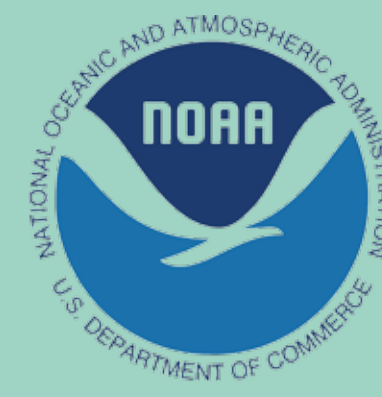


Effect of light exposure and nutrients on buoyancy of

Microcystis colonies



Tonghui Ming^{*1}, Henry A. Vanderploeg², Mark D. Rowe³, David L. Fanslow², J. Rudi Strickler⁴, Russell Miller³, Thomas H. Johengen³, Timothy W. Davis², Duane Gossiaux².

^{*}Presenter, ¹School of Natural Resources & Environment, University of Michigan; ²NOAA Great Lakes Environmental Research Laboratory; ³Cooperative Institute for Limnology and Ecosystems Research, University of Michigan; ⁴Dept. BioSci, University of Wisconsin-Milwaukee.
Contact Information: thming@umich.edu

Introduction

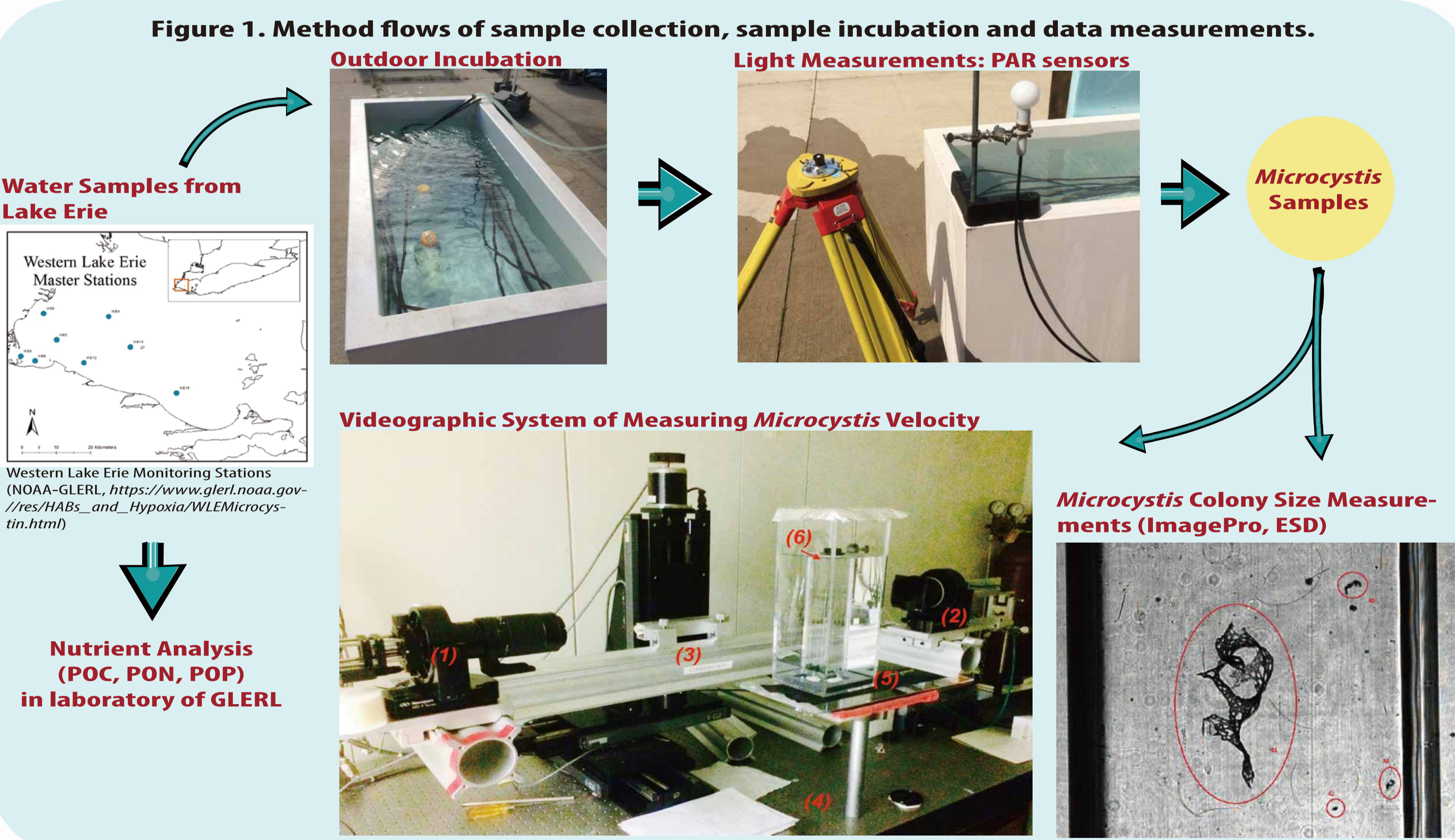
Understanding the vertical distribution of *Microcystis* is important for improving satellite-derived estimates of bloom biomass and for predicting the transport of blooms. Initialized from satellite imagery, the HAB Tracker forecast model predicts the transport and vertical distribution of harmful algal blooms (HABs) in Lake Erie over a five-day period. By focusing on buoyant velocity, we can learn more about the driven factors of *Microcystis* vertical distributions, and use the results to further improve current HAB Tracker forecast Model.

Objectives

In this study, we used an improved videographic system to **measure *Microcystis* velocity** and then developed statistical **relationships between *Microcystis* velocity, colony size, light exposure and nutrients**.

Methods

1. Experimental Setup and *Microcystis* Velocity & Size Measurements

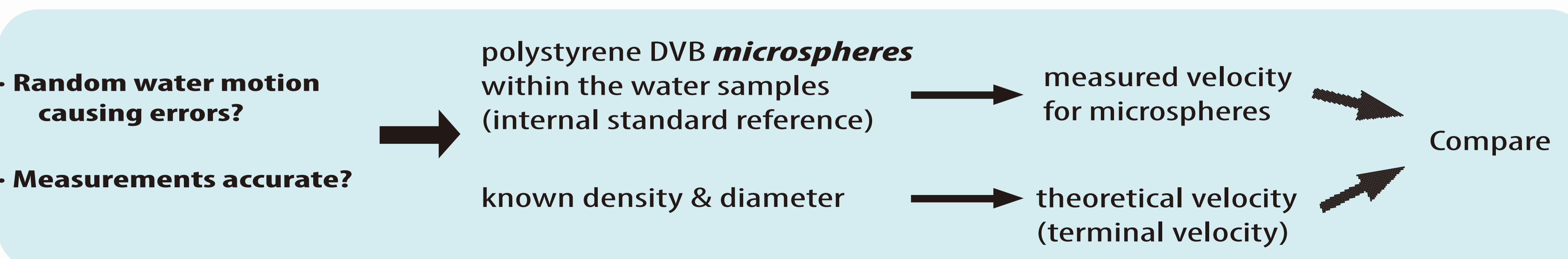


Videographic recording system: a camera system and a software system.

- Camera System:**
- temperature-controlled room
 - stationary table, 3D positioning frame
 - Shadowgraph** optics system
 - NIR laser (710 nm), Basler digital video camera (mNIR)
 - quartz prism vessel containing *Microcystis* samples
 - larger, outer container - the thermal water jacket
- Software System:**
- Contemplas Templo - collecting videos of *Microcystis* movements
 - Contemplas Vicon Motus - measuring the buoyant velocities

Colony Size - equivalent spherical diameter (ESD): calculated by converting the projected area of a *Microcystis* colony image to a diameter of a circle of equivalent area.

2. Method Verification



Results & Discussions

1. Method Verification

The mean velocities of two microspheres sizes (49.5 μm and 102 μm) on each sampling day (Figure 2) verified that the *Microcystis* velocity measurements were accurate.

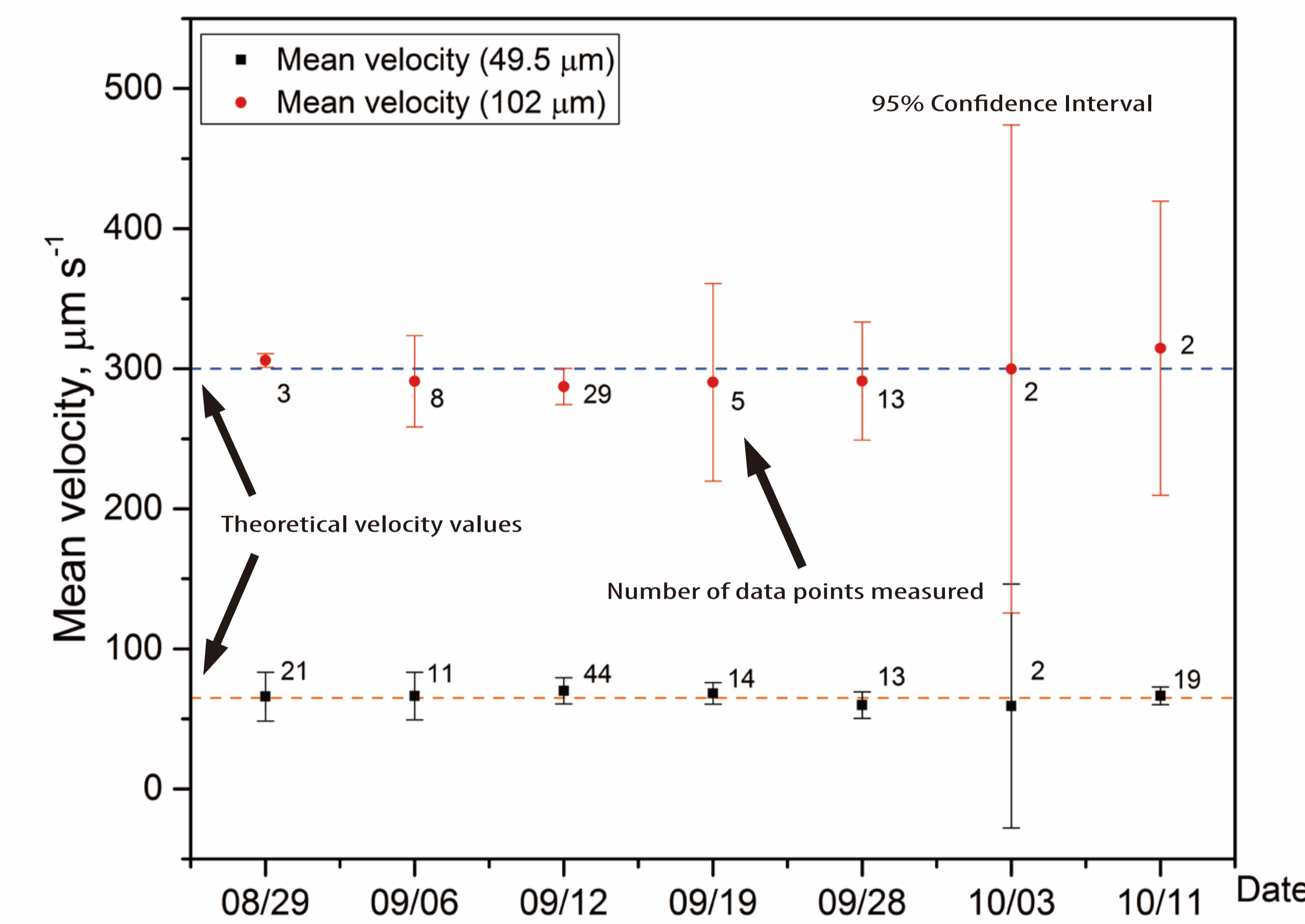


Figure 2. Mean velocity of microspheres for each size on each sampling day throughout the bloom season.

2. Velocity & Colony Size

Microcystis velocity and colony size were significantly and positively correlated, indicated by the slopes that were significantly different from zero on a log-log plot (Figure 3).

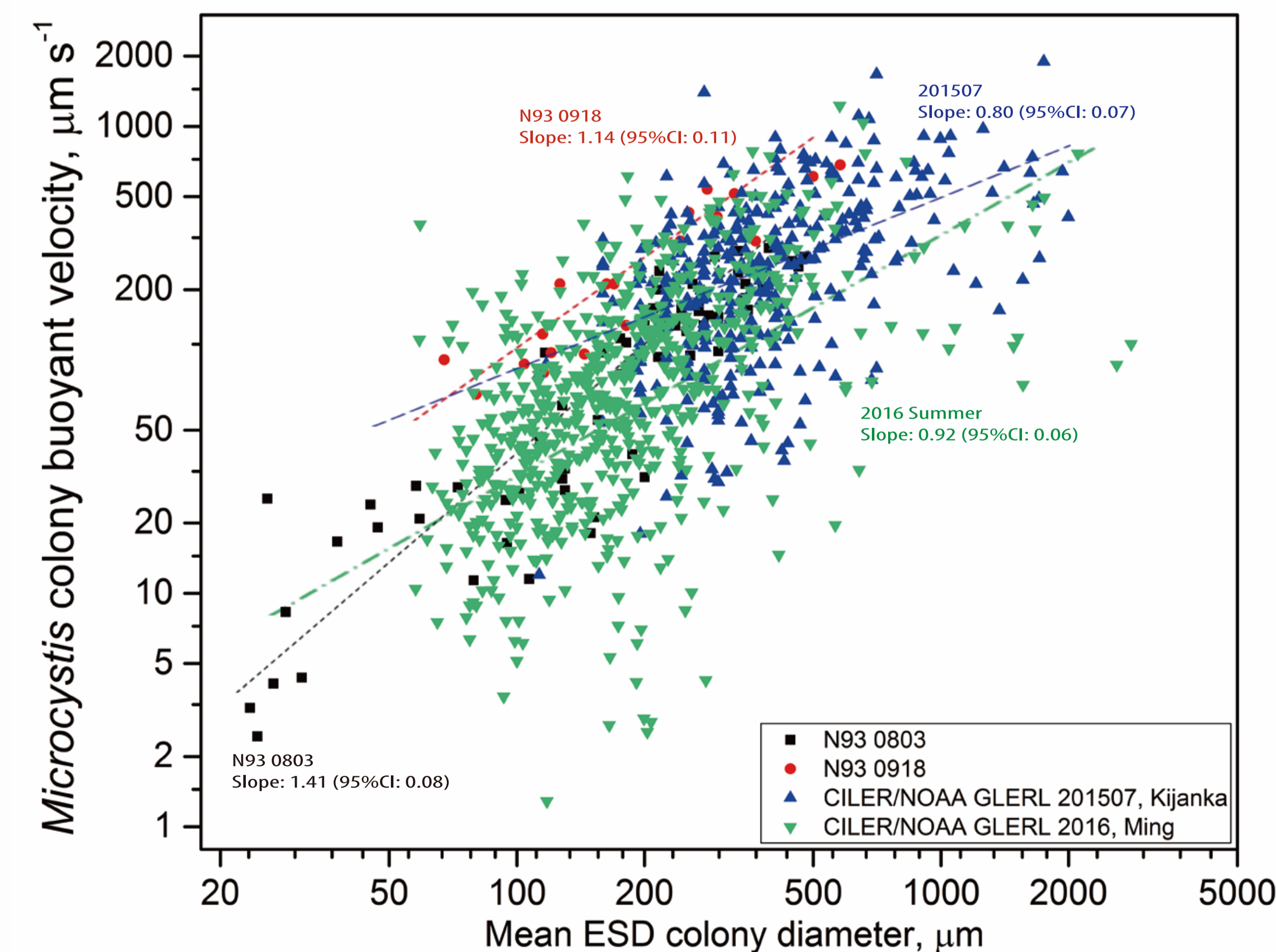


Figure 3. *Microcystis* buoyant velocity vs. colony size for data comparison. Only floating colonies are shown in this figure.

3. Velocity & Light Exposure

Mean velocity of *Microcystis* and light exposure (PAR) were significantly and negatively correlated, indicated by negative slopes in Figure 4.

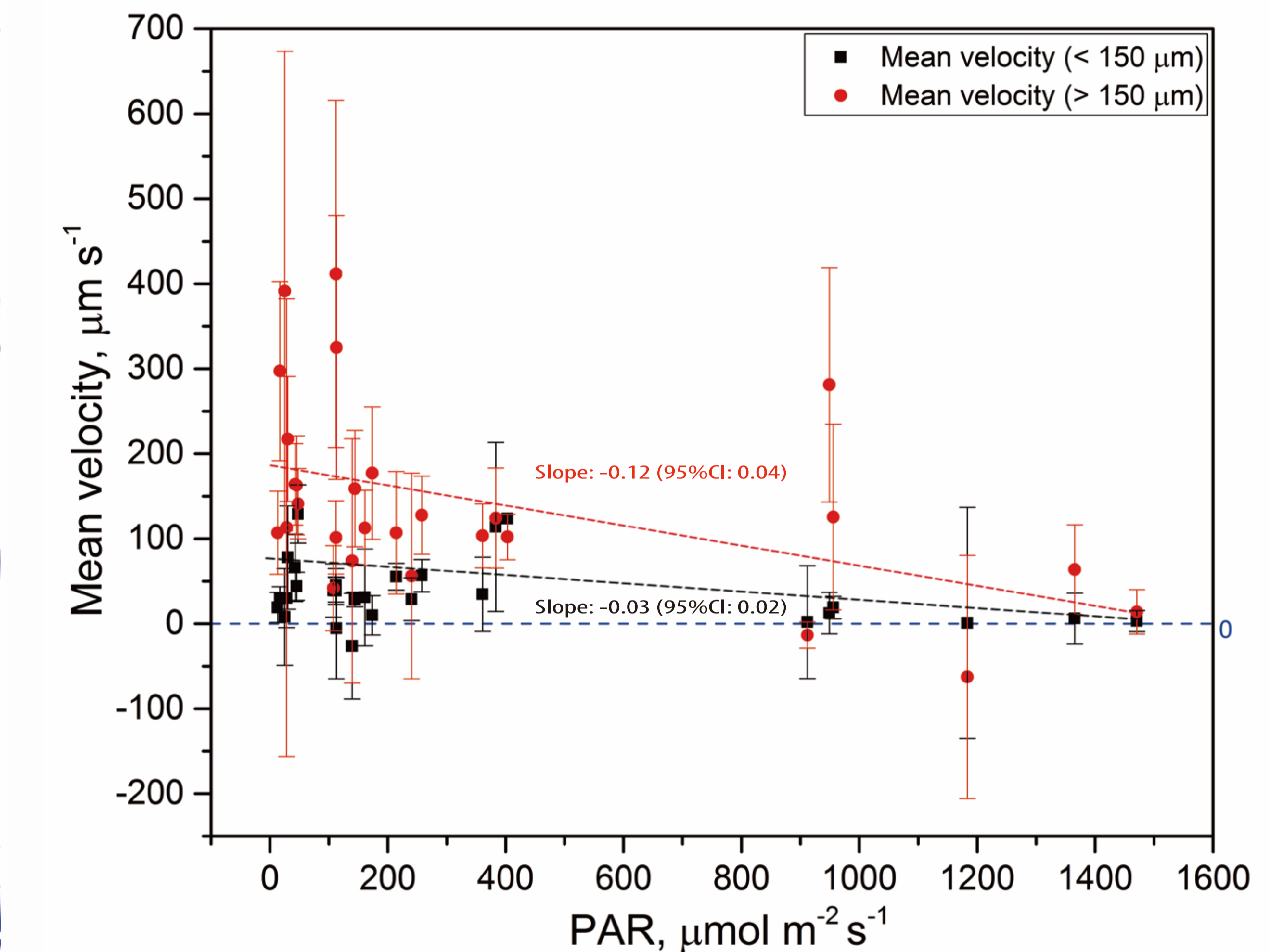


Figure 4. Mean velocity versus PAR for two colony size classes.

4. Velocity & Nutrients

Nutrient analysis showed no deficiency of nitrogen (N) or phosphorous (P) throughout the bloom season in Lake Erie, and we found no significant relationship between buoyant velocity and nutrients if the nutrients are abundant.

Conclusions

Our study has found that **strong/long light exposure can make *Microcystis* less buoyant**, while **large colony size contributes to more buoyant colonies**. As for the nutrient effects, we did not observe nutrient deficiency during the bloom season, or the relationship between N: P molar ratios and velocity under nutrient abundance conditions.

References:

- Nakamura, T., Y. Adachi, and M. Suzuki. 1993. Flotation and sedimentation of a single *Microcystis* floc collected from surface bloom. *Wat. Res.* 27 (6):979-983.
- Medrano, A. E., R. E. Uittenbogaard, L. M. Dionisio Pires, B. J. H. van de Wiel, and H. J. H. Clercx. 2013. Coupling hydrodynamics and buoyancy regulation in *Microcystis aeruginosa* for its vertical distribution in lakes. *Ecological Modelling* 248:41-56.
- Ibelings, B. W., L. R. Mur, and A. E. Walsby. 1991. Diurnal changes in buoyancy and vertical distribution in populations of *Microcystis* in two shallow lakes.
- Rowe, M. D., E. J. Anderson, T. T. Wynne, R. P. Stumpf, D. L. Fanslow, K. Kijanka, H. A. Vanderploeg, J. R. Strickler, and T. W. Davis. 2016. Vertical distribution of buoyant *Microcystis* blooms in a Lagrangian particle tracking model for short-term forecasts in Lake Erie. *Journal of Geophysical Research: Oceans* 121:1-19.
- Vanderploeg, H. A., Sarnelle, O., Liebig, J. R., Morehead, N. R., Robinson, S. D., Johengen, T. H., & Horst, G. P. 2017. Seston quality drives feeding, stoichiometry and excretion of zebra mussels. *Freshwater Biology*, 62(4), 664-680.
- Strickler, J.R. and J.S. Hwang. 1999. Matched Spatial Filters in Long Working Distance Microscopy of Phase Objects. In: Cheng, P.C., Hwang, P.P., Wu, J.L., Wang, G. and Kim H. [Eds.] *Focus on Multidimensional Microscopy*. World Scientific Publishing Pte. Ltd., River Edge, NJ. 217-239.
- Hecky, R. E., et al. (1993). The Stoichiometry of Carbon, Nitrogen, and Phosphorus in Particulate Matter of Lakes and Oceans. *Limnology and Oceanography* 38(4): 709-724.