

Note

Targeting DNA Double-Strand Breaks with TAL Effector Nucleases

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ABSTRACT

Engineered nucleases that cleave specific DNA sequences *in vivo* are valuable reagents for targeted mutagenesis. Here we report a new class of sequence-specific nucleases created by fusing transcription activator-like effectors (TALEs) to the catalytic domain of the *FokI* endonuclease. Both native and custom TALE-nuclease fusions direct DNA double-strand breaks to specific, targeted sites.

ZINC finger nucleases (ZFNs) and meganucleases cleave specific DNA target sequences *in vivo* and are powerful tools for genome modification (CARROLL 2008; CATHOMEN and JOUNG 2008; GALETTO *et al.* 2009). Chromosome breaks created by these engineered nucleases stimulate homologous recombination or gene targeting: in the presence of a template for repairing the double-strand break, specific DNA sequence changes in the template become incorporated into chromosomes at or near the break site. In the absence of a repair template, broken chromosomes are rejoined by nonhomologous end joining, which often introduces short DNA insertions or deletions that create targeted gene knock-outs. For both ZFNs and meganucleases, a barrier to their widespread adoption has been the challenge in engineering new DNA binding specificities. While significant progress has been made in recent years (URNOV *et al.* 2005; FAJARDO-SANCHEZ *et al.* 2008; MAEDER *et al.* 2008; KIM *et al.* 2009), generating the necessary reagents to modify new DNA targets is still resource intensive and to some degree empirical.

A novel DNA binding domain was recently described in a family of proteins known as transcription activator-like effectors (TALEs) (BOCH *et al.* 2009; MOSCOU and BOGDANOVE 2009). TALEs are produced by plant pathogens in the genus *Xanthomonas*, which deliver the

proteins to plant cells during infection through the type III secretion pathway (BOGDANOVE *et al.* 2010). Once inside the plant cell, TALEs enter the nucleus, bind effector-specific DNA sequences, and transcriptionally activate gene expression. Typically, activation of target genes increases plant susceptibility to pathogen colonization, but in some cases, it triggers plant defense. TALE binding to DNA is mediated by a central region of these proteins that contains as many as 30 tandem repeats of a 33- to 35-amino-acid-sequence motif (Figure 1A). The amino acid sequence of each repeat is largely invariant, with the exception of two adjacent amino acids (the repeat variable diresidue or RVD). Repeats with different RVDs recognize different DNA base pairs, and there is a one-to-one correspondence between the RVDs in the repeat domain and the nucleotides in the target DNA sequence, constituting a straightforward cipher (BOCH *et al.* 2009; MOSCOU and BOGDANOVE 2009). Using this cipher, targets of new TALEs have been correctly predicted, and functional targets of TALEs composed of randomly assembled repeats have been generated (BOCH *et al.* 2009; MOSCOU and BOGDANOVE 2009; ROMER *et al.* 2010). The ability to predict the DNA binding specificity of native or artificial TALEs suggests a variety of applications for these proteins in the targeted modification of genomes. In particular, the DNA recognition domain could direct a fused nuclease to a specific DNA sequence. Further, the apparent modularity of the repeats should enable rapid construction of TALE nucleases (TALENs) with novel specificities to target double-strand breaks at specific locations in the genome.

To test this idea, we modified TALEs by adopting the molecular architecture used for ZFNs. ZFNs function as

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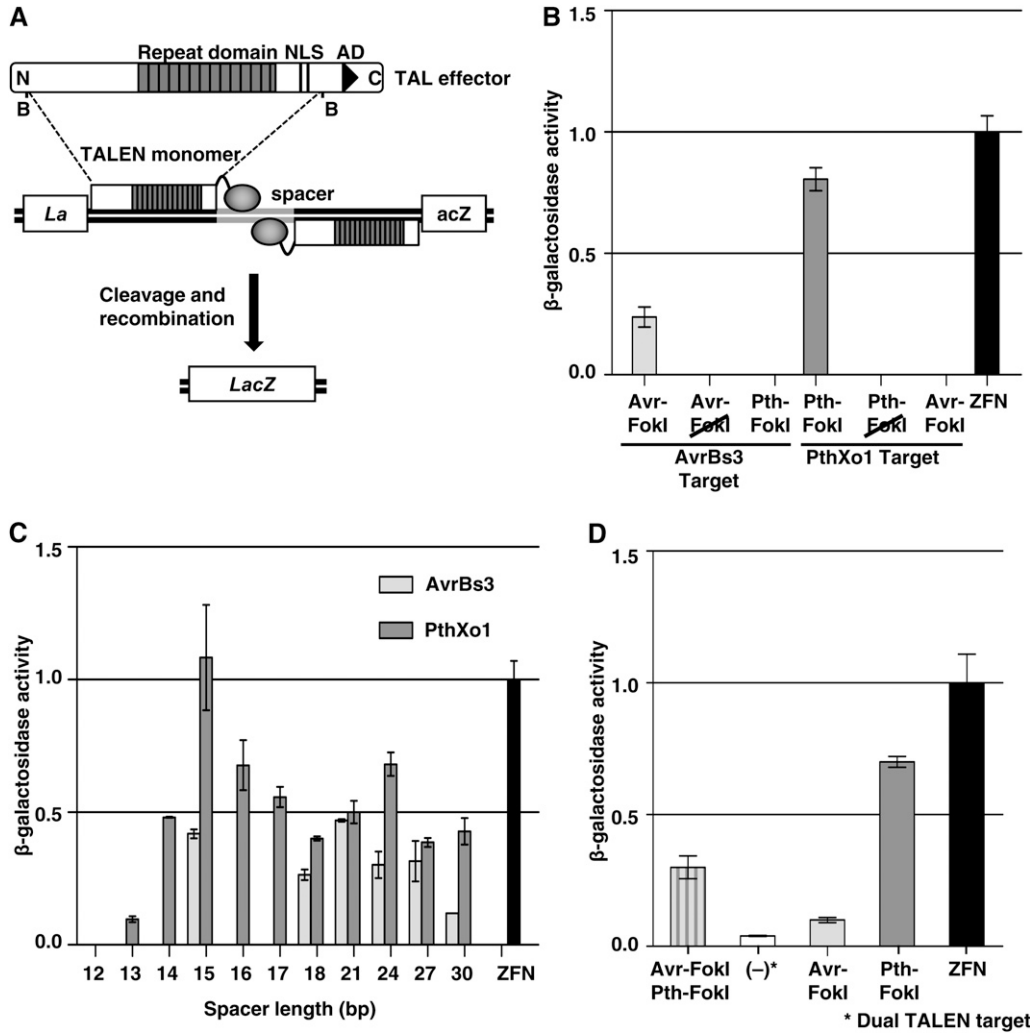


FIGURE 1.—Structure and activity of TALE nucleases (TALENs). (A) Schematic of a transcription activator-like effector (TALE) protein. *Bam*HI fragments were fused to the catalytic domain of the *Fok*I endonuclease to create TALENs. TALEN activity was measured in an *in vivo* yeast assay that is described in the text (TOWNSEND *et al.* 2009). N, N terminus; NLS, nuclear localization signal; B, *Bam*HI site; AD, acidic activation domain. (B) Activity of TALENs constructed with TALEs AvrBs3 and PthXo1. Haploid cell types containing either TALEN expression or target plasmid in 200 μ l of overnight culture were mated in YPD medium at 30°. After 4 hr, the YPD medium was replaced with 5 ml of selective medium and incubated overnight at 30°. Mated cultures were lysed, ONPG substrate added, and absorbance read at 415 nm using a 96-well plate reader (TOWNSEND *et al.* 2009). β -Galactosidase levels were calculated as a function of substrate cleavage velocity and normalized to the ZFN positive control. Avr-FokI, AvrBs3 TALEN; Pth-FokI, PthXo1 TALEN; Avr-FokI and Pth-FokI, AvrBs3 and

PthXo1 fusions to a catalytically inactive version of *Fok*I (BITINAITE *et al.* 1998); ZFN, zinc finger nuclease containing the Zif268 DNA binding domain as a positive control (PORTEUS and BALTIMORE 2003). (C) Activity of AvrBs3 and PthXo1 TALENs on targets with different spacer lengths. ZFN, Zif268-derived zinc finger nuclease. (D) Function of a heterodimeric TALEN. Activity in yeast containing PthXo1-FokI and AvrBs3-FokI expression vectors and a plasmid with a target consisting of recognition sites for each, in head-to-tail orientation separated by 15 bp is shown (Avr-FokI, Pth-FokI). Also shown for reference is activity of AvrBs3 (Avr-FokI) and PthXo1 (Pth-FokI) TALENs individually and Zif268 (ZFN) on their respective targets. As a negative control, a yeast culture with only the target site plasmid for Avr-FokI, Pth-FokI was assayed for LacZ activity [denoted as (-)].

dimers, with each monomer comprising a DNA binding domain (a zinc finger array) fused to the catalytic domain of the *Fok*I restriction enzyme (CARROLL 2008; CATHOMEN and JOUNG 2008). Two zinc finger arrays are engineered to recognize target sequences in the genome (each typically 9–12 bp) that are separated by a spacer sequence (typically 5–7 bp). Binding of the zinc finger arrays to the target allows *Fok*I to dimerize and create a double-strand break within the spacer. We reasoned that the zinc finger arrays could be substituted with the DNA recognition domain of TALEs to create TALENs that recognize and cleave DNA targets (Figure 1A). To assess TALEN function, we adapted a yeast assay in which LacZ activity serves as an indicator of DNA cleavage (TOWNSEND *et al.* 2009). In this assay, a target plasmid and a TALEN expression plasmid are brought

together in the same cell by mating. The target plasmid has a *lacZ* reporter gene with a 125-bp duplication of coding sequence. The duplication flanks the TALEN target site. When a double-strand DNA break occurs at the target site, it is repaired through single-strand annealing between the duplicated sequences, creating a functional *lacZ* gene whose expression can be measured by standard assays.

We first used two well-characterized TALEs, namely AvrBs3 from the pepper pathogen *Xanthomonas campestris* pv. *vesicatoria* and PthXo1 from the rice pathogen *X. oryzae* pv. *oryzae* (BONAS *et al.* 1989; YANG *et al.* 2006). Both TALE-encoding genes have a *Bam*HI restriction fragment that encompasses the coding sequence for the repeat domain and 287 aa prior and 231 aa after (Figure 1A). Absent from the *Bam*HI fragment is the TALE

A	Custom TALEN	1	2	3	4	5	6	7	8	9	10	11	12	13
	<i>ADH1-360-12</i>	NI A	NG T	HD C	NI A	NI A	NN G	NI A	NG T	NG T	HD C	NG T	HD C	
	<i>ADH1-408-12r</i>	HD C	HD C	HD C	NI A	NN G	NI A	NI A	NN G	NG T	NI A	NI A	NI A	
	<i>ADH1-928-12</i>	HD C	HD C	NN G	NN G	NI A	NG T	NN G	HD C	NG T	HD C	HD C	NG T	
	<i>ADH1-975-12r</i>	NI A	NN G	NI A	HD C	NI A	NI A	NI A	HD C	HD C	NI A	HD C	NI A	
	<i>gridlock-2356-13r</i>	NI A	HD C	HD C	HD C	HD C	NG T	HD C	NG T	HD C	HD C	NN G	HD C	NG T

* NI = A HD = C NN = G NG = T

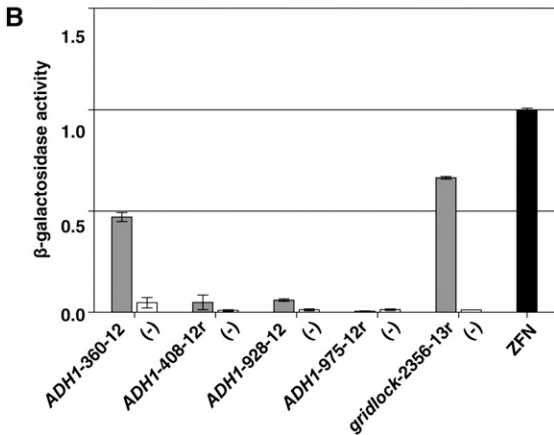


FIGURE 2.—Custom TALENs. To construct custom TALENs, repeats with the RVDs NI, HD, NN, and NG, specifying A, C, G, and T, respectively, were synthesized individually and assembled into modules of one, two, or three repeats. These modules were ligated sequentially into a derivative of the *tal1c* gene (MOSCOU and BOGDANOVA 2009) from which the original repeats had been removed. Five custom TALENs targeted to *ADH1* from Arabidopsis and the zebrafish *gridlock* gene were created. (A) The RVD sequences of individual custom TALENs and their respective DNA recognition sequences are shown. (B) The yeast *in vivo* assay was used to determine the activity of custom TALENs. Plasmids containing two recognition sites for the respective TALEN in head-to-tail orientation separated by 16- to 18-bp spacers were used as targets. We chose spacer lengths on the basis of the distance closest to 15 bp from the 3' end of the next neighboring (and opposing) candidate site, since we anticipated constructing heterodimeric TALENs that recognize native chromosomal sequences. Sixteen-base pair spacers were used for *ADH1-360-12*, *ADH1-408-12r* and 18-bp spacers for *ADH1-928-12*, *ADH1-975-12r*, and *gridlock-2356-13r*. The yeast assay was performed as described in Fig. 1B. (-), negative control with target site plasmids only; ZFN, zinc finger nuclease positive control.

transcriptional activation domain, which was replaced with a *FokI* nuclease monomer. Because the *FokI* monomers must dimerize to cleave, it was unclear what an appropriate spacer length between the two DNA recognition sites might be. For ZFNs, in which the zinc finger array is separated from *FokI* by a 4- to 7-amino-acid linker, the typical spacer between the two recognition sites is 5–7 bp. Since 235 amino acids separate the repeat domain from *FokI* in our TALEN constructs, we selected a 15-bp spacer to separate the two recognition sites, assuming that a larger spacer may be required to enable *FokI* to dimerize. As a positive control we used a well-characterized zinc finger nuclease with a DNA binding domain derived from the mouse transcription factor Zif268 (PORTEUS and BALTIMORE 2003). As negative controls, the TALE domains were fused to a catalytically inactive *FokI* variant or tested against noncognate DNA targets. Robust activity was observed for both the AvrBs3 and the PthXo1 TALENs (Figure 1B). The activity of the PthXo1 TALEN approximated that of the ZFN positive control.

We next varied the distance between the TALE binding sites (11 length variants between 12 and 30 bp) to identify spacer lengths that enable *FokI* to dimerize most efficiently (Figure 1C). Both enzymes showed two spacer length optima—one at 15 bp and the other at either 21 bp (AvrBs3) or 24 bp (PthXo1). For PthXo1, activity was observed for all tested spacer lengths 13 bp

and longer. Some spacer lengths for AvrBs3 showed no activity, however, suggesting that spacer length is critical for certain TALENs.

The above experiments tested activity of homodimeric TALENs, which bind two identical recognition sequences placed in opposition on either side of the spacer. Since such palindromic sites are unlikely to occur naturally in genomic targets, we tested whether TALENs could function as heterodimers. AvrBs3 and PthXo1 recognition sites were placed on either side of a 15-bp spacer (Figure 1D). The resulting activity of the heterodimeric TALEN approximated an average of the activities observed with the two homodimeric enzymes.

To realize the potential of TALENs for genome modification, it will be necessary to engineer them to recognize novel chromosomal DNA sequences. Randomly assembled repeat domains have been shown to bind DNA targets predicted by the cipher (BOCH *et al.* 2009); however, custom TALEs designed to recognize new target sequences have not been reported. This is an important distinction, because the former experiments validate the cipher, whereas the latter would demonstrate that DNA binding domains can be engineered with the requisite specificity for *in vivo* manipulations. To test whether repeat domains can be assembled to target TALENs to arbitrary chromosomal sequences, we chose two genes previously targeted for mutagenesis

with ZFNs—*ADHI* from Arabidopsis and *gridlock* from zebrafish (FOLEY *et al.* 2009; ZHANG *et al.* 2010). We first searched for 12- to 13-bp sequences in the coding regions, preceded by a 5' T and with a nucleotide composition similar to that of TALE binding sites identified by MOSCOU and BOGDANOVA (2009). In *ADHI* and *gridlock*, such sites occurred on average every 7–9 bp. Four 12-bp sites were selected in *ADHI* (at positions 360, 408, 928, and 975 of the chromosomal gene sequence) and one 13-bp site in *gridlock* (at position 2356 of the chromosomal gene sequence) (Figure 2A). We then constructed TALE repeat domains to recognize these targets, using the most abundant RVDs from native TALEs (NI for A, HD for C, NN for G, and NG for T). Sequences encoding the repeat domain of the native TALE *tal1c* were replaced with the custom repeat domains, and *Bam*HI fragments from these engineered TALEs were then fused to sequences encoding the catalytic domain of *FokI* (see supporting information, File S1). The resulting custom TALENs were tested in the yeast assay as homodimeric TALENs; that is, the identical DNA binding site was duplicated in inverse orientation on either side of a spacer. It is important to note that heterodimeric TALENs would need to be constructed to direct cleavage at naturally occurring DNA targets. Robust nuclease activity was observed for the *ADHI*-360-12 and *gridlock*-2356-13r TALEN (Figure 2B). The *ADHI*-928-12 TALEN had modest activity that was nonetheless significantly above the negative controls. For each TALEN that gave positive results, nuclease activity was specific to the cognate target. These results indicate that novel, functional TALENs can be created by assembly of customized repeat domains.

The experiments described here suggest that TALENs hold much promise for applications requiring sequence-specific nucleases. In particular, the ability to create TALENs that recognize novel, arbitrarily selected target sequences suggests a potential for engineering TALENs for targeted genome modification. That said, the failure of some custom TALENs suggests that yet unknown rules govern the assembly of functional repeat domains. For example, repeat composition may influence protein stability, or interactions among repeat domains may affect DNA binding activity as has been observed for finger–finger interactions in zinc finger arrays (ELROD-ERICKSON *et al.* 1996; ISALAN *et al.* 1997). Alternatively, the spacer lengths we used may have prevented dimerization of *FokI*, as appeared to be the case for some spacers with AvrBs3. Clearly, it will be important to gain a better understanding of the relationship of spacer length to function for TALENs with different repeat domains. Ascertaining the minimal DNA binding domain might help accomplish this; however, we believe the repeats alone are not sufficient for adequate DNA binding, as TALENs constructed with just the repeat domain did not function in the yeast

assay (data not shown). In the short term, we will test whether custom TALENs can be created that recognize and cleave endogenous chromosomal targets, and we will evaluate the efficiency with which custom TALENs create genome modifications by nonhomologous end joining and homologous recombination. Such experiments will be key to assessing the full utility of these reagents for eukaryotic genome engineering.

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LITERATURE CITED

- BITINAITE, J., D. A. WAH, A. K. AGGARWAL and I. SCHILDKRAUT, 1998 FokI dimerization is required for DNA cleavage. *Proc. Natl. Acad. Sci. USA* **95**: 10570–10575.
- BOCH, J., H. SCHOLZE, S. SCHORNACK, A. LANDGRAF, S. HAHN *et al.*, 2009 Breaking the code of DNA binding specificity of TAL-type III effectors. *Science* **326**: 1509–1512.
- BOGDANOVA, A. J., S. SCHORNACK and T. LAHAYE, 2010 TAL effectors: finding plant genes for disease and defense. *Curr. Opin. Plant Biol.* **13**: 394–401.
- BONAS, U., R. E. STALL and B. STASKAWICZ, 1989 Genetic and structural characterization of the avirulence gene *avrBs3* from *Xanthomonas campestris* pv. *vesicatoria*. *Mol. Gen. Genet.* **218**: 127–136.
- CARROLL, D., 2008 Progress and prospects: zinc-finger nucleases as gene therapy agents. *Gene Ther.* **15**: 1463–1468.
- CATHOMEN, T., and J. K. JOUNG, 2008 Zinc-finger nucleases: the next generation emerges. *Mol. Ther.* **16**: 1200–1207.
- ELROD-ERICKSON, M., M. A. ROULD, L. NEKLUDOVA and C. O. PABO, 1996 Zif268 protein-DNA complex refined at 1.6 Å: a model system for understanding zinc finger-DNA interactions. *Structure* **4**: 1171–1180.
- FAJARDO-SANCHEZ, E., F. STRICHER, F. PAQUES, M. ISALAN and L. SERRANO, 2008 Computer design of obligate heterodimer meganucleases allows efficient cutting of custom DNA sequences. *Nucleic Acids Res.* **36**: 2163–2173.
- FOLEY, J. E., J. R. YEH, M. L. MAEDER, D. REYON, J. D. SANDER *et al.*, 2009 Rapid mutation of endogenous zebrafish genes using zinc finger nucleases made by Oligomerized Pool ENgineering (OPEN). *PLoS One* **4**: e4348.
- GALETTO, R., P. DUCHATEAU and F. PAQUES, 2009 Targeted approaches for gene therapy and the emergence of engineered meganucleases. *Expert Opin. Biol. Ther.* **9**: 1289–1303.
- ISALAN, M., Y. CHOO and A. KLUG, 1997 Synergy between adjacent zinc fingers in sequence-specific DNA recognition. *Proc. Natl. Acad. Sci. USA* **94**: 5617–5621.
- KIM, H. J., H. J. LEE, H. KIM, S. W. CHO and J. S. KIM, 2009 Targeted genome editing in human cells with zinc finger nucleases constructed via modular assembly. *Genome Res.* **19**: 1279–1288.
- MAEDER, M. L., S. THIBODEAU-BEGANNY, A. OSIACK, D. A. WRIGHT, R. M. ANTHONY *et al.*, 2008 Rapid “open-source” engineering of customized zinc-finger nucleases for highly efficient gene modification. *Mol. Cell* **31**: 294–301.
- MOSCOU, M. J., and A. J. BOGDANOVA, 2009 A simple cipher governs DNA recognition by TAL effectors. *Science* **326**: 1501.
- PORTEUS, M. H., and D. BALTIMORE, 2003 Chimeric nucleases stimulate gene targeting in human cells. *Science* **300**: 763.
- ROMER, P., S. RECHT, T. STRAUSS, J. ELSAESSER, S. SCHORNACK *et al.*, 2010 Promoter elements of rice susceptibility genes are bound and activated by specific TAL effectors from the bacterial blight pathogen, *Xanthomonas oryzae* pv. *oryzae*. *New Phytol.* (in press).
- TOWNSEND, J. A., D. A. WRIGHT, R. J. WINFREY, F. FU, M. L. MAEDER *et al.*, 2009 High-frequency modification of plant genes using engineered zinc-finger nucleases. *Nature* **459**: 442–445.

- URNOV, F. D., J. C. MILLER, Y. L. LEE, C. M. BEAUSEJOUR, J. M. ROCK *et al.*, 2005 Highly efficient endogenous human gene correction using designed zinc-finger nucleases. *Nature* **435**: 646–651.
- YANG, B., A. SUGIO and F. F. WHITE, 2006 Os8N3 is a host disease-susceptibility gene for bacterial blight of rice. *Proc. Natl. Acad. Sci. USA* **103**: 10503–10508.
- ZHANG, F., M. L. MAEDER, E. UNGER-WALLACE, J. P. HOSHAW, D. REYON *et al.*, 2010 High frequency targeted mutagenesis in *Arabidopsis thaliana* using zinc finger nucleases. *Proc. Natl. Acad. Sci. USA* **107**: 12028–12033.

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Supporting Information

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