaryl	I-C								0.010	0.015	0.021	0.016	0.005	0.007														
Chlorpyrifos	I-OP			0.034	0.061	0.033																						
Diazinon	I-OP																					0.021						
Dicamba I	H									0.017		0.015	0.014						0.015	0.010				0.014				
Dichlobenil	H		0.005		0.012																							
Diuron	H									0.053				0.060														
Imidacloprid	I-N											0.005		0.006	0.005	0.004	0.005	0.005				0.005	0.005	0.004		0.003	0.003	
MCPA	H											0.024																
Norflurazon	H											0.030																
Oxamyl oxime	D-C			0.026																								
Prometon	H								0.009																			
Total Suspended Solids	NA	2.0	2.0	2.0	30.0	14.0	3.0	9.0	7.0	11.0	13.0	30.0	18.0	8.0	7.0	2.0	2.0	2.0	7.0	4.0	5.0	12.5	12.0	13.0	12.0	3.0	3.0	2.0

C: Carbamate, D: Degradate, H: Herbicide, I: Insecticide, N: Neonicotinoid, NA: Not Applicable, OP: Organophosphate WP: Wood Preservative, OP: Organophosphate

Marion Drain

A total of 23 pesticides and degradates were detected in Marion Drain in 2010 (Table E-11).

Malathion was detected above EPA chronic invertebrate assessment criteria once in 2010; however, this single event did not exceed the 21-day time component of the chronic invertebrate criteria. No other detections were above assessment criteria or water quality standards.

Table E-11. Marion Drain 2010.

Month		March				April				May				June					July				August					Sep				Oct			
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37	38	39	40	41	42	43	44
2,4-D	H							0.045	0.051	0.064	0.034	0.056	0.029	0.037		0.040	0.033	0.042	0.038	0.050	0.048	0.079	0.059	0.040	0.045	0.043	0.081	0.032							
Atrazine	H									0.041																									
Bentazon	H																0.051	0.073	0.160	0.250	0.150	0.230	0.094	0.170	0.130			0.097							
Bromacil	H									0.052							0.051	0.026																	
Bromoxynil	H							0.051	0.076	0.050	0.032	0.035																							
Carbaryl	I-C								0.009	0.011	0.016						0.008	0.007																	
Chlorpyrifos	I-OP																										0.023		0.027		0.022	0.020	0.020		
Dicamba I	H								0.017	0.019	0.016	0.023	0.011	0.011		0.011	0.019	0.011	0.013	0.014	0.029	0.027	0.032	0.016	0.014	0.011		0.015							
Disulfoton sulfone	I-OP																								0.023		0.044		0.045						
Disulfoton sulfoxide	D-OP																								0.011		0.110								
Diuron	H								0.030	0.210	0.016	0.097		0.033		0.037																			
Eptam	H										0.028																								
Ethoprop	I-OP																									0.057	0.110								
Imidacloprid	I-N		0.004						0.004			0.006	0.006	0.005	0.005	0.005	0.006	0.004	0.005	0.004		0.005	0.006	0.005	0.006	0.006	0.009								
Malathion	I-OP											0.062	0.044																						
MCPA	H							0.066		0.032		0.036	0.022				0.025				0.026														
Methomyl	I-C												0.004						0.003	0.003		0.043				0.004									
Metolachlor	H													0.034		0.032		0.023																	
Pendimethalin	H							0.099	0.075	0.087	0.075	0.098	0.039	0.042	0.032	0.064	0.049	0.037		0.025															
Propargite	I-SE															0.110																			
Simazine	H									0.081				0.023																					
Terbacil	H							0.200	0.160	0.420	0.097	0.160	0.050	0.064	0.059	0.180	0.190	0.080	0.032	0.270	0.220	0.087	0.050	0.043		0.034	0.057	0.270	0.580	0.505	0.150	0.038	0.170		
Trifluralin	H							0.020		0.030	0.023	0.023	0.015	0.028	0.012	0.027																			
Total Suspended Solids	NA	6.3	10.0	16.5	10.0	16.0	47.0	48.3	19.0	16.5	13.0	12.3	21.0	23.0	23.0	9.0	4.0	5.0	1.0	3.0	5.0	3.0	26.0	6.0	2.0	5.0	11.0	9.0	8.0	11.0	17.0	4.0	8.8	5.0	9.0

C: Carbamate, D: Degradate, H: Herbicide, I: Insecticide, N: Neonicotinoid, NA: Not Applicable, OP: Organophosphate SE: Sulfite Ester

Sulphur Creek Wasteway

A total of 22 pesticides and degradates were detected in Sulphur Creek Wasteway in 2010 (Table E-12).

In March there were two consecutive detections of chlorpyrifos that did not meet (were above) the chronic water quality standard and EPA's chronic invertebrate criteria. In addition, one of these detections also did not meet the acute water quality standard and EPA's acute invertebrate criteria. Chlorpyrifos chronic water quality standard and NRWQC are not met when the 4-day average concentration is above the numerical criteria. Because chlorpyrifos was detected in two consecutive weeks, the time component of the chronic water quality standard and NRWQC was not met.

Table E-12. Sulphur Creek Wasteway 2010.

Month		**March**				**April**				**May**				**June**					**July**				**August**					**Sep**
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
2,4-D	H				0.041	0.036	0.028	0.073	0.074	0.068	0.120	0.093	0.040	0.057	0.031	0.058	0.210	0.054	0.038	0.440	0.100	0.087		0.050	0.110	0.270	0.350	0.210
Acetochlor	H									0.032				0.041														
Bentazon	H	0.052	0.049																									
Bromacil	H	0.048	0.047			0.018	0.024			0.020		0.028		0.044		0.045	0.041	0.036		0.017	0.024							
Carbaryl	I-C								0.023	0.015	0.013	0.040	0.005	0.009		0.011	0.007	0.012	0.009					0.004				
Chlorpyrifos	I-OP			0.096	0.053	0.028	0.024																					
DCPA	H		0.044							0.031							0.029	0.047	0.038	0.033								
DDVP	I-OP																	0.069										
Diazinon	I-OP																										0.033	
Dicamba I	H									0.026	0.024	0.024			0.015		0.015	0.013	0.015	0.021	0.017			0.017	0.019			0.018
Dichlobenil	H		0.004											0.005									0.009					
Disulfoton sulfoxide	D-OP																											0.026
Diuron	H								0.030	0.097	0.051	0.260		0.540		0.083	0.044											
Hexazinone	H	0.062									0.410			0.057														
Imidacloprid	I-N			0.005								0.005	0.005	0.006	0.005	0.004	0.005	0.042			0.004	0.004	0.003	0.003	0.003		0.003	
MCPA	H								0.029	0.029		0.037																
Methomyl	I-C											0.004	0.004															
Oxamyl	I-C																			0.003								
Pendimethalin	H							0.055																				
Simazine	H													0.049														
Terbacil	H																	0.036								0.025		0.095
Trifluralin	H										0.017					0.025												
Total Suspended Solids	NA	10.0	7.0	251.0	48.0	56.0	50.0	94.0	160.0	45.0	26.0	39.0	53.0	60.0	39.0	9.0	27.0	15.0	16.4	15.0	17.0	8.0	41.0	16.0	18.0	17.0	18.0	21.0

C: Carbamate, D: Degradate, H: Herbicide, I: Insecticide, N: Neonicotinoid, NA: Not Applicable, OP: Organophosphate

Wenatchee and Entiat Basins

Peshastin Creek

A total of four pesticides and degradates were detected in Peshastin Creek in 2010 (Tables E-13).

Endosulfan I was detected above the ESLOC for rainbow trout once in March. No other detected pesticides were above assessment criteria or water quality standards.

Table E-13. Peshastin Creek 2010.

Month		**March**				**April**				**May**				**June**					**July**				**August**				**Sep**	
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
3-Hydroxycarbofuran	D-C					0.004																						
Diuron	H																											0.120
Endosulfan I	I-OC			0.045																								
Simazine	H																											0.047
Total Suspended Solids	NA	2.0	2.0	1.0	7.0	2.0	<3.0	39.0	9.5	2.0	2.0	55.0	5.0	12.0	10.0	12.0	5.0	3.5	2.0	2.0	1.0	1.0	4.0	2.0	1.0	<1.0	6.0	42.0

C: Carbamate, D: Degradate, H: Herbicide, I: Insecticide, NA: Not Applicable, OC: Organochlorine

Mission Creek

In 2010 two compounds were detected in Mission Creek, an insecticide and a pesticide synergist (Tables E-14).

No detections were above assessment criteria or water quality standards.

Table E-14. Mission Creek 2010.

Month		**March**				**April**				**May**				**June**					**July**				**August**				**Sep**	
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
Carbaryl	I-C								0.007				0.006															
Piperonyl Butoxide	Sy			0.660																								
Total Suspended Solids	NA	7.0	4.0	2.0	25.0	8.0	3.0	268.0	105.0	10.0	10.0	143.0	24.0	22.5	427.0	95.0	32.0	30.0	12.0	9.0	8.0	5.0	6.0	2.0	2.0	2.0	3.0	4180

C: Carbamate, I: Insecticide, NA: Not Applicable, Sy: Synergist

Brender Creek

A total of 15 pesticides and degradates were detected in Brender Creek in 2010 (Tables E-15).

On March 24, total endosulfan was detected above the ESLOC for fish and above the chronic water quality standard.

One detection of chlorpyrifos on April 12 was above the acute and chronic water quality standard and EPA criteria. Because there was only one detection, the time component of the 4-day exposure criteria for the chronic water quality standard and NRWQC was likely not exceeded. The 21-day exposure criterion for the EPA chronic criteria was probably not exceeded with only two consecutive weeks of detections.

On September 8, one detection of diazinon was above EPA's acute and chronic criteria.

DDT and DDT degradates were found consistently throughout 2010; however, these compounds were detected less than in previous years. In 2010 there were 18 sample events out of 27 where total DDT concentrations were above the chronic water quality standard. DDT and DDT metabolite concentrations were above the chronic water quality standard and NRWQC. The chronic water quality standard is based on a 24-hour average concentration.

Table E-15. Brender Creek 2010.

Month		March				April				May				June					July				August				Sep	
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
4,4'-DDD	D-OC									0.018						0.025	0.027	0.020	0.023	0.024				0.014		0.012	0.013	0.027
4,4'-DDE	D-OC								0.042	0.042	0.026					0.043	0.029	0.012	0.006	0.014	0.024	0.021	0.038	0.011		0.024	0.033	0.045
4,4'-DDT	I-OC									0.028	0.023	0.023	0.017		0.020	0.045	0.041	0.023	0.024	0.027	0.024			0.026		0.021	0.026	0.045
Carbaryl	I-C									0.028	0.017		0.005	0.006														0.006
Chlorpyrifos	I-OP					0.024	0.120	0.029	0.027																			
Diazinon	I-OP								0.028																	0.019		0.230
Dichlobenil	H														0.004													
Diuron	H								0.031	0.180	0.024	0.860	0.025	0.038		0.070							0.067					0.047
Endosulfan I	I-OC			0.054	0.027																							
Endosulfan II	I-OC			0.029					0.029	0.035																		
Endosulfan Sulfate	D-OC			0.043	0.052	0.035	0.052	0.058	0.100	0.065	0.054	0.059	0.037	0.045	0.044	0.056	0.046	0.040	0.049	0.049				0.022		0.021	0.027	0.062
Imidacloprid	I-N											0.006		0.005								0.005	0.003	0.008	0.003	0.003		0.037
Norflurazon	H											0.470						0.049	0.022						0.032		0.040	0.120
Pendimethalin	H									0.041							0.048											
Pentachlorophenol	WP						0.016	0.016		0.020	0.015			0.015														
Total Suspended Solids	NA	11.0	13.0	7.0	7.0	7.0	44.0	14.0	249.0	108.0	53.5	50.5	25.0	7.0	36.0	83.0	68.0	25.0	30.0	54.0	37.0	25.0	103.0	21.0	12.0	31.0	143.0	125.0

C: Carbamate, D: Degradate, H: Herbicide, I: Insecticide, N: Neonicotinoid, NA: Not Applicable, OC: Organochlorine, OP: Organophosphate, WP: Wood Preservative

Wenatchee River

A total of five pesticides were detected in the Wenatchee River in 2010 (Tables E-16). No pesticide detections were above assessment criteria or water quality standards.

Table E-16. Wenatchee River 2010.

Month		**March**				**April**				**May**				**June**					**July**				**August**				**Sep**	
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
2,4-D	H																											0.040
Carbaryl	I-C													0.006														
Chlorpyrifos	I-OP					0.025																						
Dicamba I	H																											0.017
Diuron	H																											0.027
Total Suspended Solids	NA	2.0	2.0	2.0	4.0	3.0	4.0	25.5	6.0	6.0	3.0	70.0	8.0	12.0	18.0	17.0	15.0	10.0	4.0	6.0	6.0	3.0	9.0	3.0	2.0	2.0	2.0	30.0

C: Carbamate, H: Herbicide, I: Insecticide, NA: Not Applicable, OP: Organophosphate

Entiat River

A total of four pesticides and one synergist were detected in the Entiat River in 2010 (Tables E-17).

On September 1, there was one detection of DDT that was above the chronic water quality standard and EPA criteria. The chronic water quality standard is based on a 24-hour average concentration.

Table E-17. Entiat River 2010.

Month		**March**				**April**				**May**				**June**					**July**				**August**				**Sep**	
Chemical	Type	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37
2,4-D	H					0.040	0.095																					
4,4'-DDT	I-OC																										0.021	
Carbaryl	I-C								0.003		0.017																	
Imidacloprid	I-N														0.006													
Piperonyl Butoxide	Sy		0.280																									
Total Suspended Solids	NA	3.0	4.0	2.0	4.0	3.0	4.0	21.0	7.0	4.0	5.0	31.0	8.0	11.0	7.0	31.0	13.0	8.0	7.0	6.0	4.0	3.0	5.0	3.0	3.0	2.0	2.0	2.0

H: Herbicide, I: Insecticide, N: Neonicotinoid, NA: Not Applicable, OC: Organochlorine, Sy: Synergist